

Duchenne and Becker Muscular Dystrophy: Clinical Features Genetics, and Diagnostics

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Outline

- The dystrophinopathies: Duchenne and Becker muscular dystrophy
- Clinical features
- Lessons from mutational analysis
 - Genotypically and phenotypically catalogued patients
 - Newborn screening
- Emerging therapeutic approaches





Duchenne de Boulogne



École nationale supérieure des beaux-arts

Paralysie musculaire pseudohypertrophique ou paralysie myosclérosique

Duchenne:

- *De l'électrisation localisée et son application à la pathologie et à la thérapeutique.* (1861)
- *Récherches sur la paralysie musculaire pseudohypertrophique, ou paralysie myosclérosique.* Arch Gén Méd. (1868)

Edward Meryon (England):

- “granular and fatty degeneration of the voluntary muscles” in eight boys (1852)

Gaetano Conte (Italy):

- affected brothers (1836)



Gowers

Duchenne Muscular Dystrophy

- Progressive skeletal muscle degeneration

- Onset age 3-5:

- Pelvic girdle weakness (difficulty arising/climbing stairs)
 - Gait abnormalities (toe walking)
 - Serum CK 50-100X normal



- Commonly cited incidence of 1:3500 live male births
- MDA estimates up to 12,000 boys with DMD registered in clinics




(a)



(b)



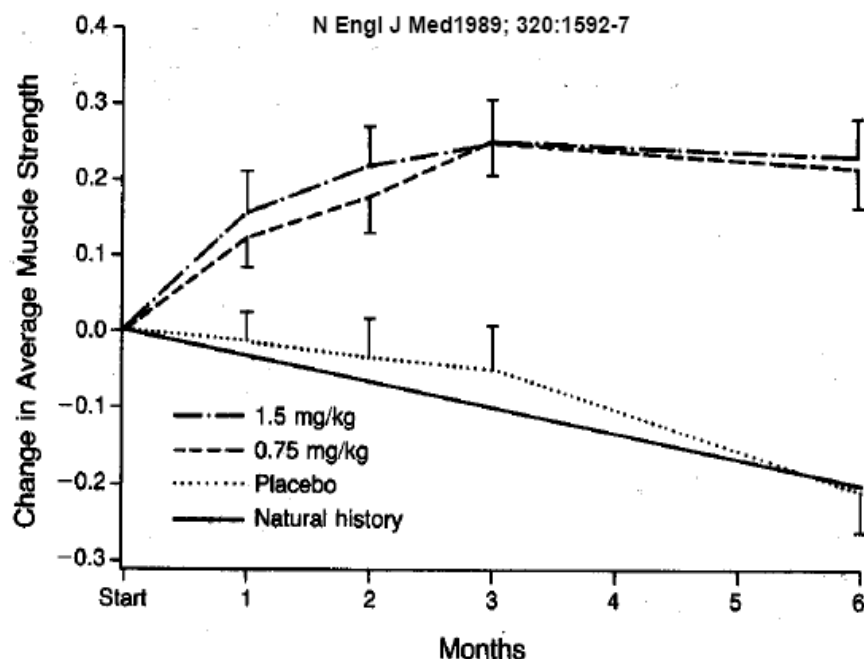
(c)

- 
- Loss of ambulation by age 12 years (range 7-12)
 - Mean age at death around 19 years
 - Dilated cardiomyopathy
 - Ventilatory insufficiency

Randomized, double-blind six-month trial of prednisone in Duchenne's muscular dystrophy

Mendell JR, Moxley RT, Griggs RC, et al

N Engl J Med. 1989 Jun 15;320(24):1592-7.




■ Improve muscle strength

- Prolongs ambulation by 2-3 years
- Prevents scoliosis

■ Limitations

- No change in functional grades
- Many cannot tolerate side effects

- 
- Loss of ambulation by age 12 years (range 7-12)
 - Mean age at death around 19 years
 - Dilated cardiomyopathy
 - Ventilatory insufficiency
 - Glucocorticoid corticosteroids
 - Prednisone 0.75 mg/kg/day
 - Deflazacort 0.9 mg/kg/day
 - AAN Practice Parameters; Cochrane review
 - Prolonged ambulation (up to 1-3 years)
 - Significant side effects
 - Supportive Care
 - Nocturnal ventilatory support
 - Spinal surgery in appropriate candidates



Dystrophinopathies: Clinical diagnosis

■ DMD:

- Onset age 3-5
- Pelvic girdle weakness
- Tight heel cords
- CK 50-100X normal
- Loss of ambulation by age 12

■ BMD:

- Classic definition: loss of ambulation > age 12
- Alternatively:
 - “intermediate muscular dystrophy” for loss of ambulation ages 12 through 15
 - BMD for loss of ambulation > age 15
- Limb-girdle syndromes in adulthood
- Myalgias
- Isolated cardiomyopathy



Dystrophin Mutations

- Dystrophin gene (Xp21.1) is huge:
 - 2.4 million base pairs
 - 79 exons and 8 promoters
- Large deletions (≥ 1 exon) account for ~65% of DMD/BMD patients
- ~5% have duplications
- ~15% of boys have nonsense mutations
- Remainder are frameshifting insertions/deletions, splice site mutations, missense mutations


Distribution of mutations in an unselected cohort

(Dent *et al*; AJMG, 2005 Apr 30;134(3):295-8)

Mutation Type	DMD	BMD	Carrier	Total
≥1 exon deletion	32	13		45 (66%)
Premature Stop	5	3	1	9 (13%)
Missense	1	2		3 (4%)
Frameshift insertion or deletion	1		1	2 (3%)
≥1 exon duplication	3	1		4 (6%)
No mutation detected	3	2		5 (7%)
Total	45	21	2	68

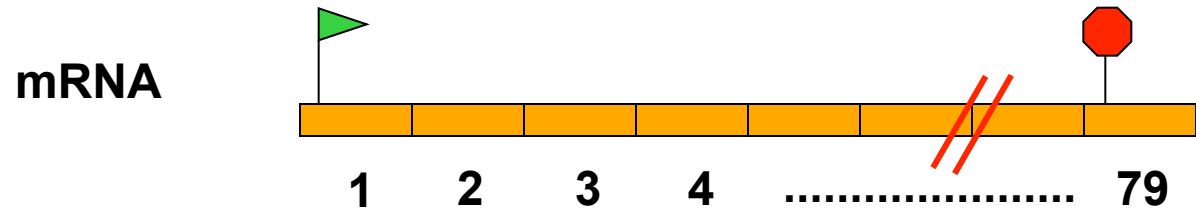
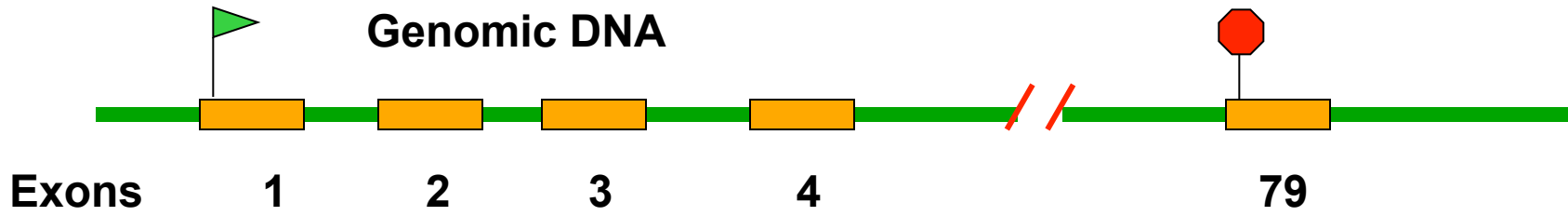
Currently available methodology can detect 93%-96% of dystrophinopathy mutations from blood samples.

(Yan *et al*, Hum Mutat 2004; 23:203-204).

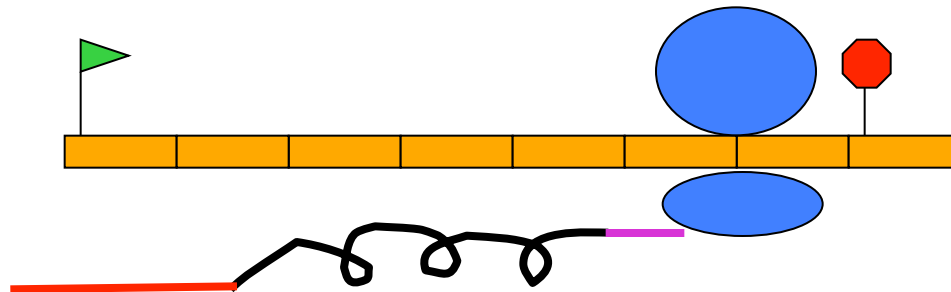


Dystrophin mutations: Duchenne vs Becker

- Size of deletion does not correlate well with phenotype
- in-frame deletions are more likely to result in translation of a protein with partial function
 - (i.e., out-of-frame deletions are DMD ~90% of the time)



Protein synthesis



Reading Frame

THE BIG RED DOG RAN AND SAT

T HEB IGR EDD OGR ANA NDS AT

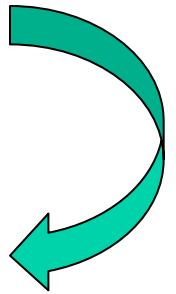
TH EBI GRE DDO GRA NAN DSA T

Reading Frame

- THE BIG RED DOG RAN AND SAT

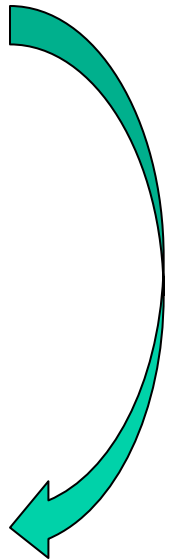
Reading Frame

- THE BIG RED DOG RAN AND SAT
- THE BIR EDD OGR ANA NDS AT



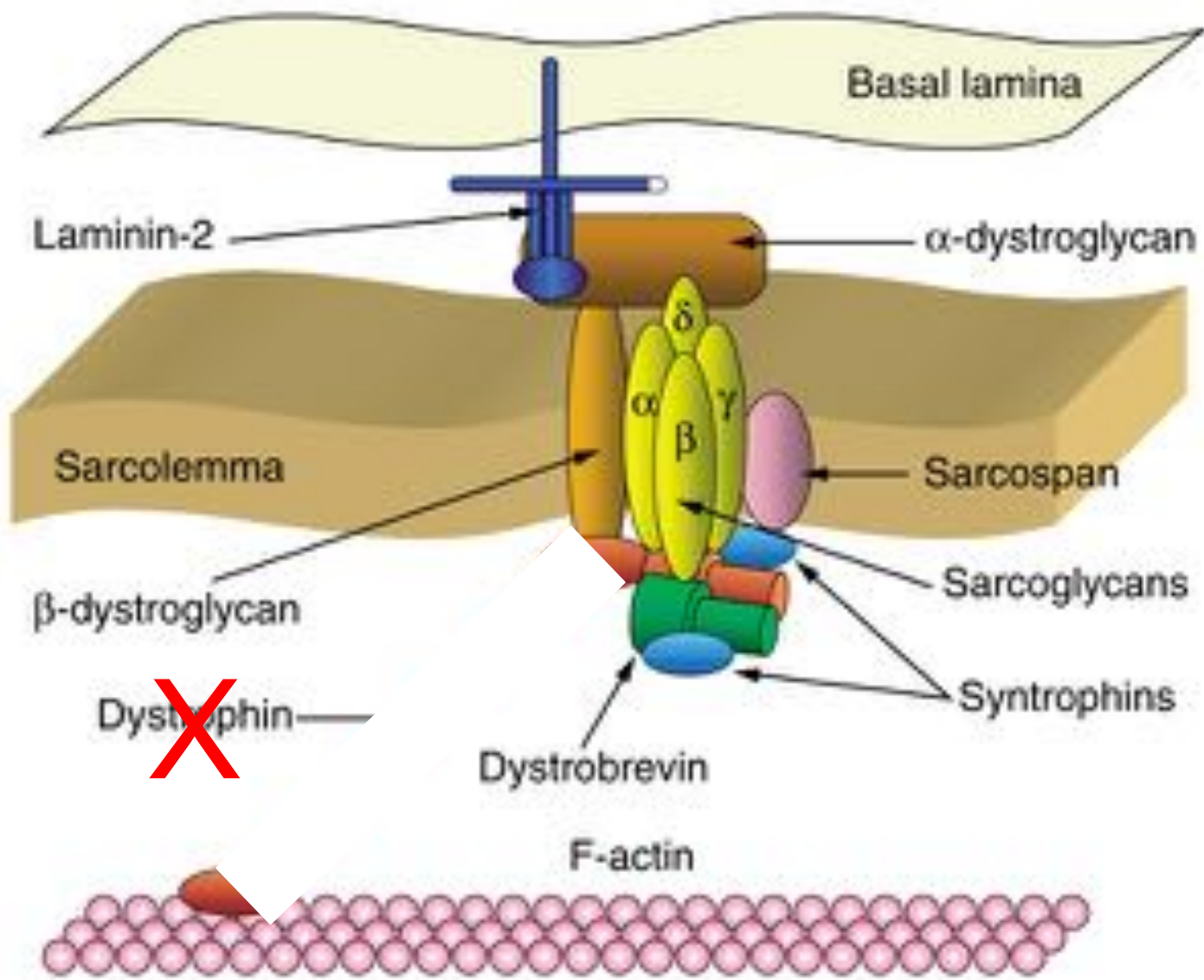
Reading Frame

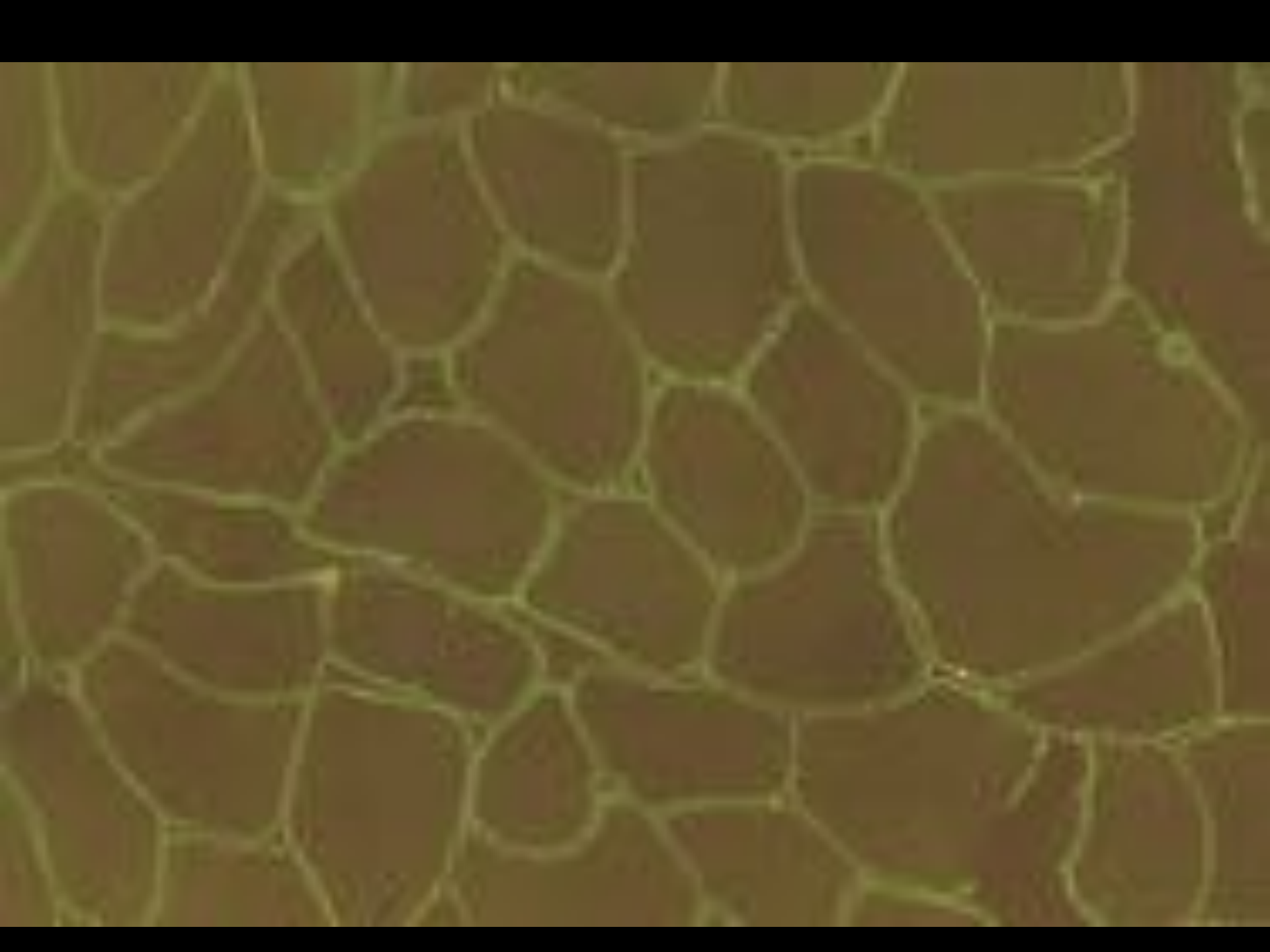
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- ~~THE BIR EDD OGR ANA NDS AT~~
- THE DOG RAN AND SAT

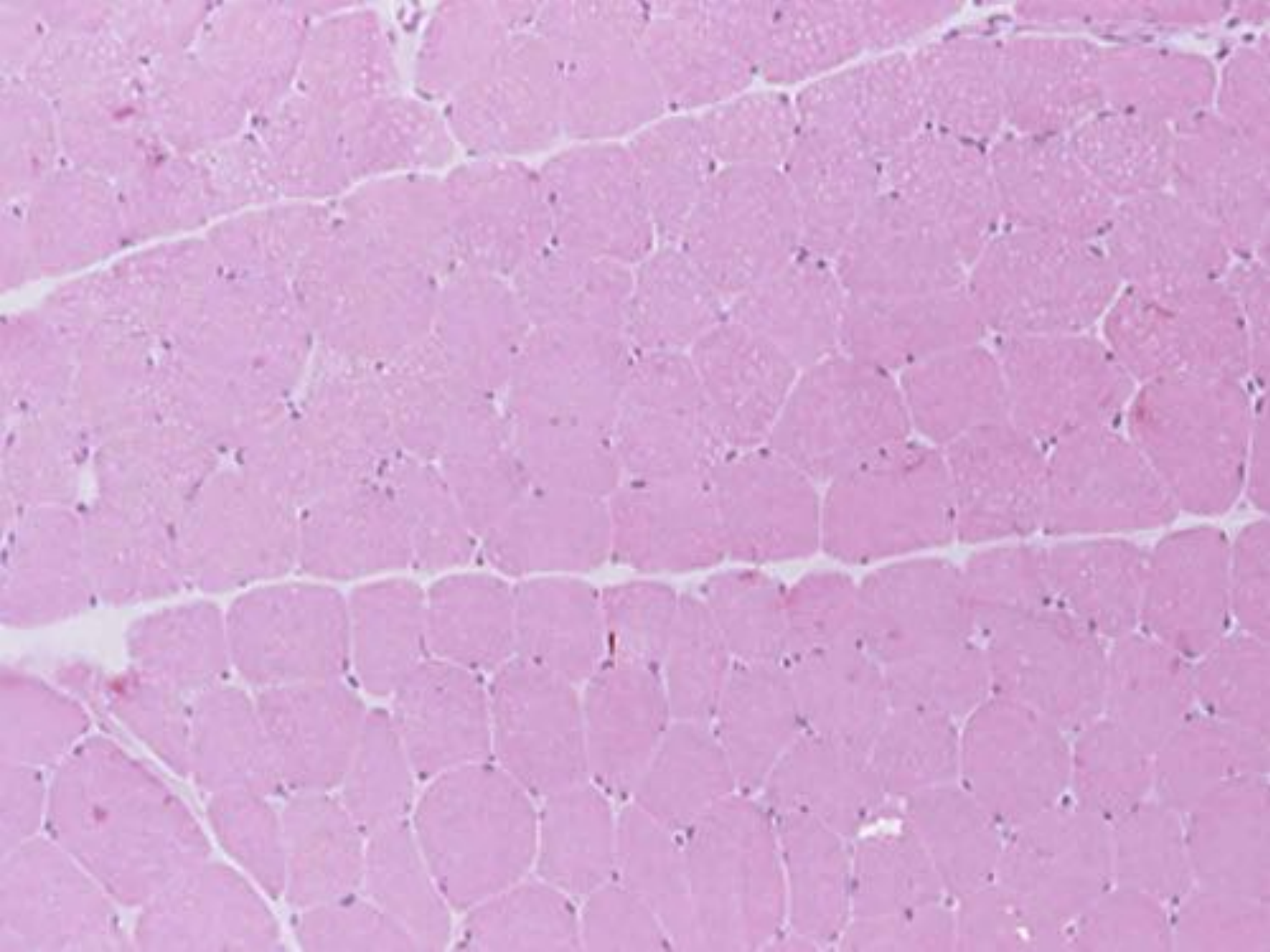


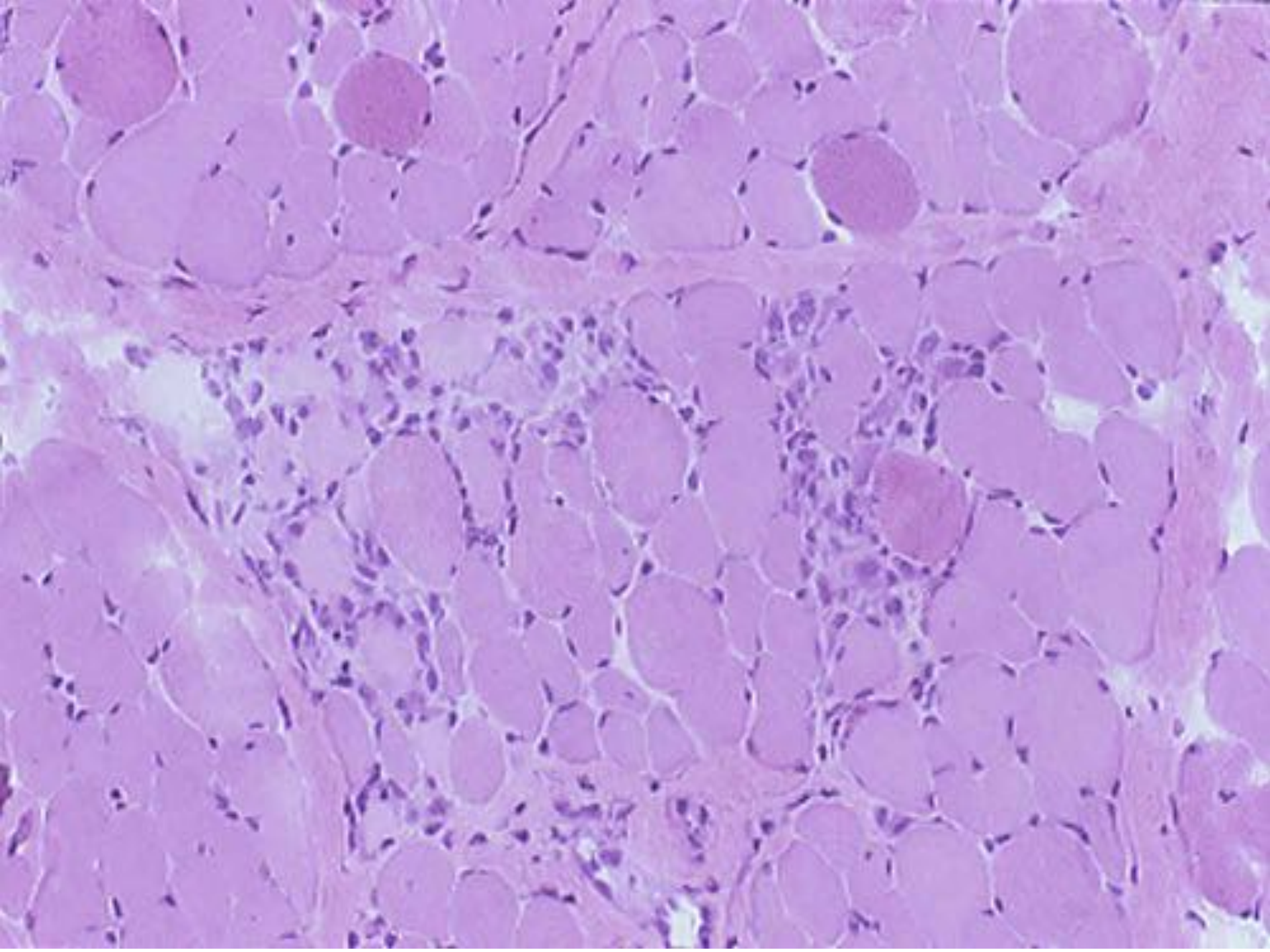
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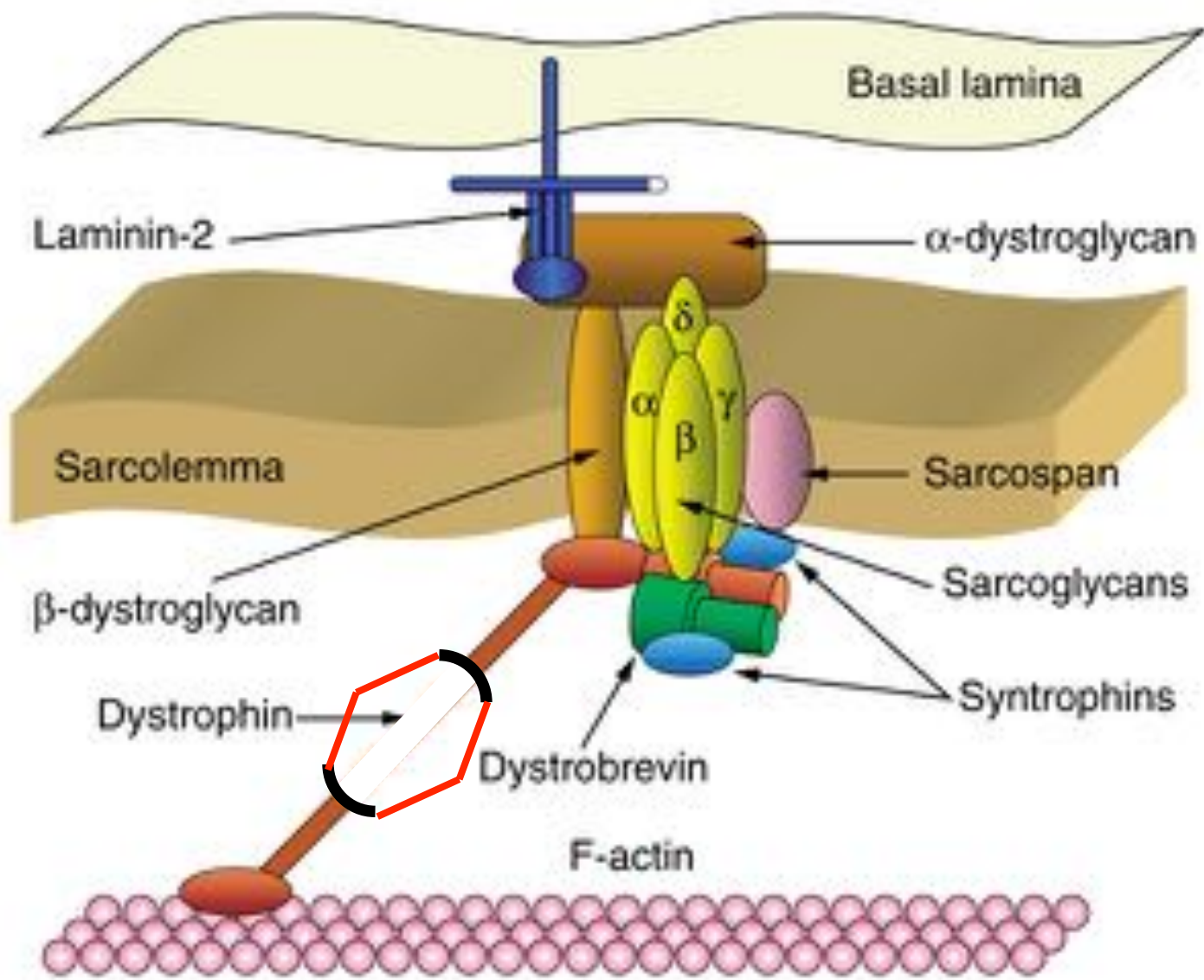
- THE BIG RED DOG RAN AND SAT
- THE BIR EDD OGR ANA NDS AT
= Duchenne Muscular Dystrophy
- THE DOG RAN AND SAT
= Becker Muscular Dystrophy







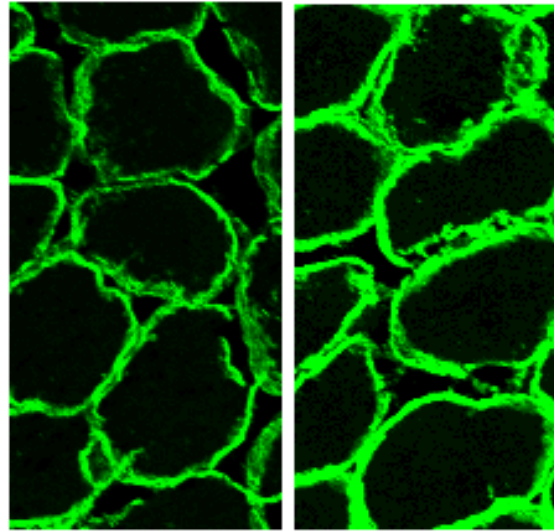




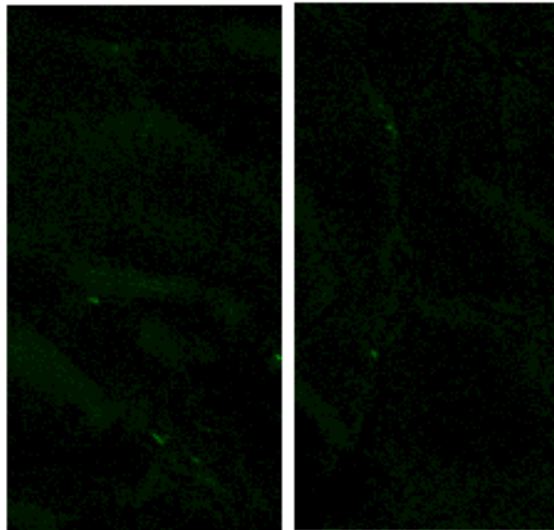
Mandys 1
(exon 31-32)

Mandys 8
(exon 32)

Normal



Pt 42719



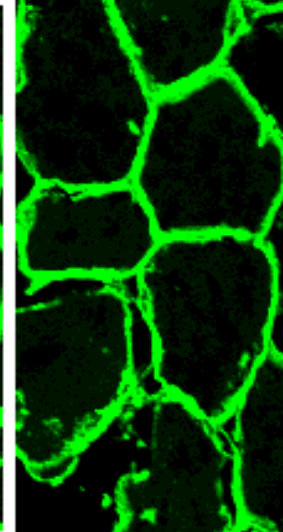
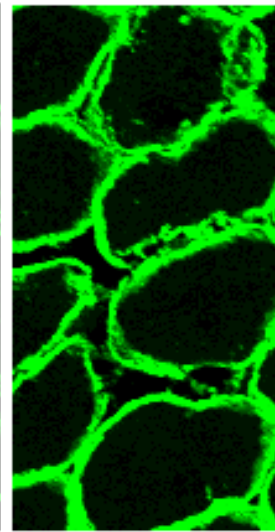
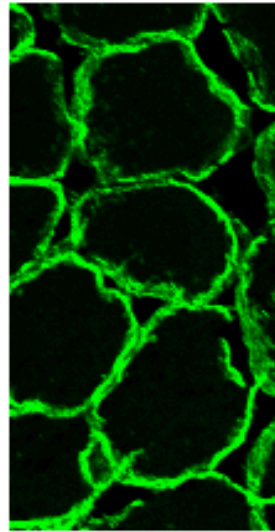
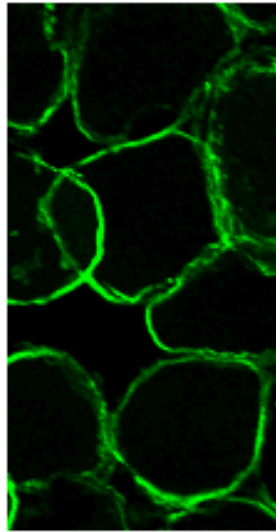
Mandys 19
(exon 20-21)

Mandys 1
(exon 31-32)

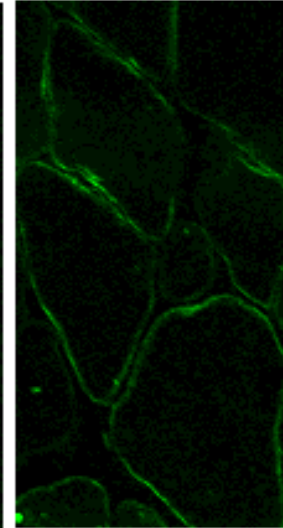
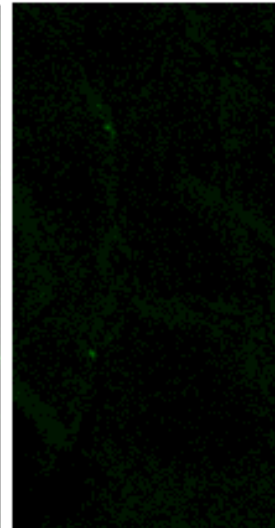
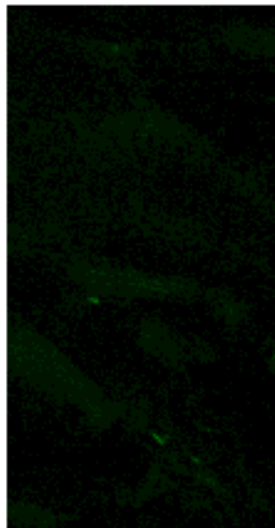
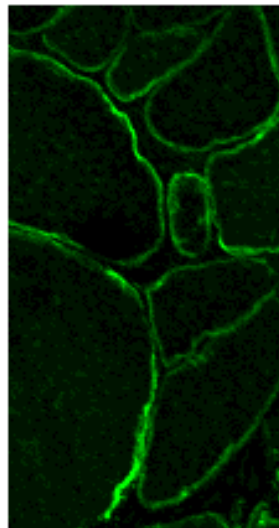
Mandys 8
(exon 32)

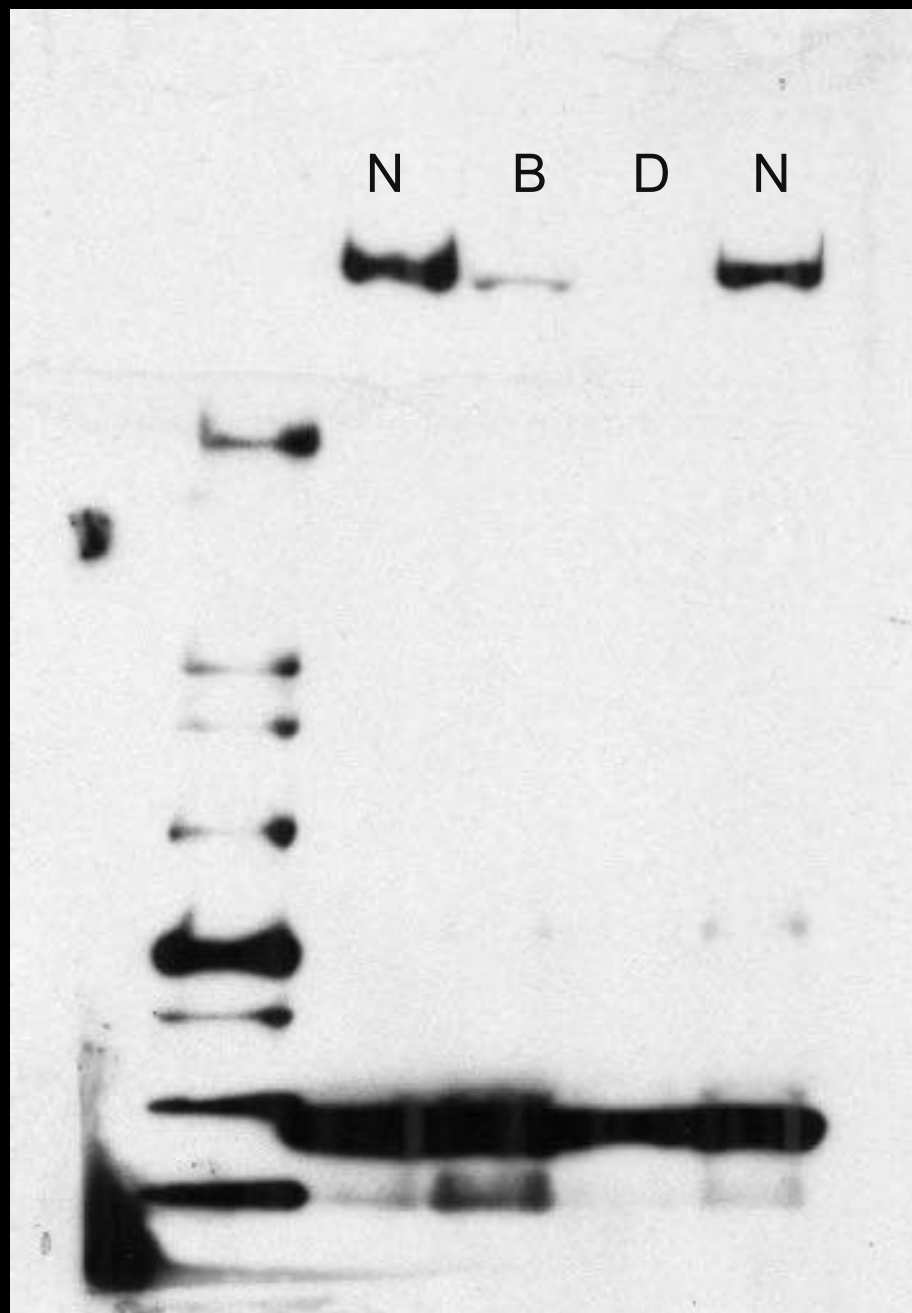
Dys 2
(exon 77-79)

Normal



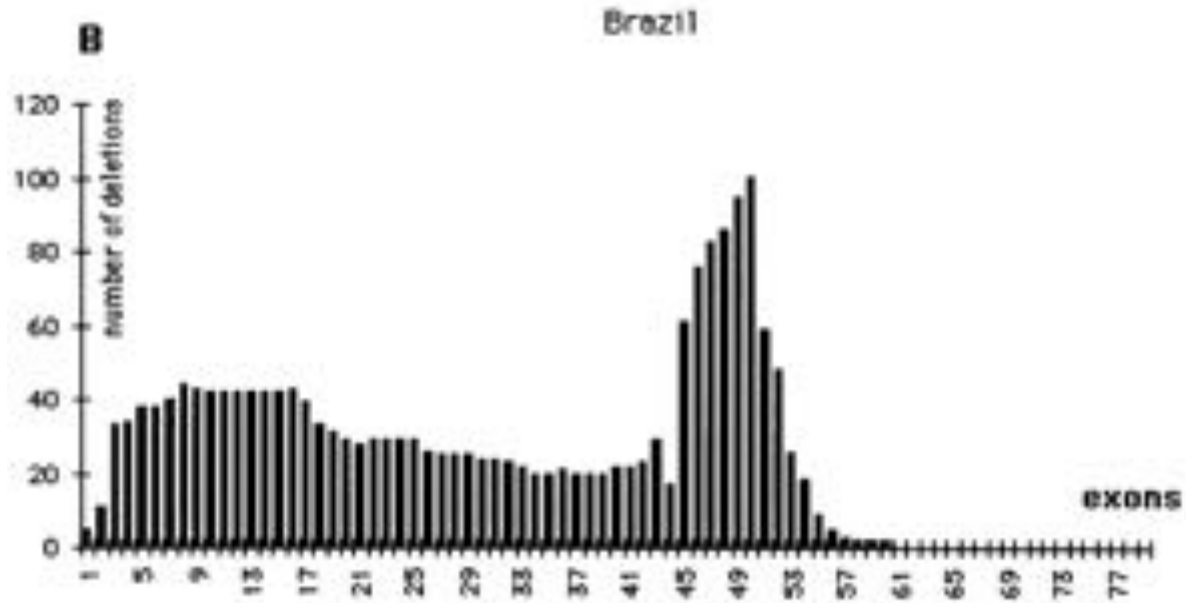
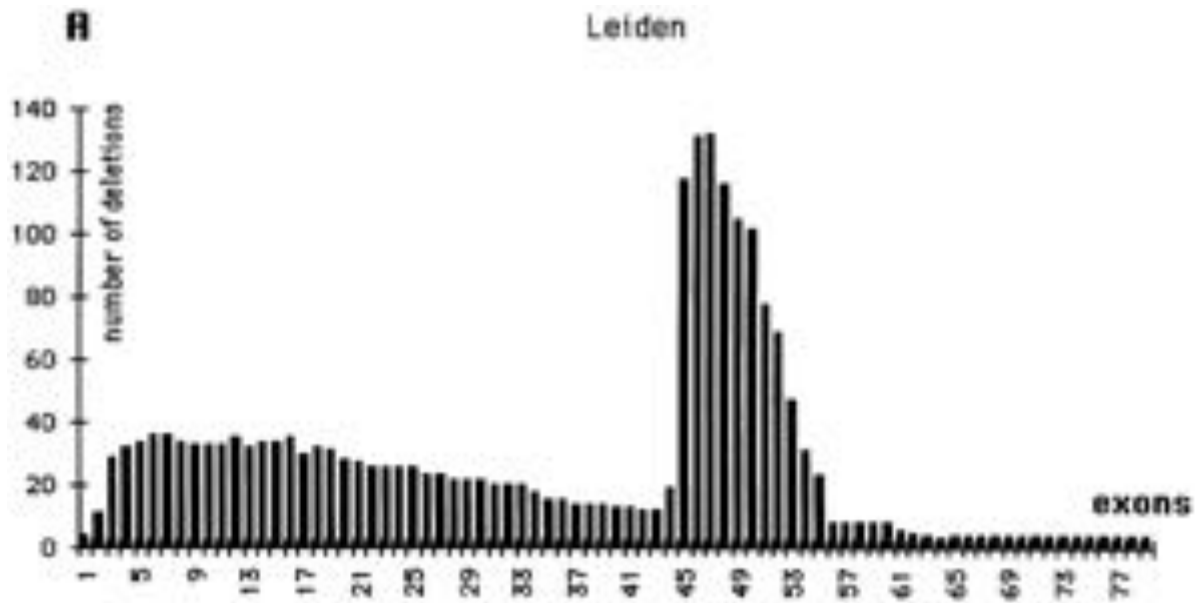
Pt 42719

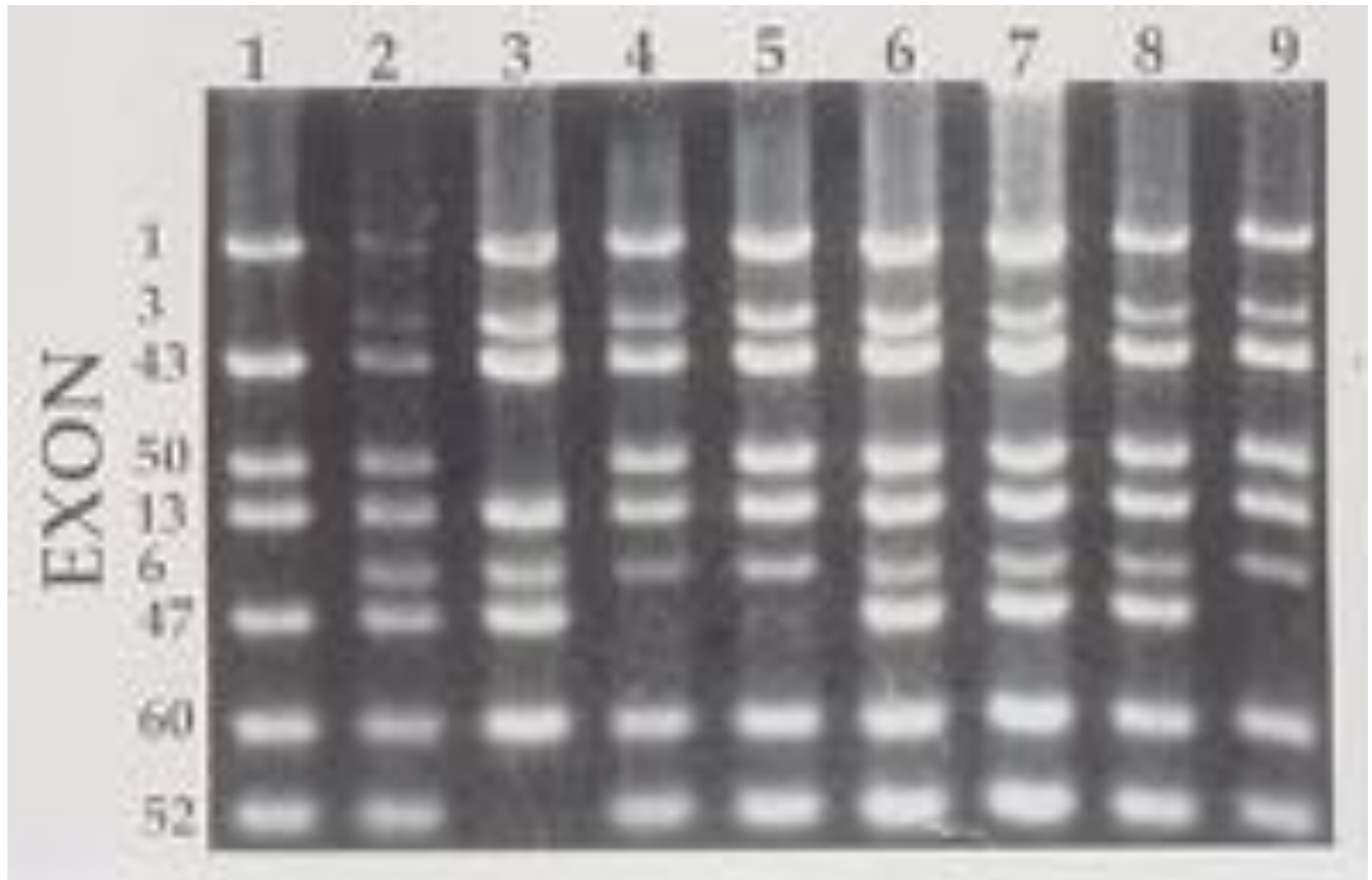




**Dystrophin
(427 kd)**

GAPDH

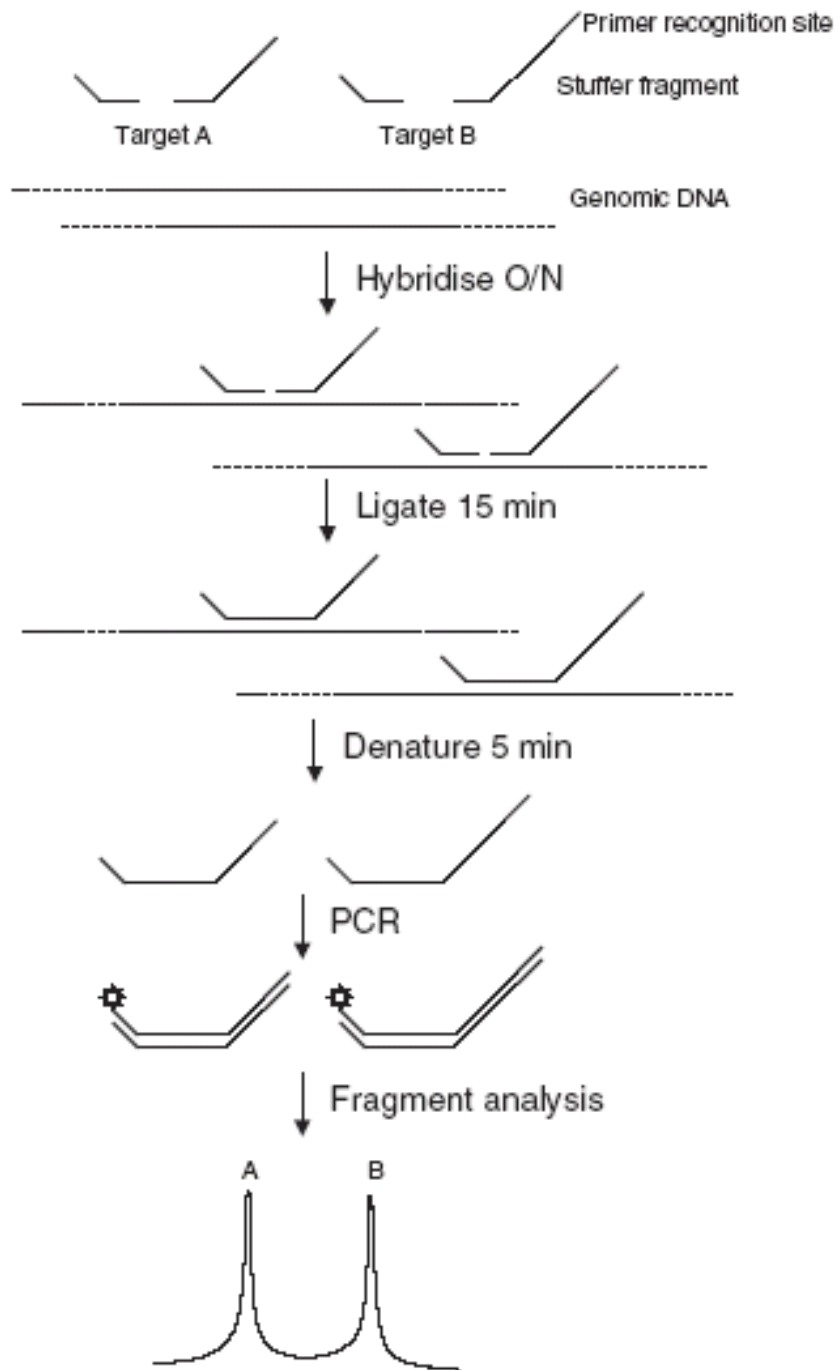




Amplification of 25 exons detects ~98% of exonic deletions

Quantitative techniques detect duplications, and carriers of deletions/duplications

- ~~Multiplex PCR~~
- Southern blotting
- Quantitative PCR (radiolabeled/fluorophore/dye concentration)
- Multiplex Amplifiable Probe Hybridization (MAPH)
- Multiplex Ligation-dependent Probe Amplification (MLPA)
- Comparative Genomic Hybridization (CGH)

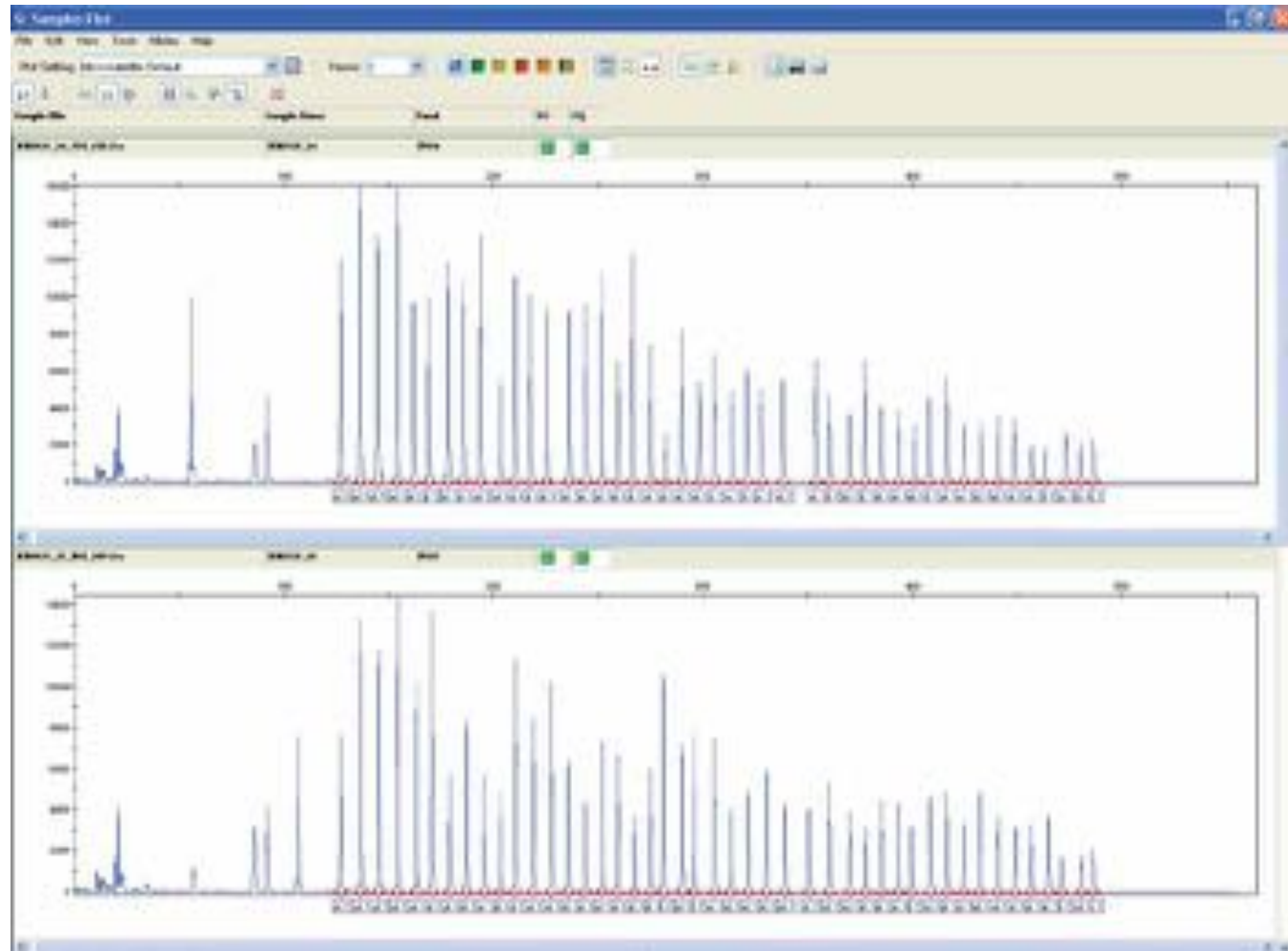
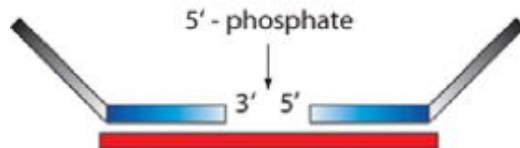


Multiplex Ligation-Dependent Probe Amplification (MLPA)

Sellner & Taylor, Hum Mutat. 2004
May;23(5):413-9

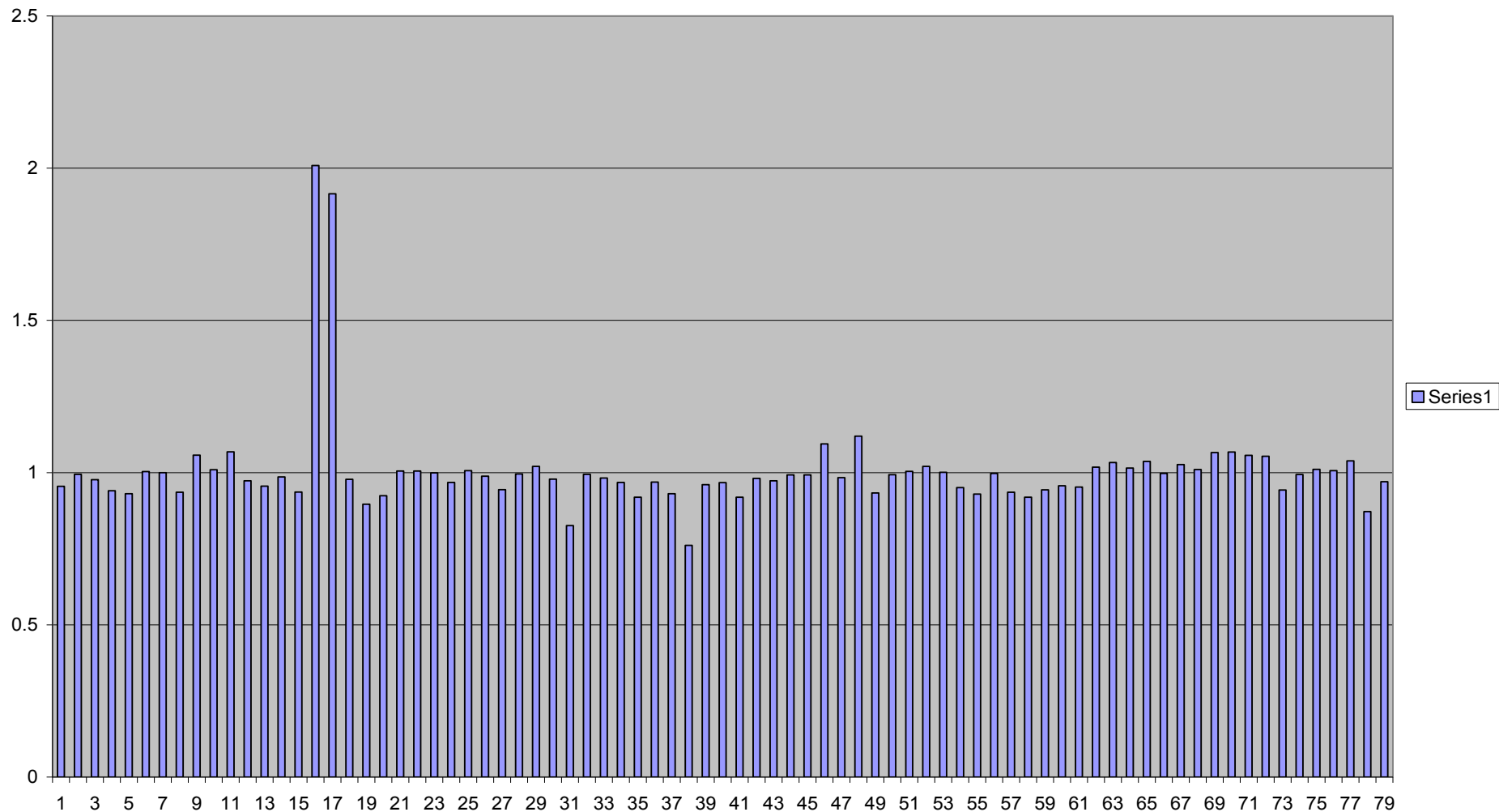
MLPA as first step

MLPA Probes: total length 115-400 bp



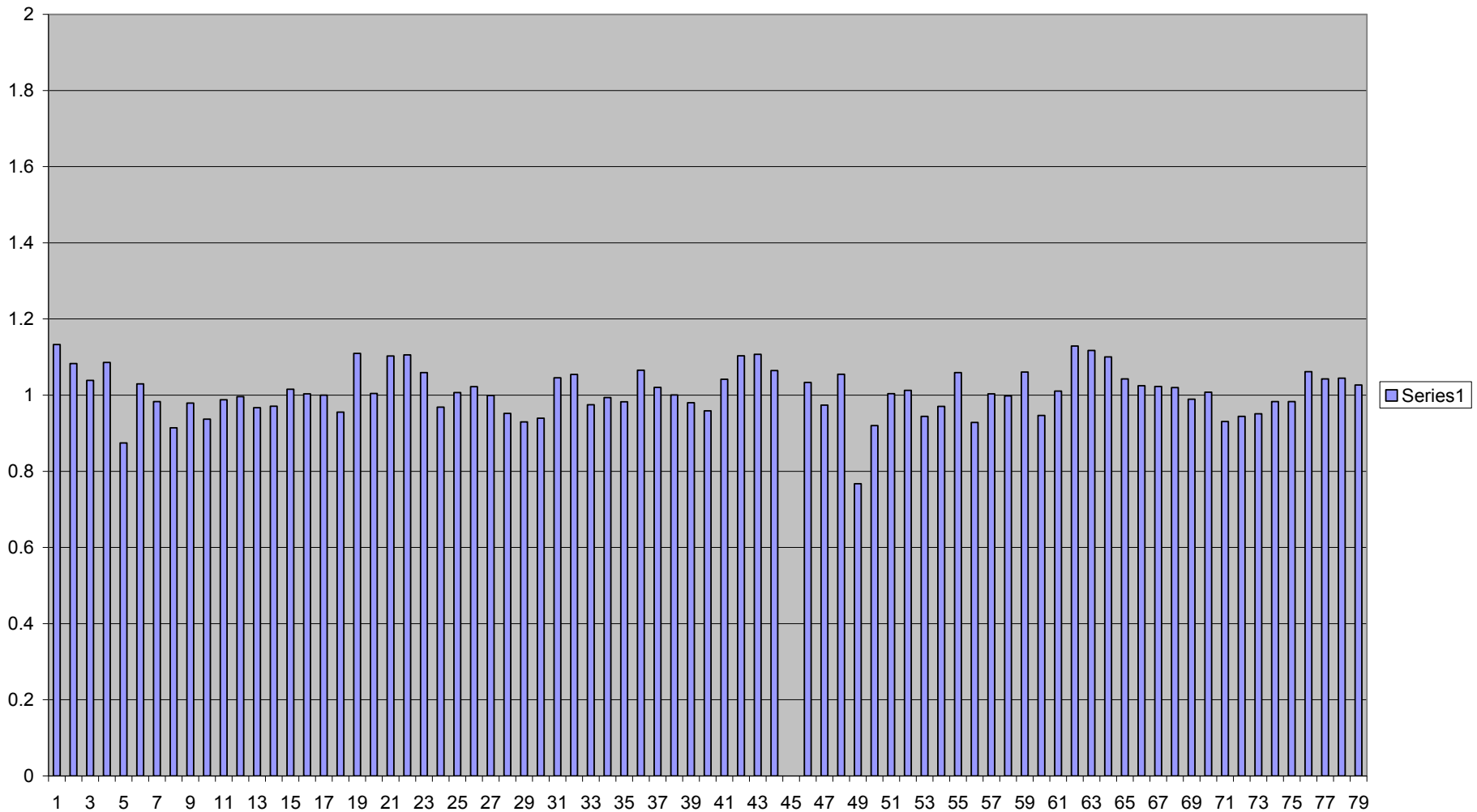
Duplication exons 18-19

43460



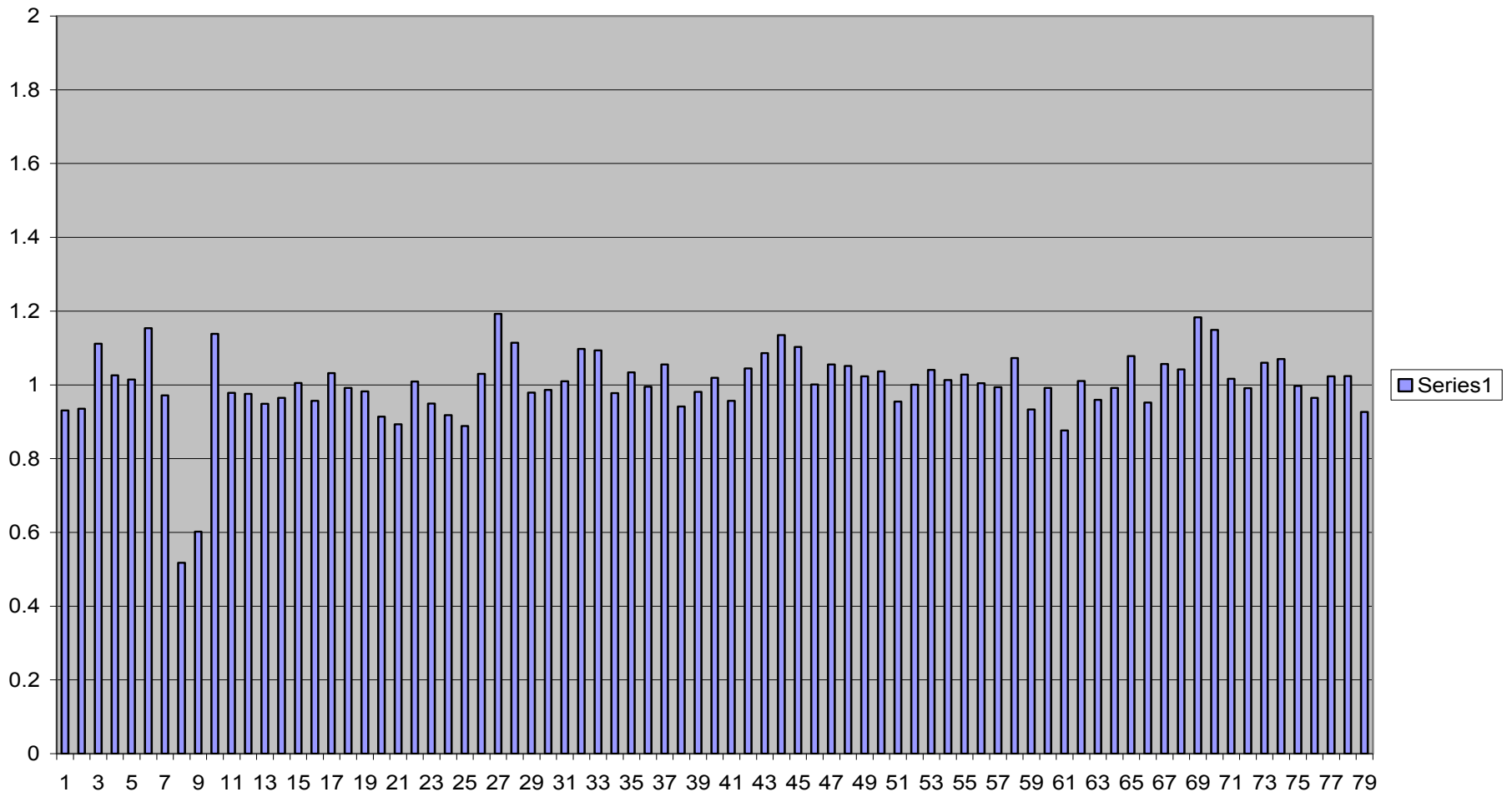
Deletion exon 45

43060



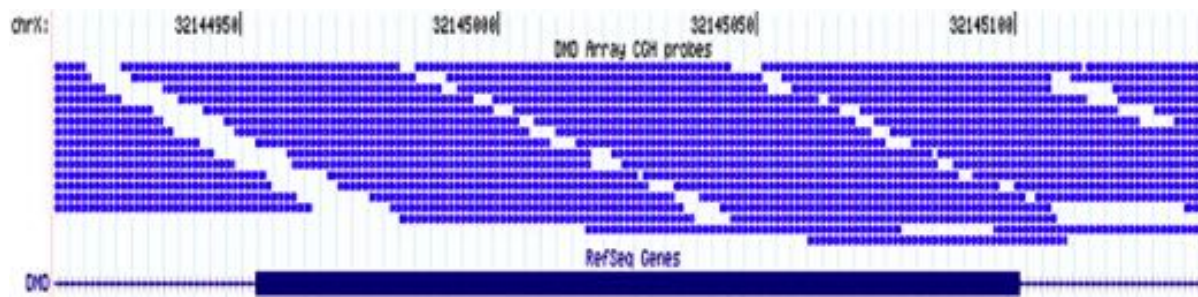
Deletion exons 8-9

43410(F)

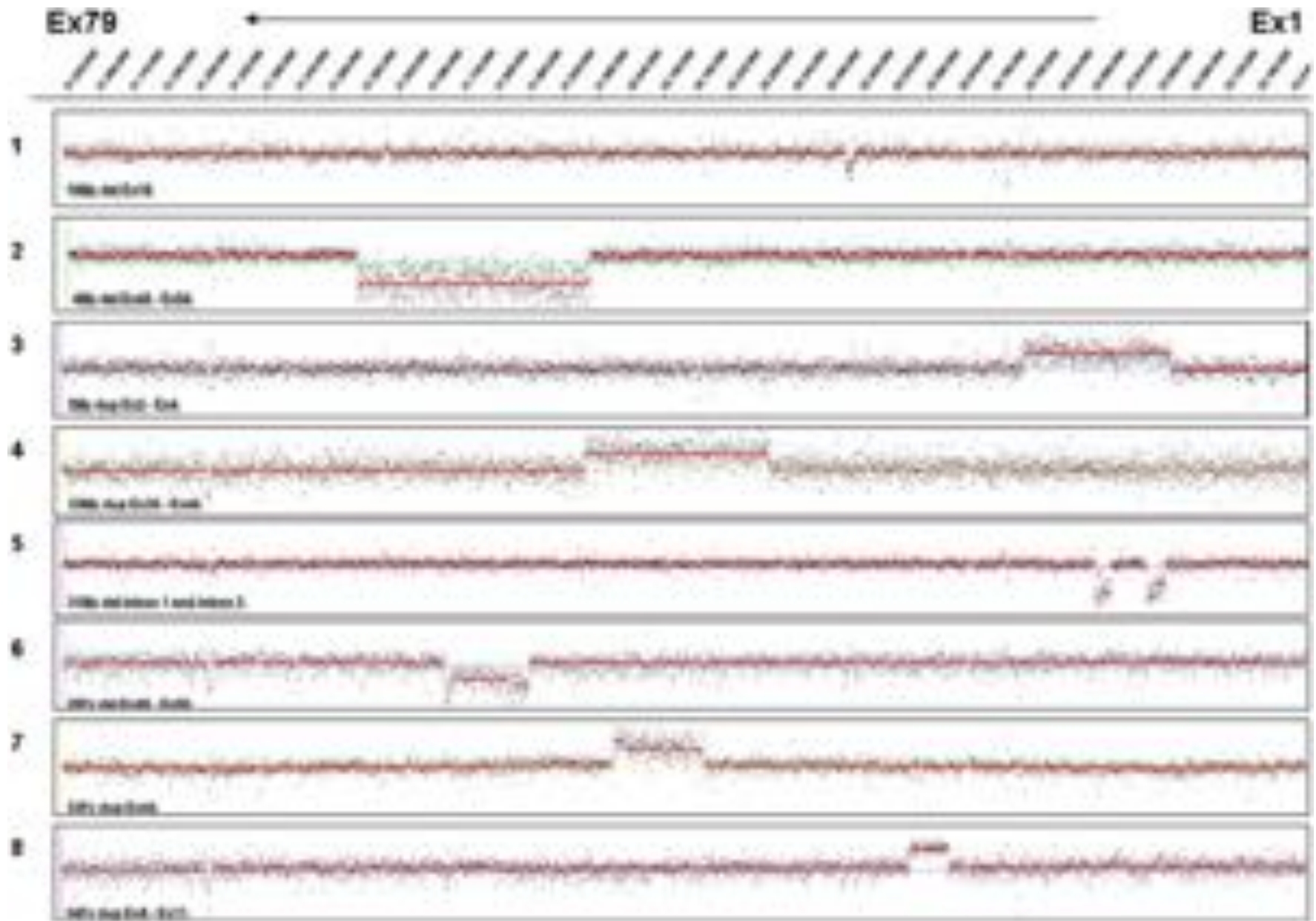


Array-CGH

(comparative genomic hybridization)



Hegde et al, Human Mutation 29(9),1091-1099, 2008



Hegde et al, Human Mutation 29(9),1091-1099, 2008

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Currently available methodology can detect 93%-96% of dystrophinopathy mutations from blood samples.

(Yan *et al*, Hum Mutat 2004; 23:203-204).

Mutational Spectrum of DMD Mutations in Dystrophinopathy Patients: Application of Modern Diagnostic Techniques to a Large Cohort

Kevin M. Flanigan,^{1-4*} Diane M. Dunn,¹ Andrew von Niederhausern,¹ Payam Soltanzadeh,¹ Eduard Gappmaier,¹ Michael T. Howard,¹ Jacinda B. Sampson,² Jerry R. Mendell,⁵ Cheryl Wall,⁵ Wendy M. King,⁵ Alan Pestronk,^{6,7} Julaine M. Florence,⁶ Anne M. Connolly,⁶ Katherine D. Mathews,⁸ Carrie M. Stephan,⁸ Karla S. Laubenthal,^{1,8} Brenda L. Wong,^{9,10} Paula J. Morehart,¹⁰ Amy Meyer,¹⁰ Richard S. Finkel,^{11,12} Carsten G. Bonnemann,^{11,12} Livija Medne,¹¹ John W. Day,¹³ Joline C. Dalton,¹³ Marcia K. Margolis,¹³ Veronica J. Hinton,¹⁴ the United Dystrophinopathy Project Consortium,[†] and Robert B. Weiss^{1*}

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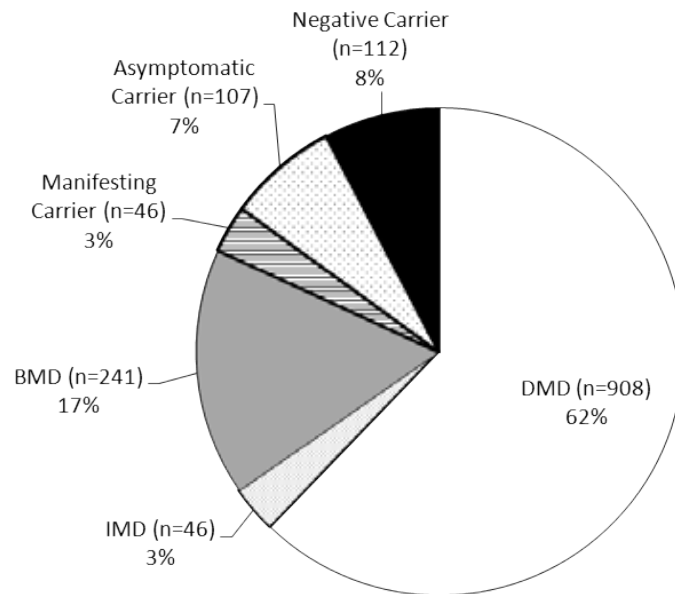
Published online 30 August 2009 in Wiley InterScience (www.interscience.wiley.com). DOI 10.1002/humu.21114

1,111 mutation-positive patients

The United Dystrophinopathy Project: a patient catalogue and clinical research resource

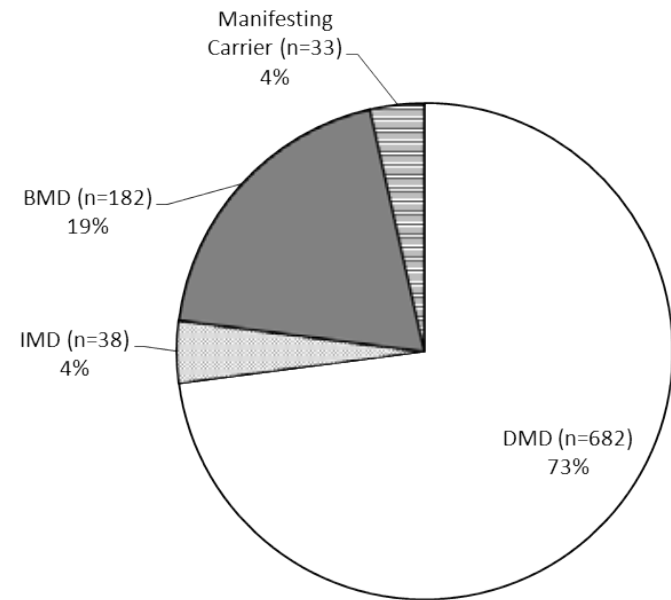
A

Total subjects enrolled (n=1195)



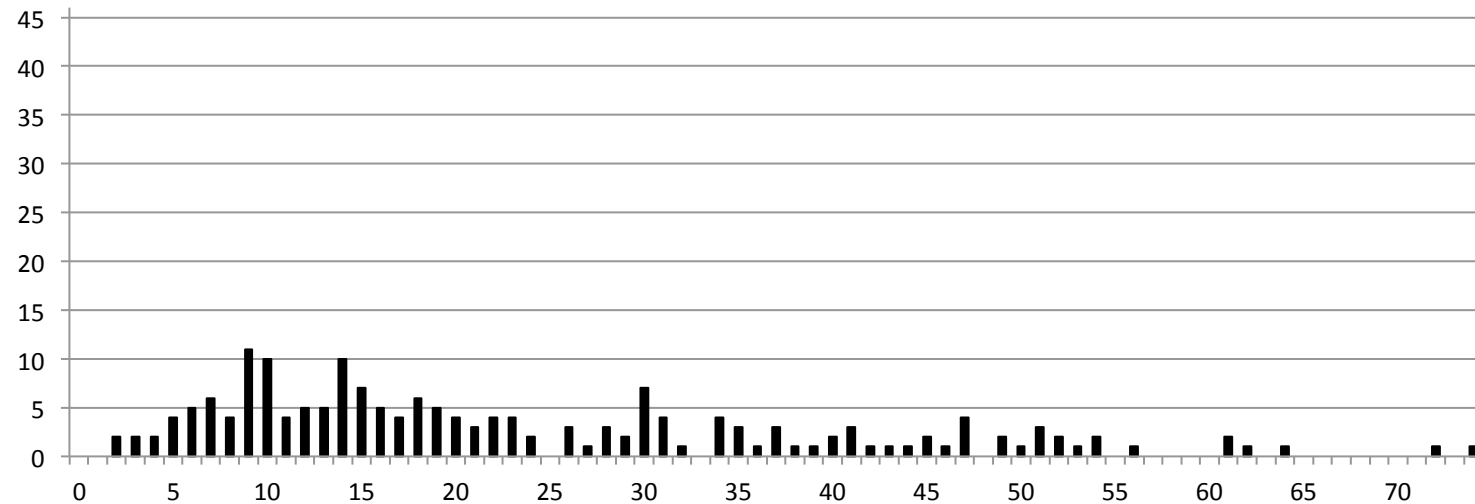
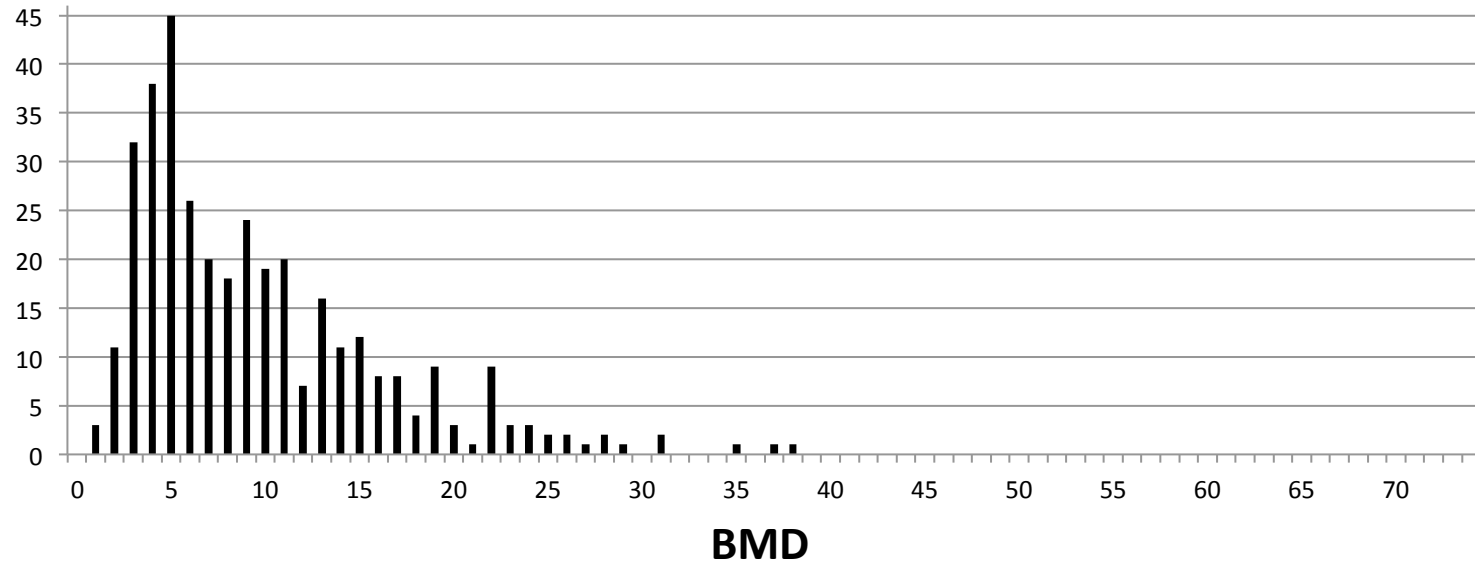
B

Total subjects with clinical evaluation (n=935)



Age at enrollment

DMD

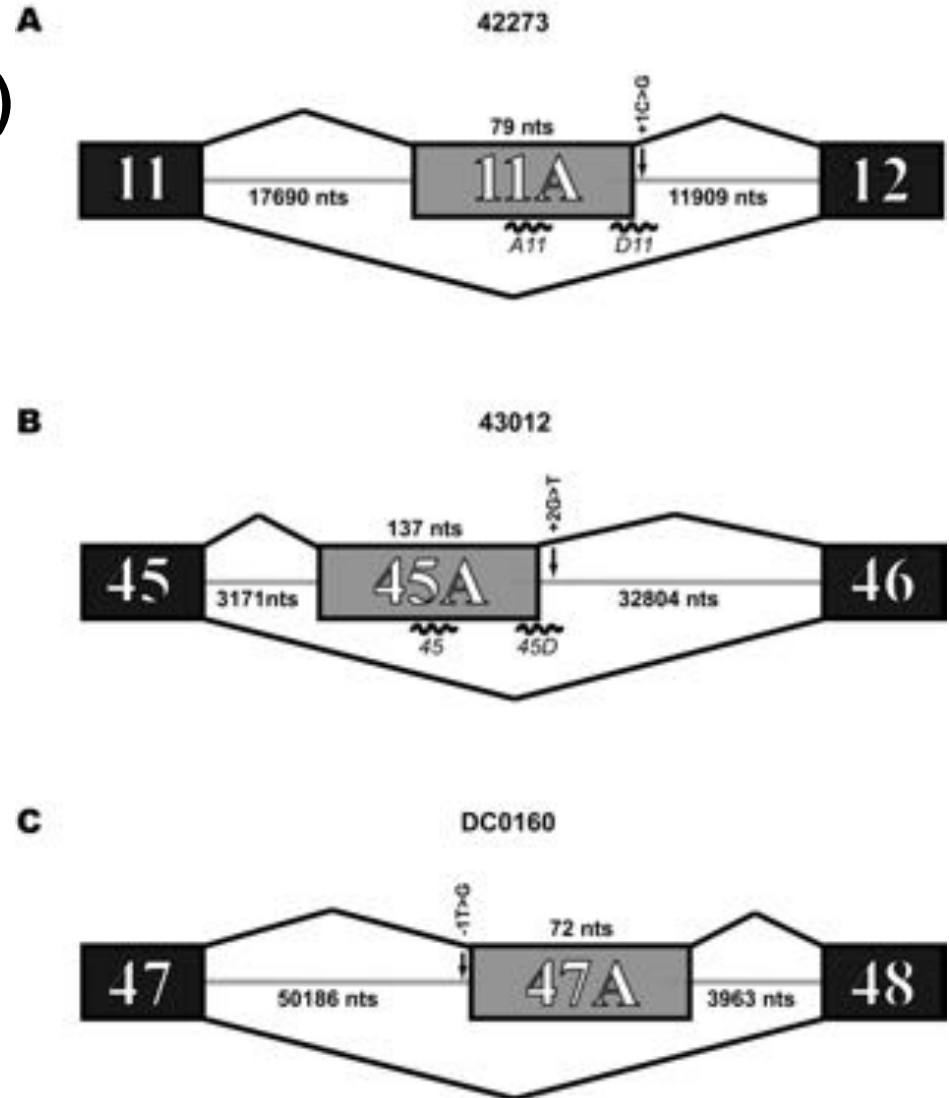


MUTATION CLASS	DMD	IMD	BMD	Unknown (B/DMD)	Manifesting Carrier ^b	Carrier (all phenotypes) ^a	Total	%
DELETION	283	15	55	107	3	14	477	42.9%
<i>in</i>	30	2	36	17	1	2	88	
<i>out</i>	243	13	18	88	1	12	375	
<i>other</i>	10	0	1	2	1	0	14	
STOP	176	4	30	46	4	34	294	26.5%
UGA	60	1	13	20	3	15	112	
UAG	71	0	11	13	0	4	99	
UAA	45	3	6	13	1	15	83	
SUBEXONIC	70	0	10	32	1	14	127	11.4%
FS Ins	22	0	1	7	1	6	37	
FS	46	0	4	23	0	8	81	
FS Ins/Del	1	0	2	2	0	0	5	
<i>in-frame deletion</i>	1	0	3	0	0	0	4	
DUPLICATION	87	7	10	8	5	5	122	11.0%
SPLICE	22	3	7	18	2	12	64	5.8%
MISSENSE	2	1	6	6	0	0	15	1.4%
PSEUDOEXON	0	2	2	0	0	2	6	0.5%
POTENTIAL	2	0	0	3	0	1	6	0.5%
OTHER	0	0	0	0	0	0	0	0.0%
TOTAL MUTATIONS	642	32	120	220	15	82	1111	100.0%

Mutations may not be detectable by genomic DNA analysis

Pseudoexon mutations (e.g.)

- Deep intronic point mutations
- Create splice donor or acceptor sites
- Intronic DNA included as a “pseudoexon” in mRNA
- Undetectable from blood



The reading frame rule in DMD/BMD

Table 2. The Value of Mutational Reading Frame in Predicting a Phenotype of Duchenne Muscular Dystrophy

	DMD	BMD		
Exonic deletions only				
Truncating (out-of-frame) mutations	254	32	88.8%	Positive predictive value
Non-truncating (in-frame) mutations	30	38	55.9%	Negative predictive value
Sensitivity	89.4%			
Specificity	—	54.3%		
All mutations*				
Truncating mutations	519	79	86.8%	Positive predictive value
Non-truncating mutations	37	63	63.0%	Negative predictive value
Sensitivity	93.3%			
Specificity	—	44.4%		

45-55% of BMD patients have out-of-frame mutations



Nonsense mutations do not always predict DMD

- Mutations 5' in the gene may result in altered translational initiation
 - Founder allele: Trp3X (exon 1)

Pt. 42790
c.9G>A; Trp3X (*DMD*)

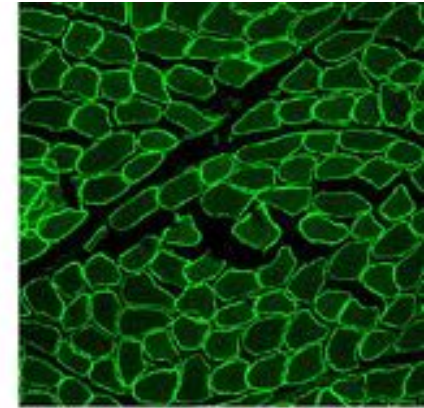
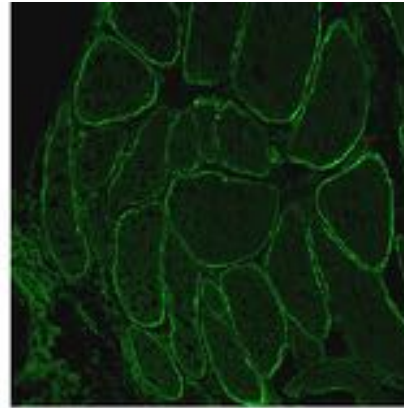
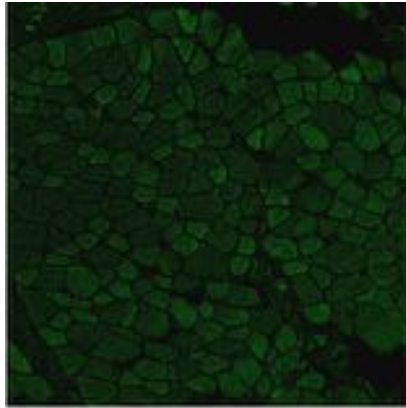
- 65 year old man
 - onset of proximal weakness at age 20
 - wheelchair age 62.
- The proband's brother (examined at age 58)
 - denied symptoms
 - only minimal-to-mild pelvic girdle weakness upon examination.
- Five other historically unrelated families: all had proband boys (<age 10) with no symptoms, but were diagnosed due to elevated serum CPK.

DMD control

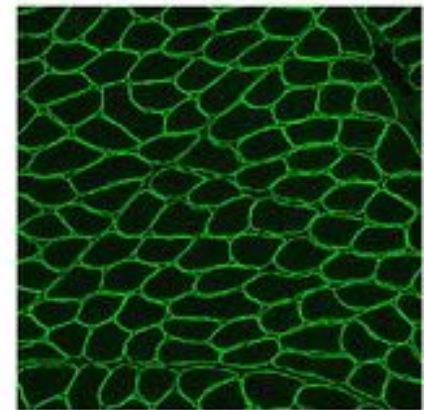
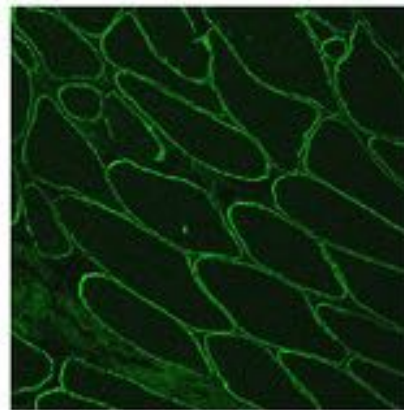
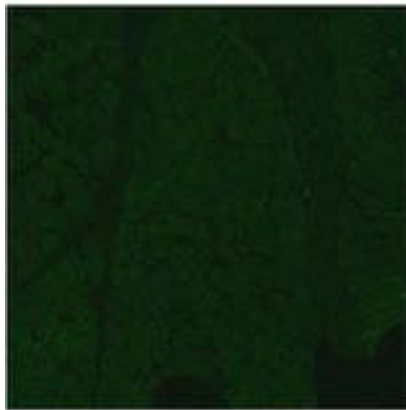
Trp3X

WT

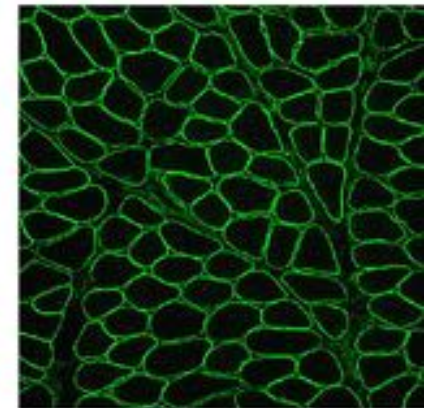
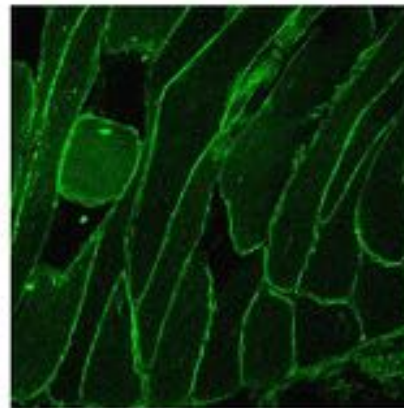
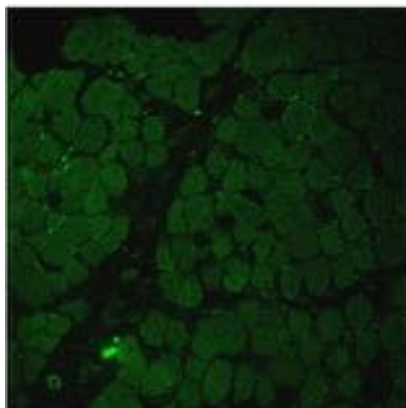
Mandra1
ex.66-79
(C-term)



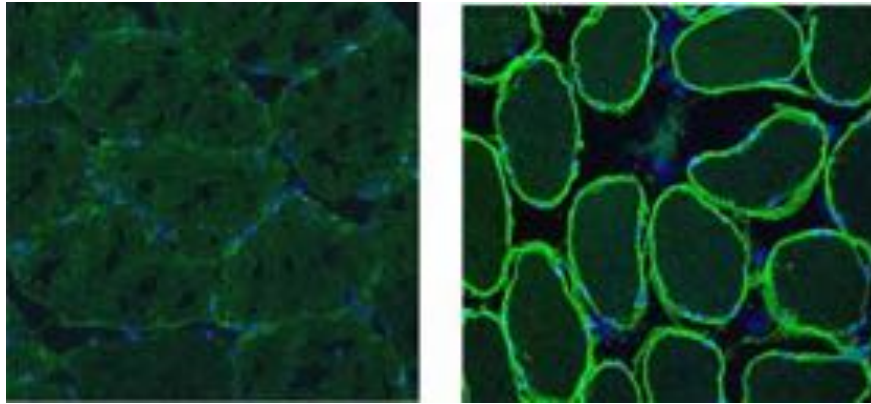
Dys3
ex.10-12
(N-term)



Manex7B
ex.7-8
(N-term)



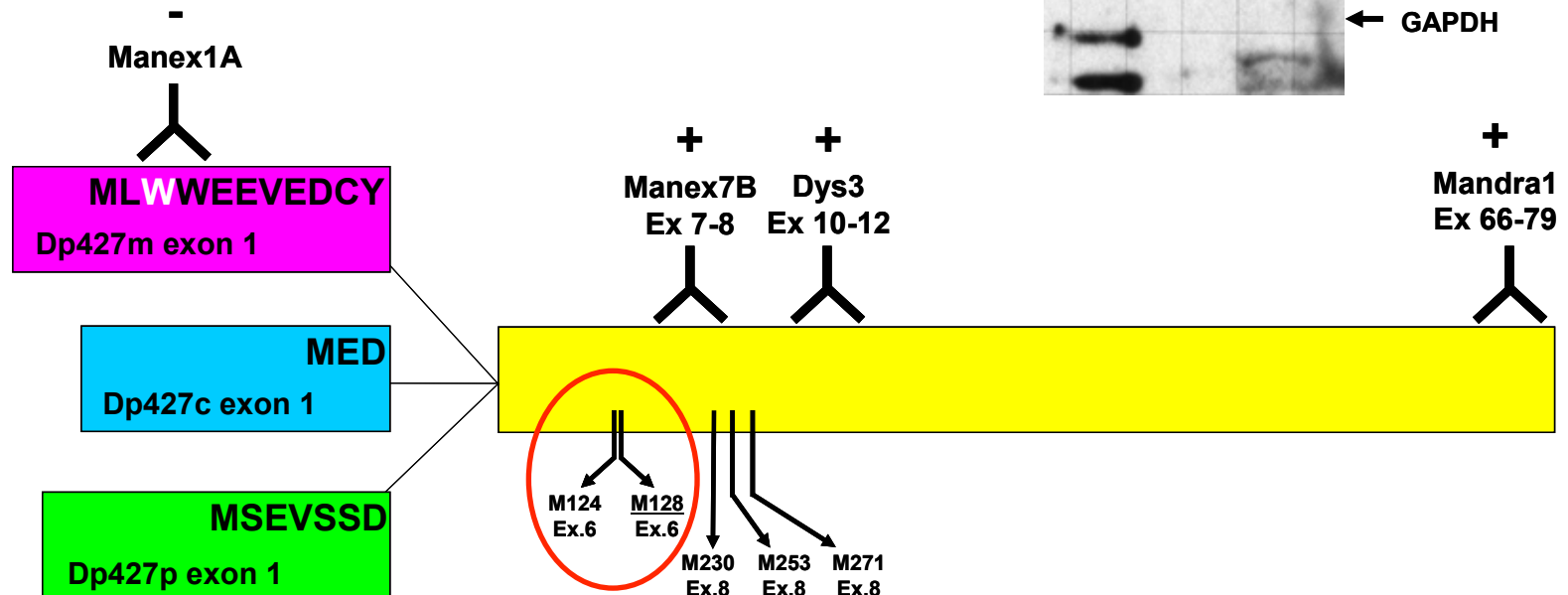
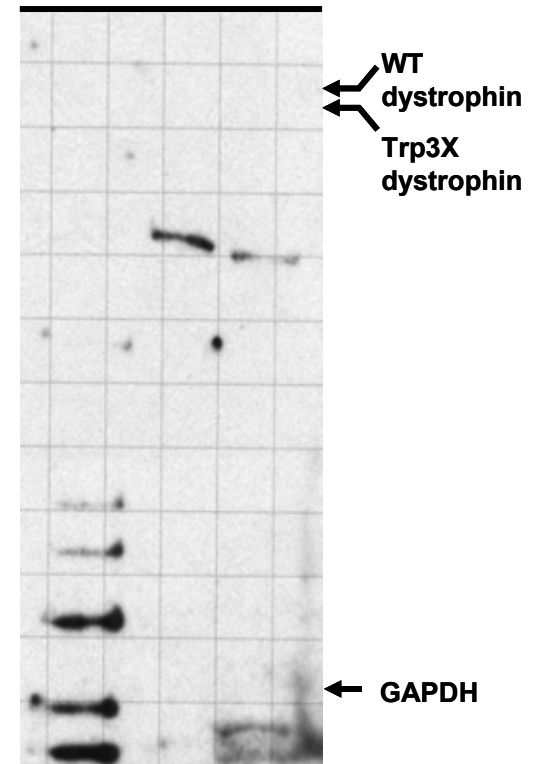
Manex1A



Trp3X

WT

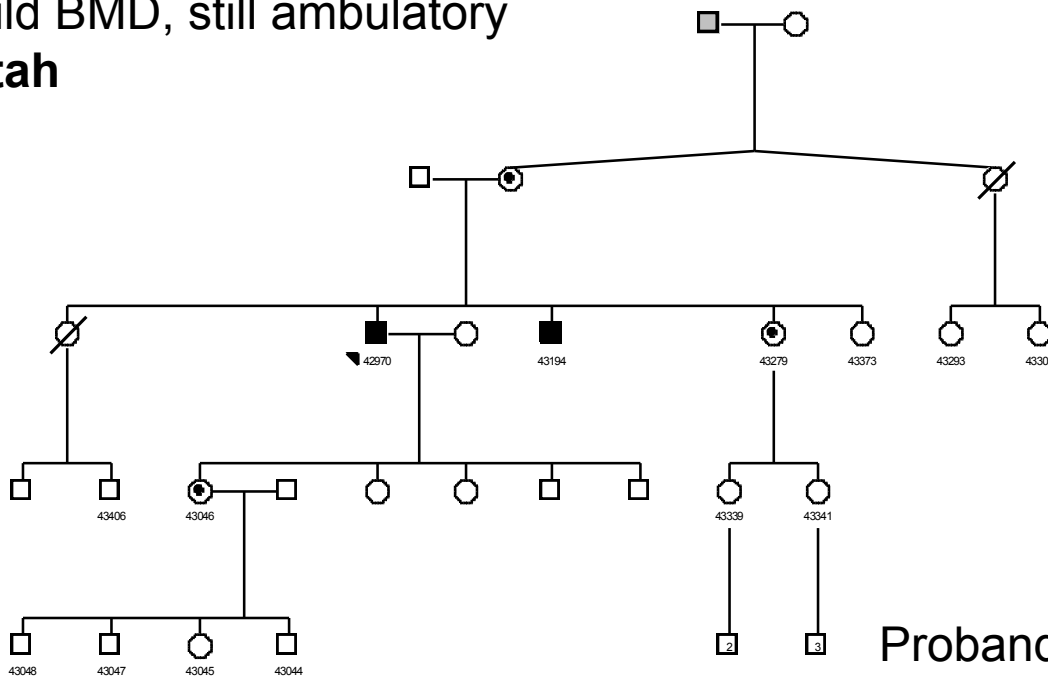
M WT W3X



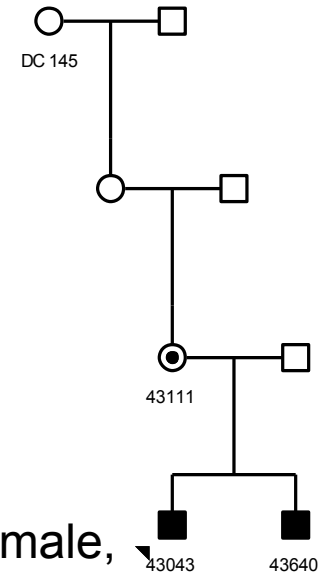
p.Trp3X BMD Pedigrees

Proband = 62 yr. old male,
mild BMD, still ambulatory
Utah

Family 1



Family 2



Proband = 3.5 yr. old male,
incidental to elevated CK levels,
Younger brother CK = 5080 iu/L
Michigan



Concordant resequencing haplotypes across the DMD gene



Trp3X is a founder allele in the *DMD* gene

- Associated with childhood hyperCKemia, with or without myoglobinuria
- Compatible with no significant weakness at age 70
- No effect on reproductive fitness

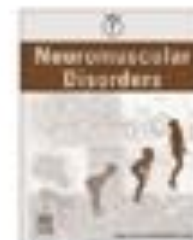
Neuromuscular Disorders 19 (2009) 743–748



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journal homepage: www.elsevier.com/locate/nmd



DMD Trp3X nonsense mutation associated with a founder effect in North American families with mild Becker muscular dystrophy

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Nonsense mutations do not always predict DMD

- Mutations 5' in the gene may result in altered translational initiation
 - Founder allele: Trp3X (exon 1)
- Mutations predicted as nonsense mutations may instead affect exon splice regulatory signals
 - This results in exclusion of exons
 - The remaining mRNA may be in-frame

Nonsense-induced exon skipping

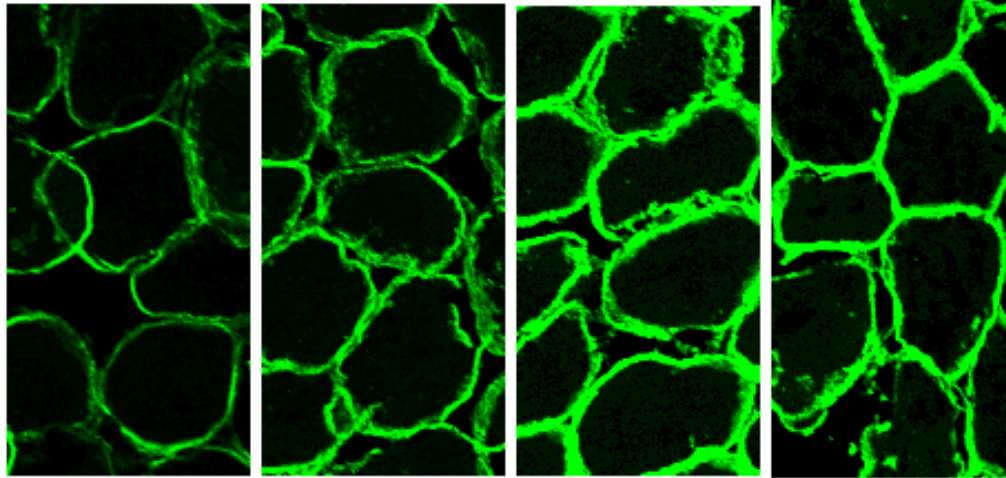
Mandys 19
(exon 20-21)

Mandys 1
(exon 31-32)

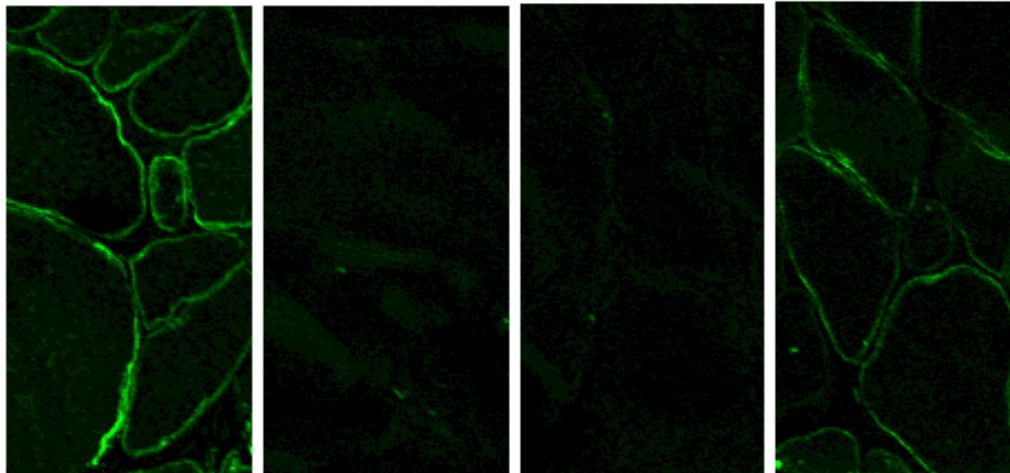
Mandys 8
(exon 32)

Dys 2
(exon 77-79)

Normal



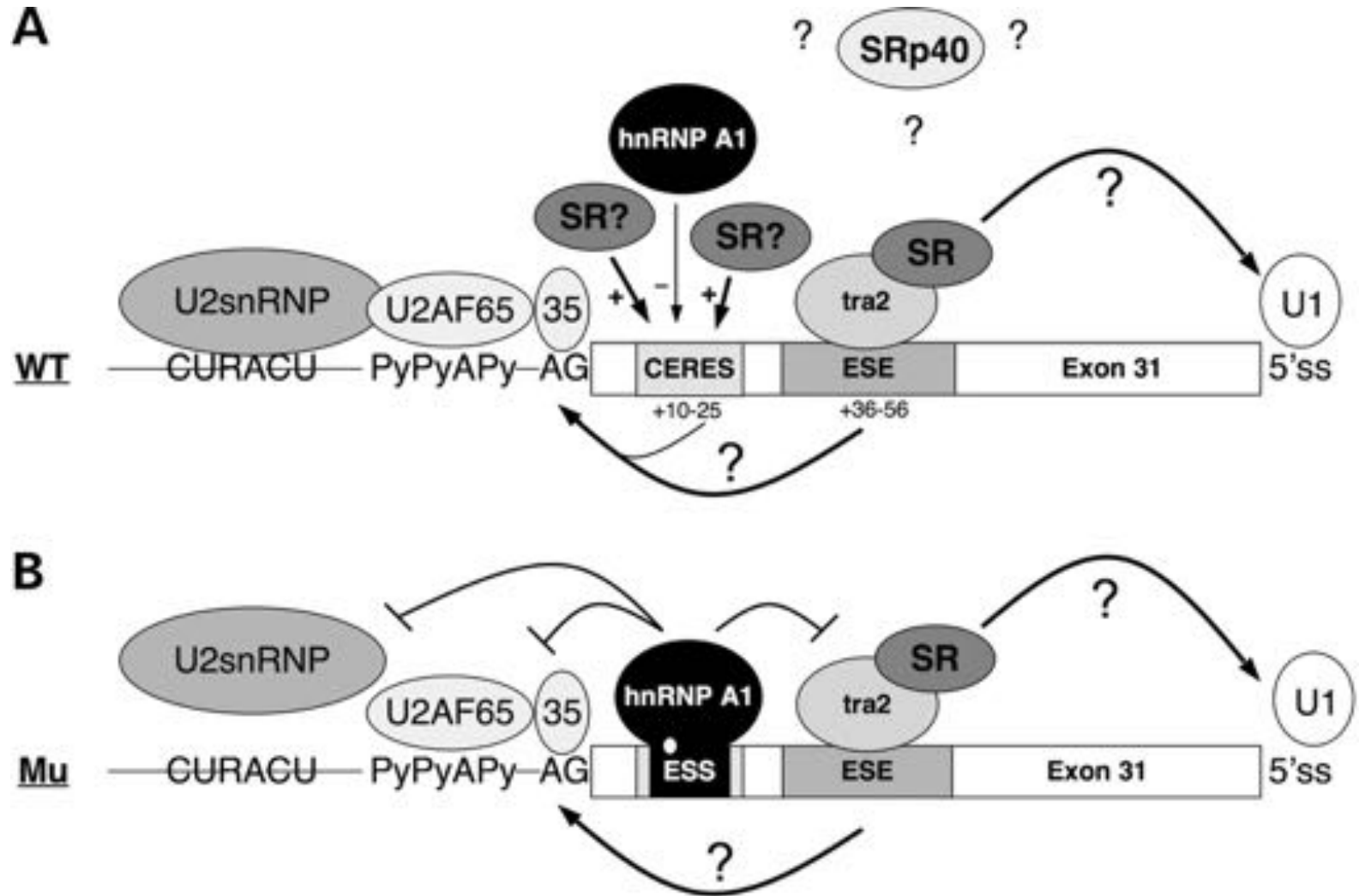
Pt 42719



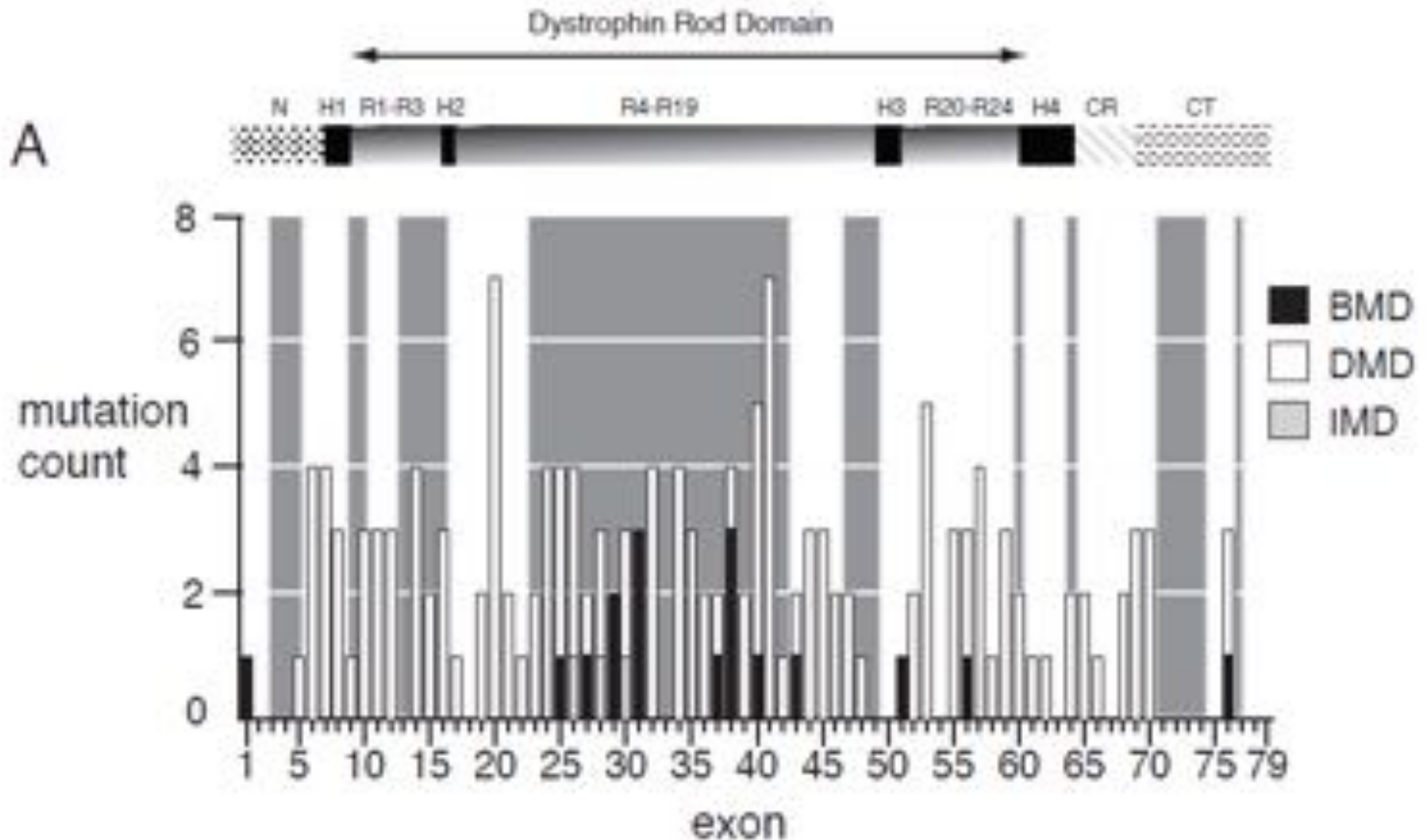
c.4240C>T
p.1414Gly>X

BMD-associated stop codon mutation in exon 31

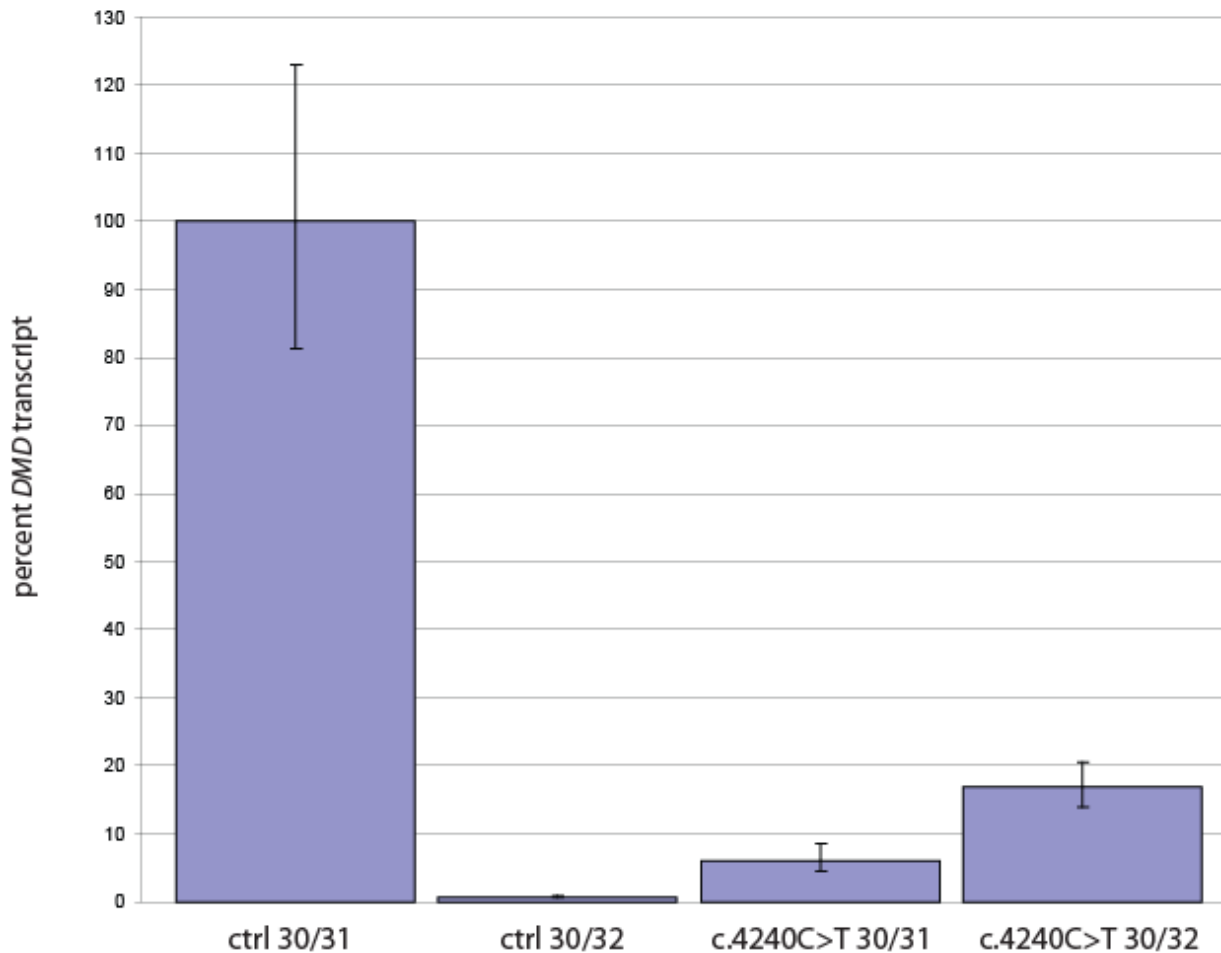
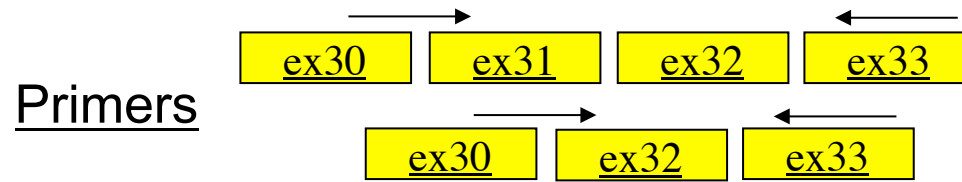
c.4250T>A p.1417Leu>X



161 unique nonsense mutations



Quantitative RT-PCR of exon 31 (c. 4240C>T)





Dystrophinopathies: key points

- Point (non-exon deletion) mutations are nearly all “private” mutations
- Family history is most helpful (but not perfect) in guiding prognosis
- In the absence of family history, reading frame alone should be used cautiously in predicted prognosis
- A place for muscle biopsy remains in the clinical workup



Emerging therapeutic approaches

- Nonsense suppression
- Antisense oligomer-induced exon skipping
- Myostatin inhibition
- Viral gene transfer
 - Gene replacement (microdystrophin, *SGCA*)
 - Expression of other genes (*Galgt2*)
- Recombinant protein approaches
 - NF-kB modulation
 - Biglycan



Acknowledgements

■ University of Utah

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- Olga Gurvich, PhD
- Jay Maiti, MD, PhD

■ Nationwide Children's

- Jerry Mendell, MD
- Adeline Vulin-Chaffiol, PhD
- Nicolas Wein, PhD
- Linda Lowes
- Lindsay Alfano
- Wendy King
- Jack Kaminoh
- Laura Taylor

■ Collaborators

- Alan Pestronk, MD
- Julaine Florence, PhD
- Anne Connolly, MD
- Richard Finkel, MD
- Carsten Bonneman, MD
- Livija Medne, MS
- Brenda Wong, MD
- Kathy Mathews, MD
- John Day, MD
- Craig McDonald, MD

What was thought before the cloning of the Dystrophin gene ?

- 1) Many hypotheses about how DMD worked. A) The vascular hypothesis
B) The neural hypothesis C) The membrane theory (delta lesion in muscle plasma membrane)
- 2) It was known to be an X-linked recessive disorder. This allowed the first mapping.
- 3) Many people said positional cloning of DMD would not work. But geneticists thought it would.
- 4) Many biochemical experiments were done and showed all kinds of differences
For instance as the membrane theory was popular and many cell types were investigated. Fibroblasts, blood cells etc. Indeed abnormalities were reported in many of these cell types later were found not to have dystrophin. Relevance ?
- 5) Cloning of DMD is pre-PCR, pre-BACs and large genomic maps. It involved fun stuff like phage and cosmid walking, partial restriction digest and making the phage and cosmid libraries yourself (no kits). Finding single copy probes, lots of radioactivity and best of all southern blots.

Construction of a genetic linkage map in man using restriction fragment length polymorphism.

[Botstein, D., R.L. White, M. Skolnick, and R.W. Davis. 1980.](#)
Am. J. Hum. Genet. 32: 314. 1980

Cloning of a representative genomic library of the human X chromosome after sorting by flow cytometry. Nature

[Davies KE, Young BD, Elles RG, Hill ME, Williamson R.](#)
[Nature.](#) 1981 Oct 1;293(5831):374-6.

Human X chromosomes were physically separated using a fluorescence-activated cell sorter. A library of genomic fragments was constructed.

Linkage relationship of a cloned DNA sequence on the short arm of the X chromosome to Duchenne muscular dystrophy

[Murray JM, Davies KE, Harper PS, Meredith L, Mueller CR, Williamson R.](#)
[Nature.](#) 1982 Nov 4;300(5887):69-71.

Established that DMD was on short arm of the X flanking markers indicated Xp21

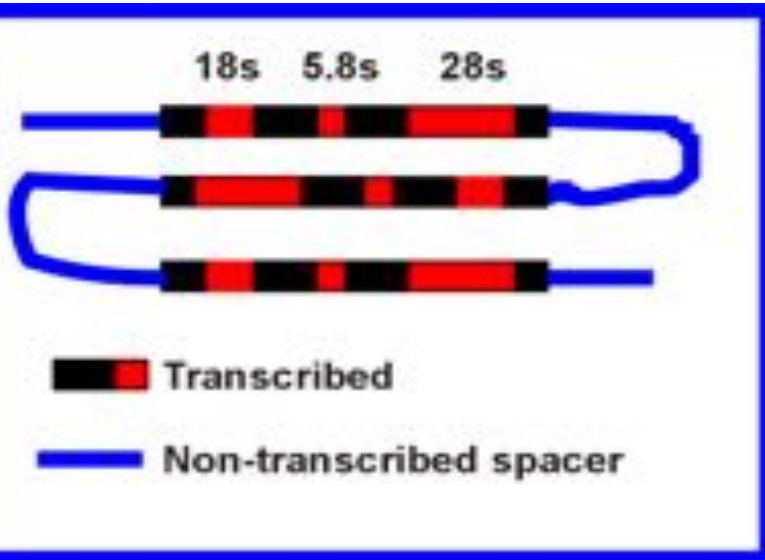
Two key patients for the cloning of DMD

Deletions

Bruce Bryer BB born in 1966 in Spokane WA and was an adopted child. At age three he was identified to have Chronic granulomatous disease (CGD), then at 9 DMD as well. Hans Ochs made cell lines from Bruce Bryer. Roberta Pagon determined that BB also suffered retinitis pigmentosa. Thus three X linked diseases. Pagon speculated that a deletion existed. Thus cytogenetics was performed and a deletion identified this was a large deletion but only just visible with the techniques of the day.

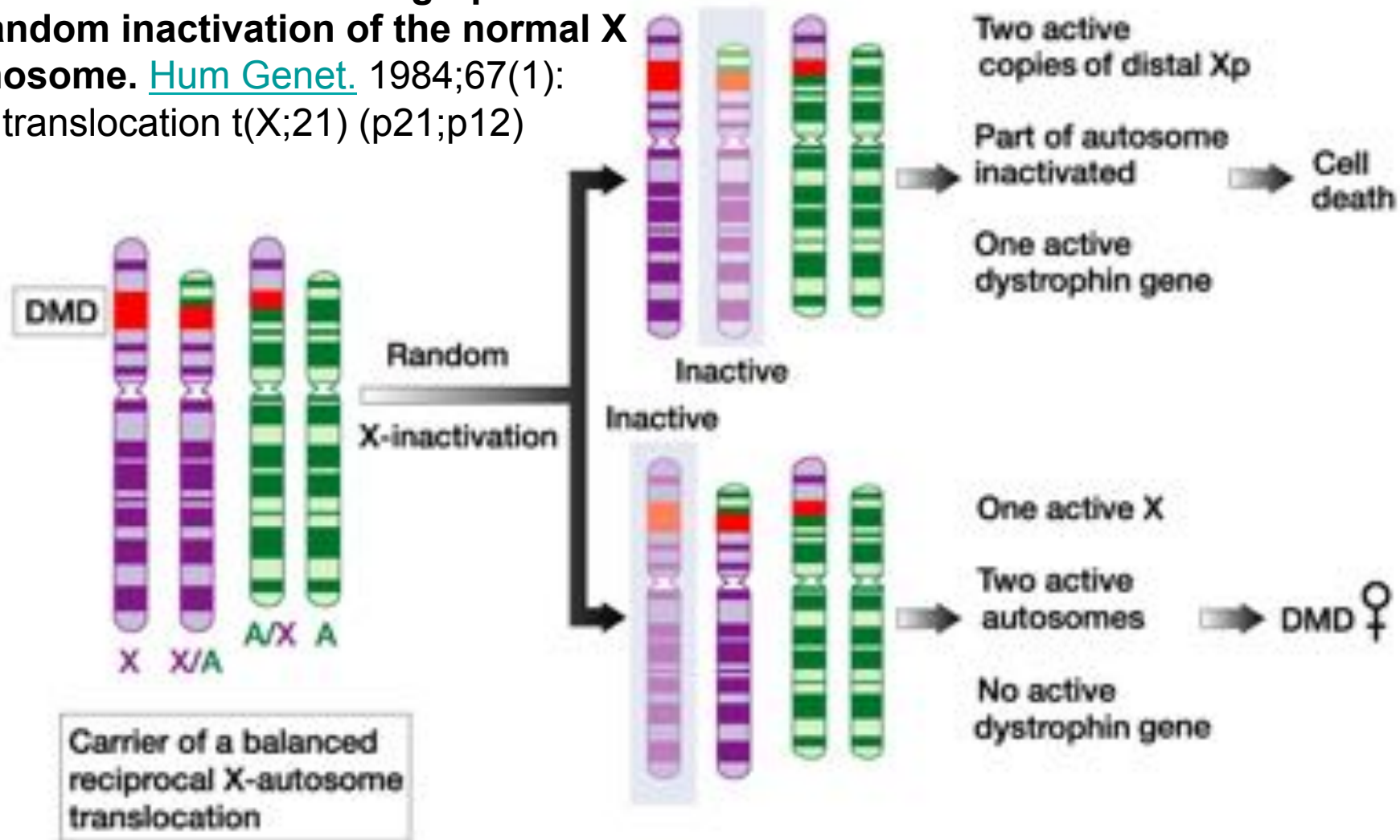
Translocations

Seven females with DMD all with an X-autosome balanced translocation. The X-chromosome break was in Xp21 in all cases. One occurred in a Belgium girl Anne and was $t(X;21)(p21;p12)$ breaking the ribosomal gene cluster on chromosome 21.



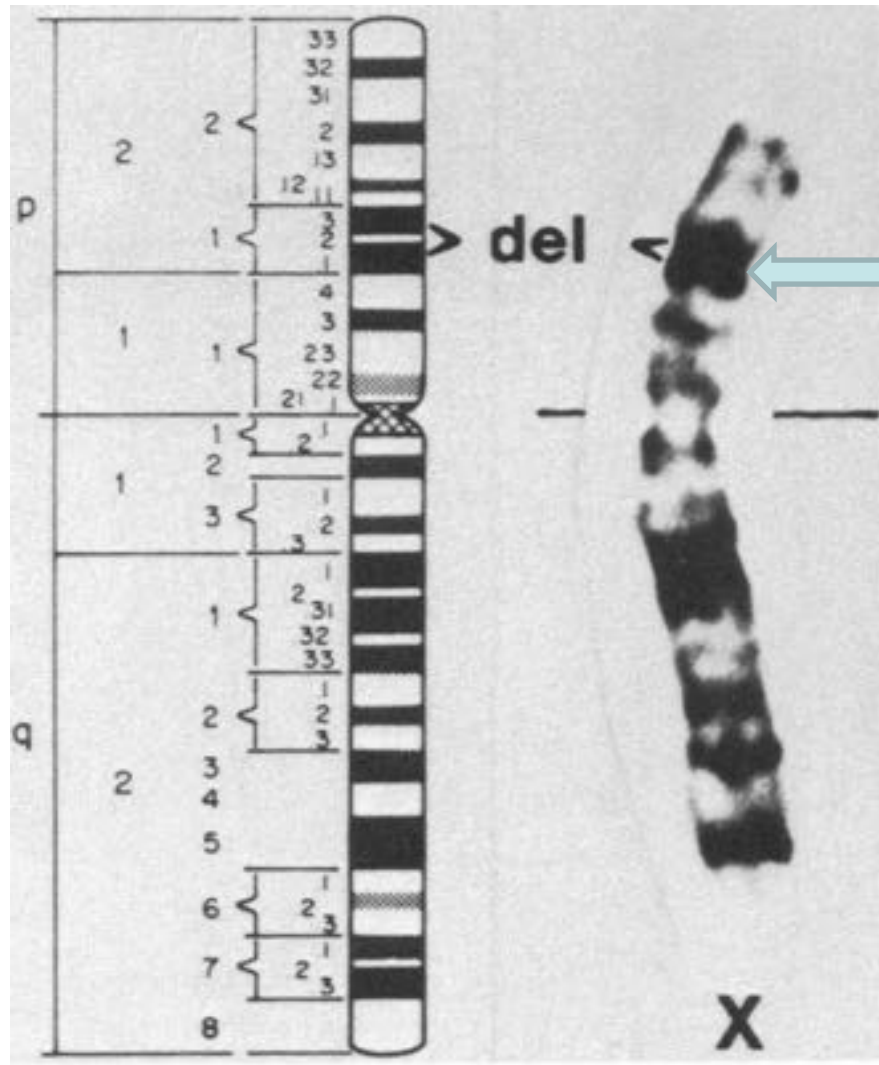
[Science](#). 1984 Jun 29;224(4656):1447-9.
Duchenne muscular dystrophy involving translocation of the dmd gene next to ribosomal RNA genes.
[Worton RG](#), [Duff C](#), [Sylvester JE](#),
[Schmickel RD](#), [Willard HF](#).

[Verellen-Dumoulin C](#), [Freund M](#), [De Meyer R](#),
[Laterre C](#), [Frédéric J](#), [Thompson MW](#),
[Markovic VD](#), [Worton RG](#). **Expression of an
X-linked muscular dystrophy in a female
due to translocation involving Xp21 and
non-random inactivation of the normal X
chromosome.** [Hum Genet.](#) 1984;67(1):
115-9. translocation t(X;21) (p21;p12)



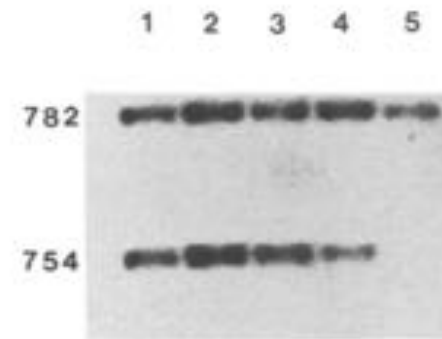
MINOR XP21 CHROMOSOME DELETION IN A MALE ASSOCIATED WITH EXPRESSION OF DUCHENNE MUSCULAR-DYSTROPHY, CHRONIC GRANULOMATOUS-DISEASE, RETINITIS PIGMENTOSA, AND MCLEOD SYNDROME

Francke, U., Ochs, H. D., Demartinville, B., Giacalone, J., Lindgren, V., Distèche, C., Pagon, R. A., Hofker, M. H., Van-Ommen, G. J., Pearson, P. L., Wedgwood, R. J. AMERICAN JOURNAL OF HUMAN GENETICS 1985; 37 (2): 250-267



White space of
Xp21 missing in BB

Deletion confirmed by southern
using probe 754



BB

**Cloning of the breakpoint of an X;21 translocation associated with
Duchenne muscular dystrophy.**

[Ray PN](#), [Belfall B](#), [Duff C](#), [Logan C](#), [Kean V](#), [Thompson MW](#),
[Sylvester JE](#), [Gorski JL](#), [Schmickel RD](#), [Worton RG](#).

Nature. 1985 Dec 19-1986 Jan 1;318(6047):672-5.

Genetics Department and Research Institute, The Hospital for
Sick Children and The Departments of Medical Genetics and
Biophysics, University of Toronto, 555 University Avenue,
Toronto, Ontario M5G 1X8, Canada

**Detection of deletions spanning the Duchenne muscular dystrophy locus using
a tightly linked DNA segment.**

[Monaco AP](#), [Bertelson CJ](#), [Middlesworth W](#), [Colletti CA](#), [Aldridge J](#),
[Fischbeck KH](#), [Bartlett R](#), [Pericak-Vance MA](#), [Roses AD](#), [Kunkel LM](#).

Nature. 1985 Aug 29-Sep 4;316(6031):842-5

Division of Genetics, Mental Retardation Program,
Department of Pediatrics, Harvard Medical School,
The Children's Hospital, Boston, Massachusetts
02115, USA

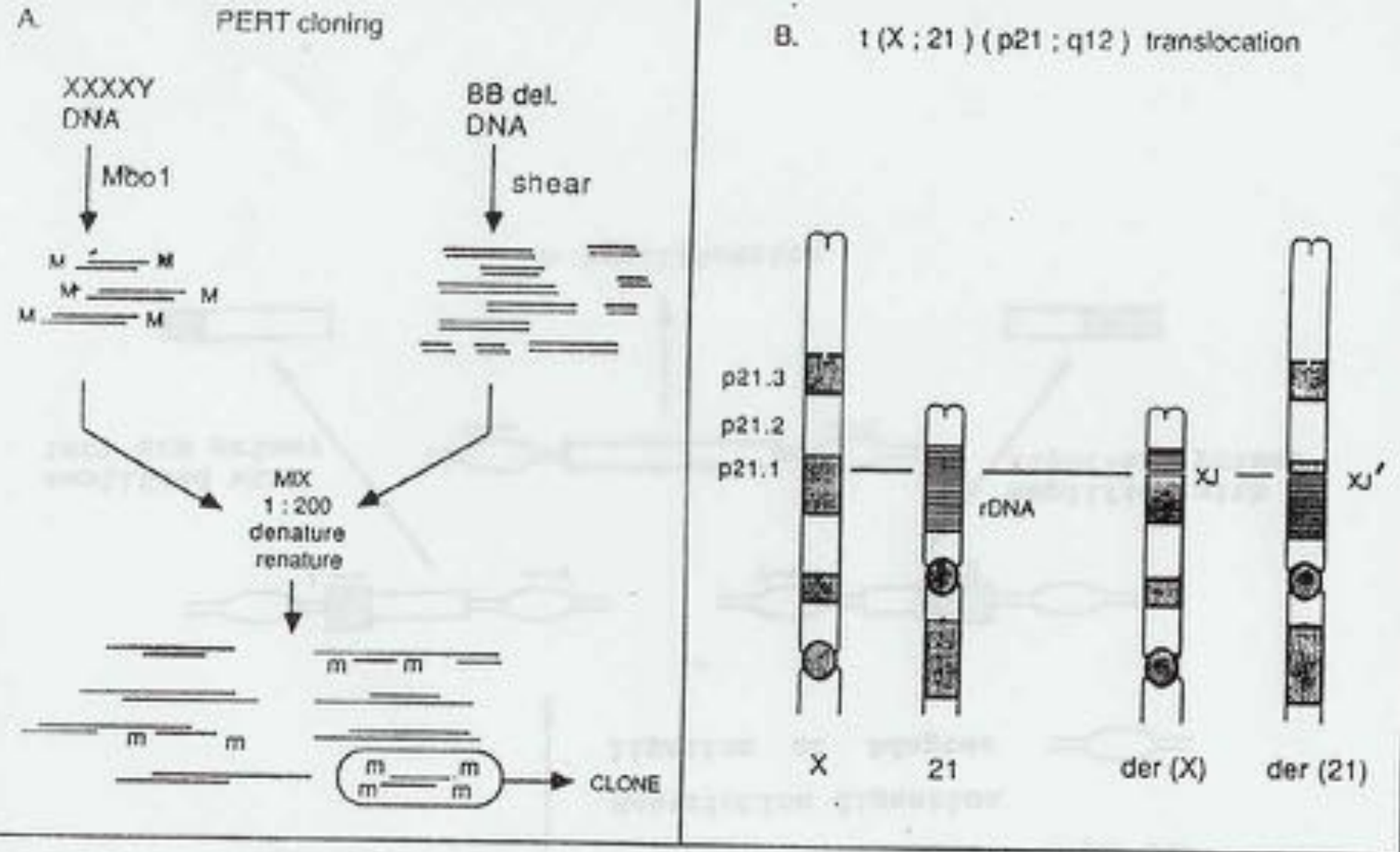
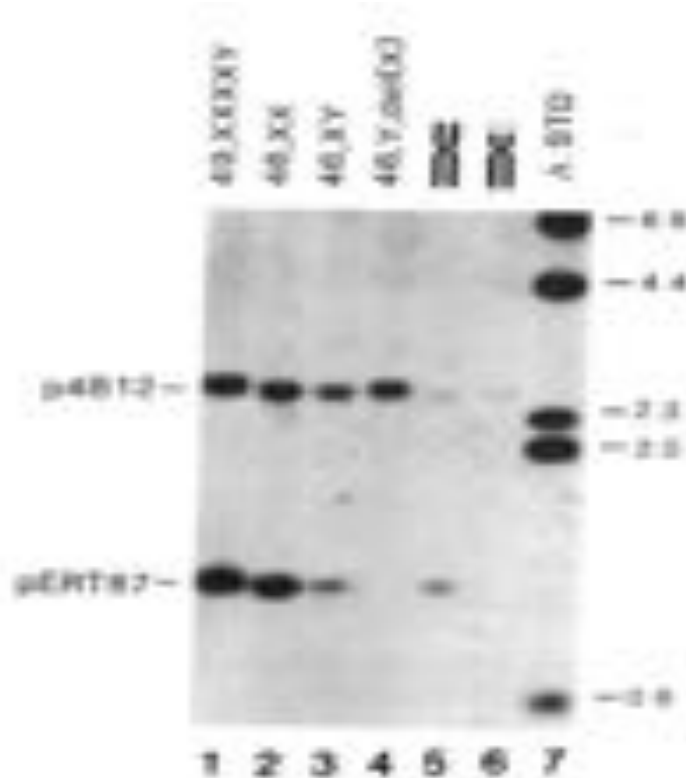


Figure 2 Schematic to illustrate the two successful cloning strategies.

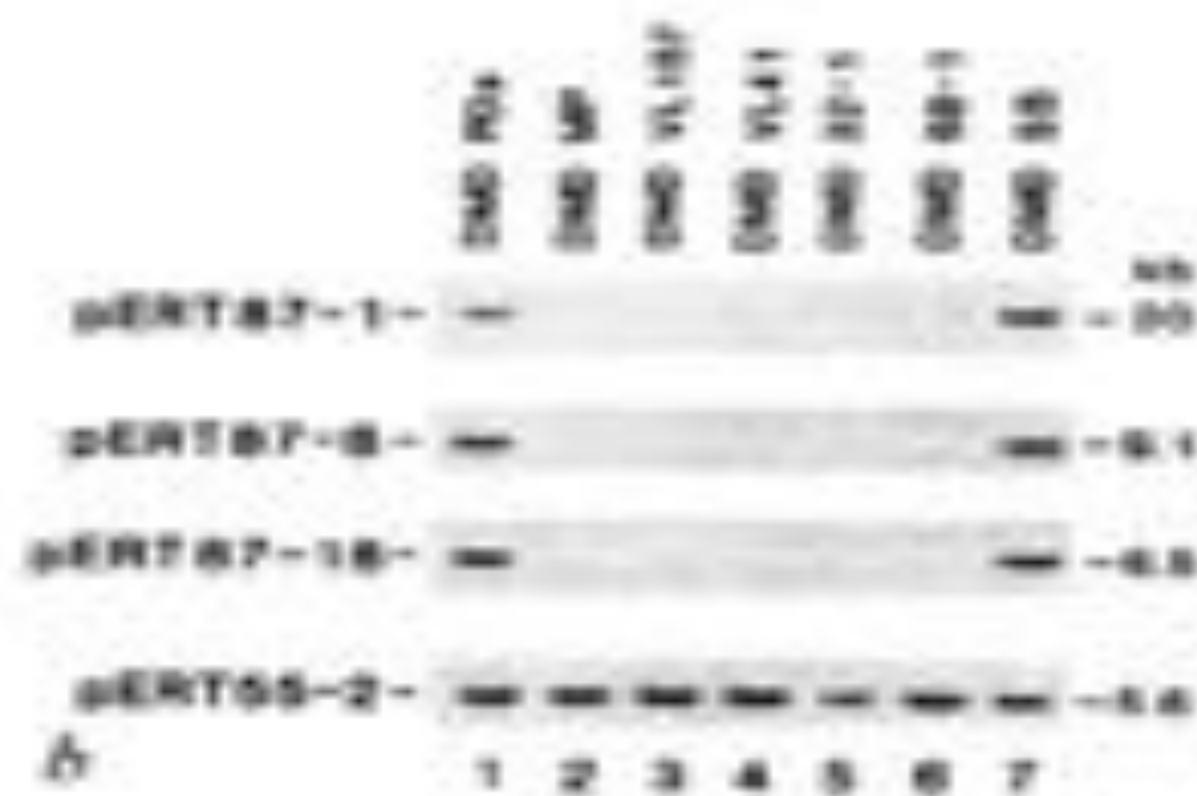
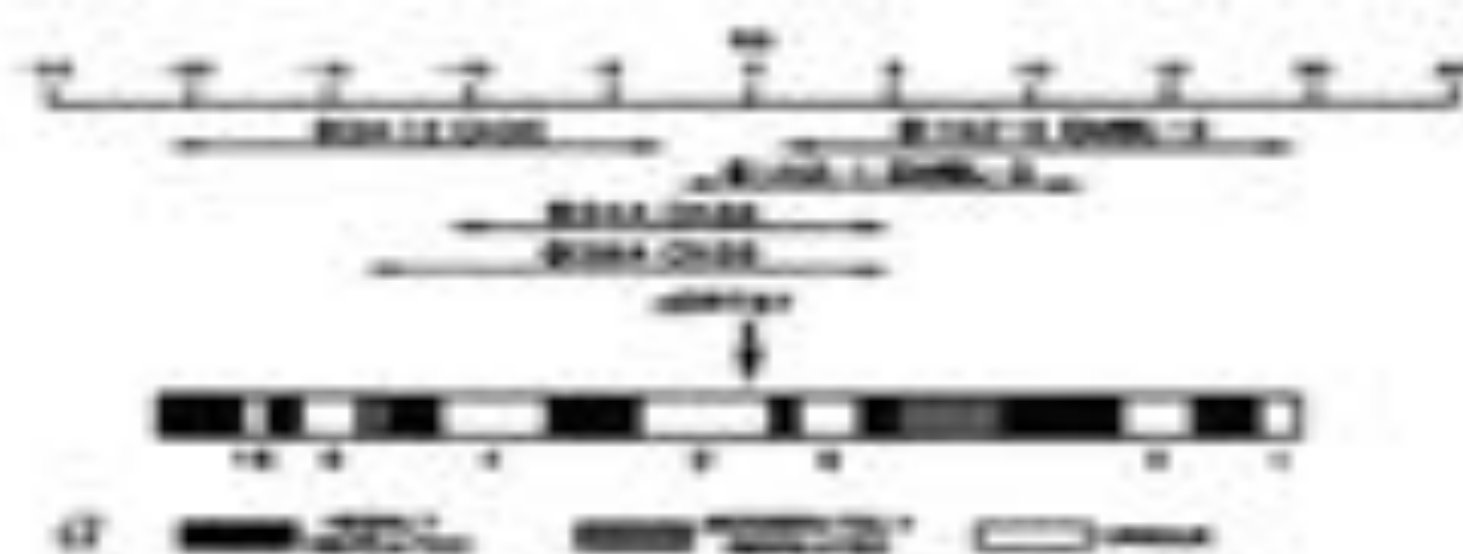
A. The pERT cloning scheme is described in the text. The ends of the *MboI*-digested molecules are indicated as M (double stranded) or m (single stranded). The ends of the sheared molecules are unlabeled. Among the reassociated molecules, only the perfectly aligned *MboI*-digested molecules are clonable, and these are enriched for sequences missing from the DNA of patient DD.

Specific cloning of DNA fragments absent from the DNA of a male patient with an X chromosome deletion

Louis M. KUNKEL, ANTHONY P. MONACO, WILLIAM MIDDLESWORTH, HANS D. OCHS, AND SAMUEL A. LATT Proc. Natl. Acad. Sci. USA Vol. 82, pp. 4778-4782, July 1985



Royer-Pokora, B., L.M. Kunkel, A.P. Monaco, S.C. Goff, P.E. Newburger, R.L. Baehner, ES. Cole, J.T. Curnutte, and S.H. Orkin. 1986. Cloning the gene for an inherited human disorder-chronic granulomatous disease-on the basis of its chromosomal location. *Nature* 322: 32.



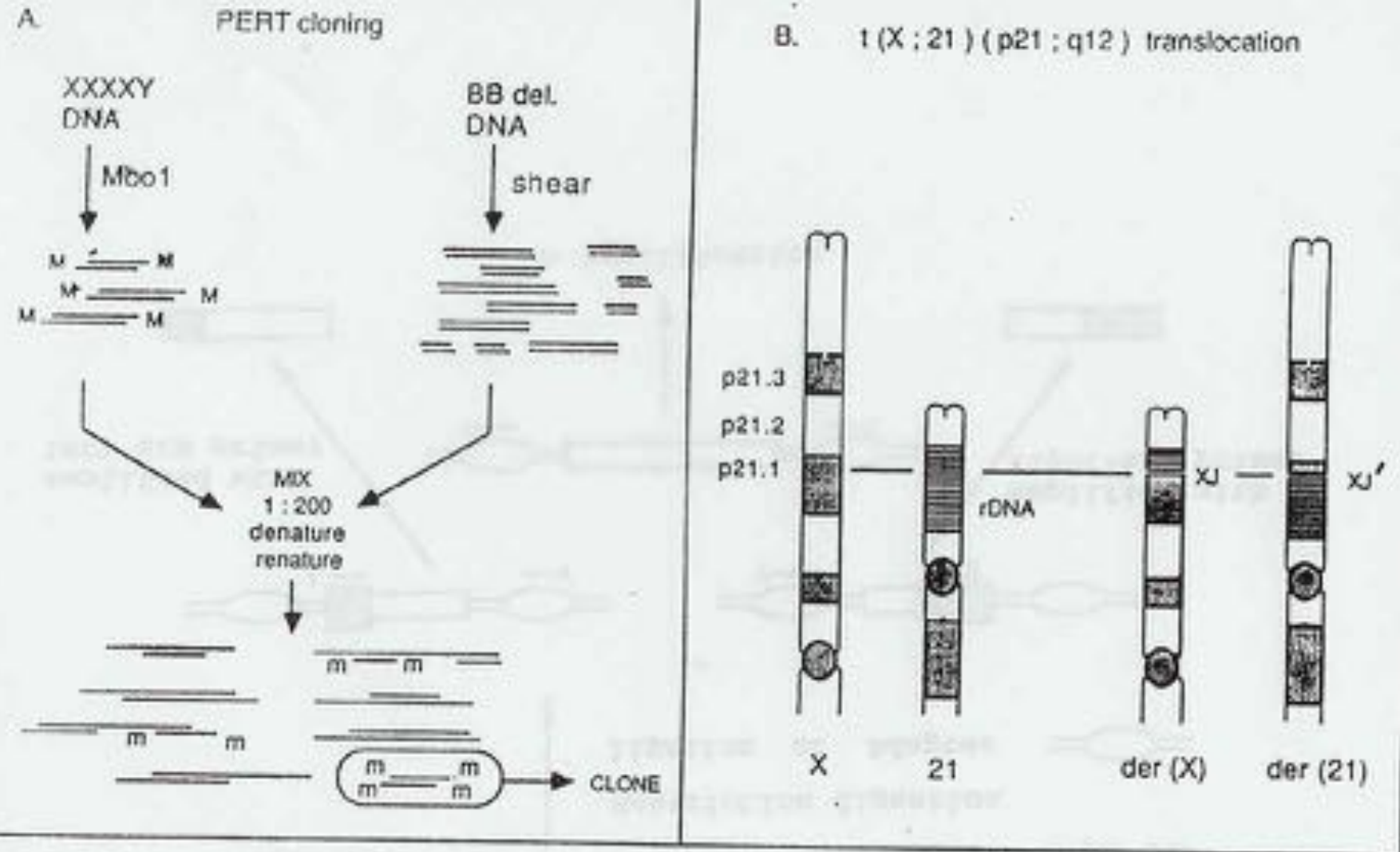
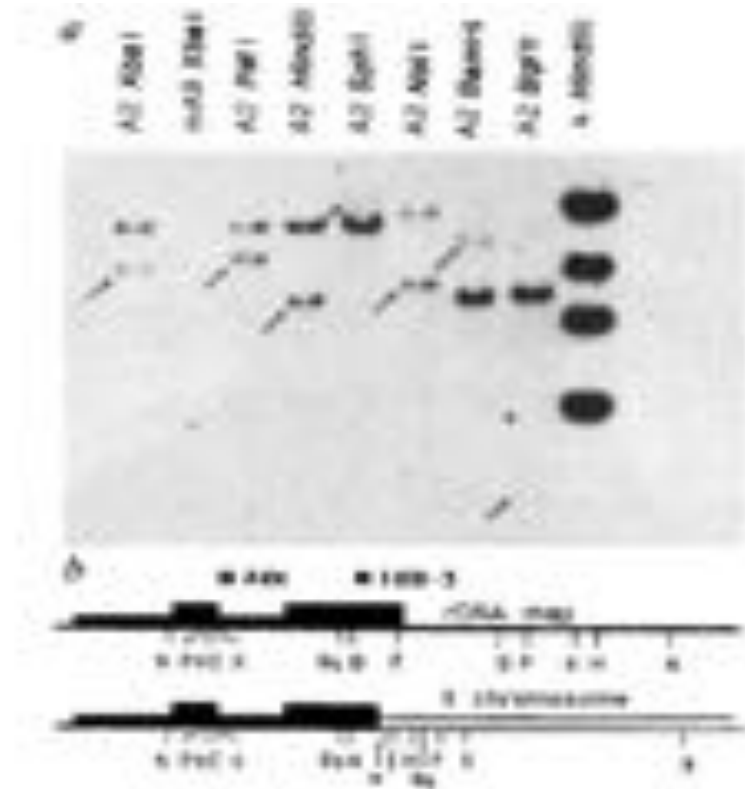
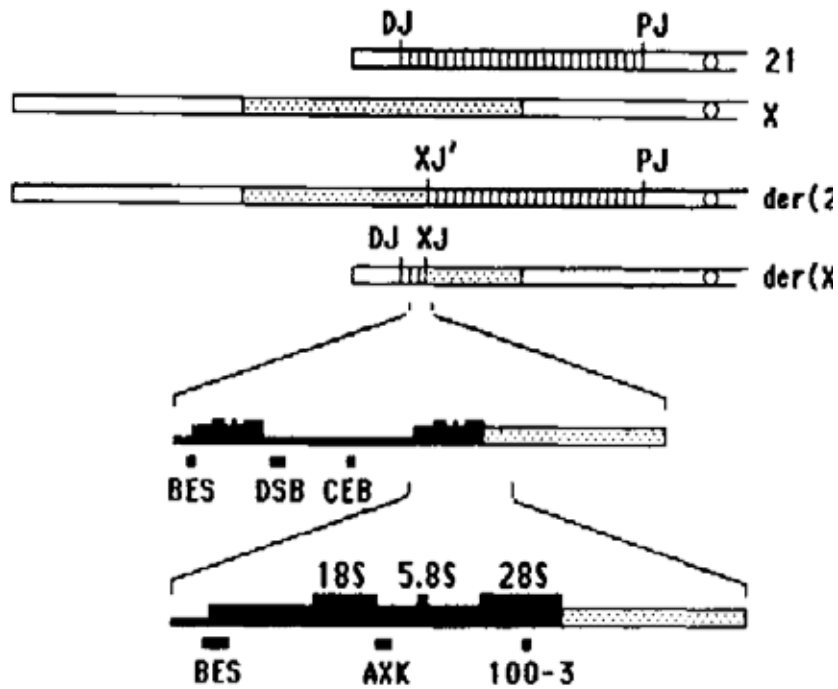


Figure 2 Schematic to illustrate the two successful cloning strategies.

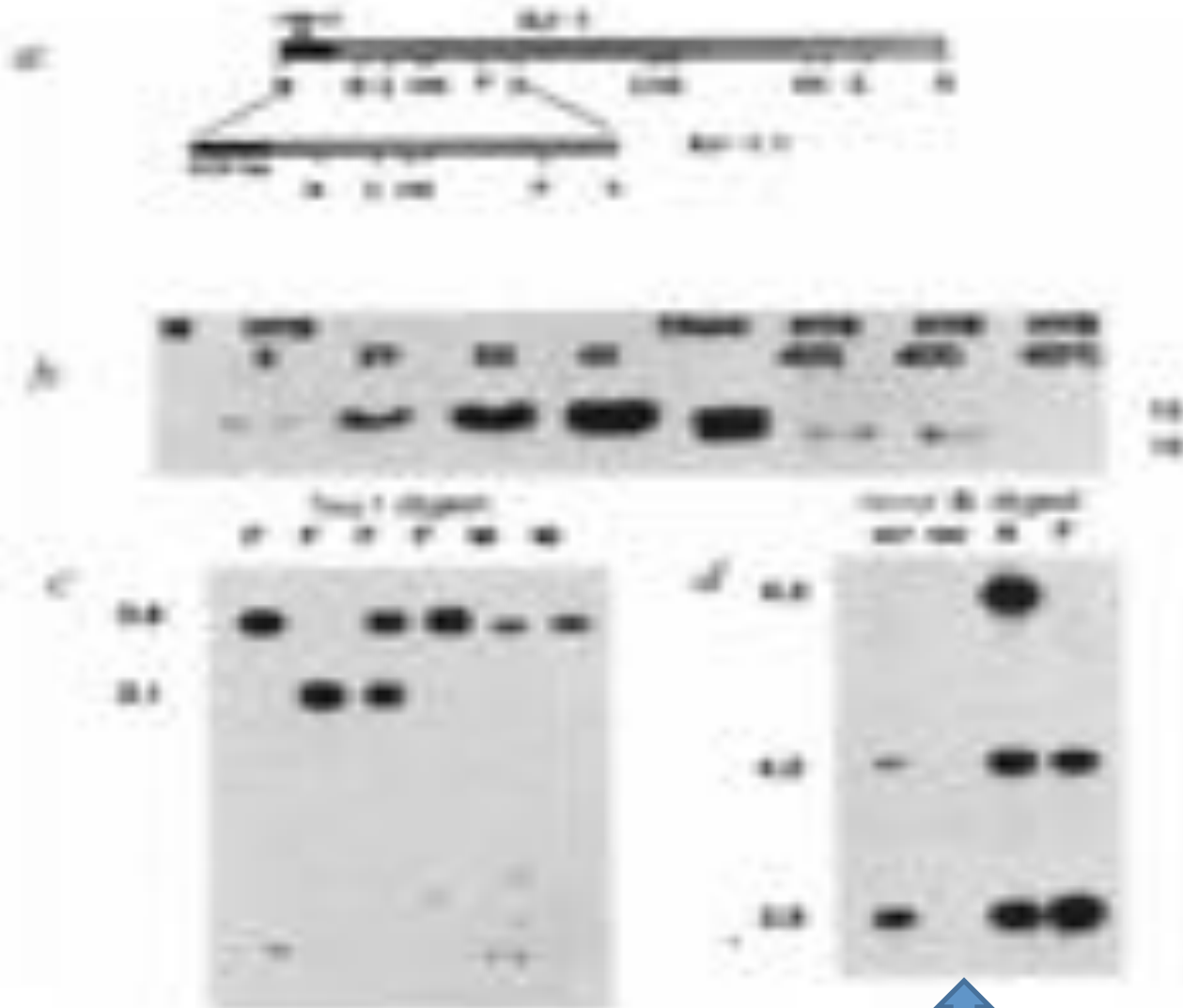
A. The pERT cloning scheme is described in the text. The ends of the *MboI*-digested molecules are indicated as M (double stranded) or m (single stranded). The ends of the sheared molecules are unlabeled. Among the reassociated molecules, only the perfectly aligned *MboI*-digested molecules are clonable, and these are enriched for sequences missing from the DNA of patient BB.

Translocation Junction Cloned

Ray et al.



Probe XJ 1.1 Detects deletions in patients with just DMD



DMD with deletion

Isolation of candidate cDNAs for portions of the Duchenne muscular dystrophy gene

ANTHONY P. MONACO*†, RACHAEL L. NEVE*†, CHRIS COLLETTI-FEENER*,
CORLEE J. BERTELSON*, DAVID M. KURNIT* & LOUIS M. KUNKEL*†‡

*Division of Genetics, Mental Retardation Program, Department of Pediatrics,
Harvard Medical School, The Children's Hospital, Boston, Massachusetts 02115,
USA

†The Program in Neuroscience, Harvard University, Cambridge, Massachusetts
02138, USA

‡To whom correspondence should be addressed.

Nature. 1986 Oct 16-22;323(6089):646-50

A cDNA clone from the Duchenne/Becker muscular dystrophy gene

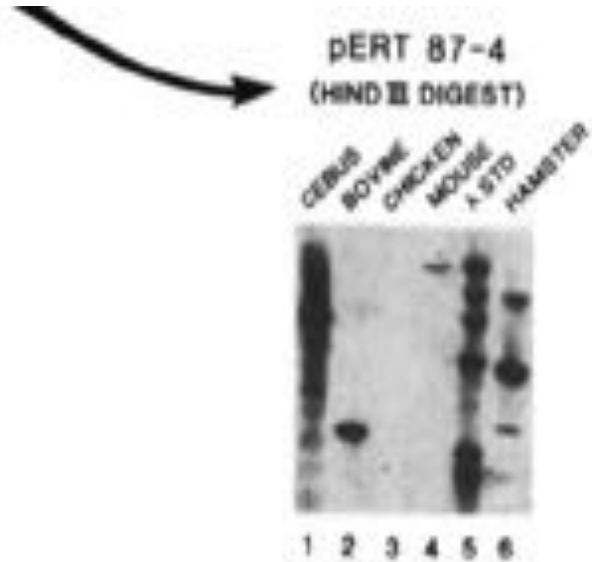
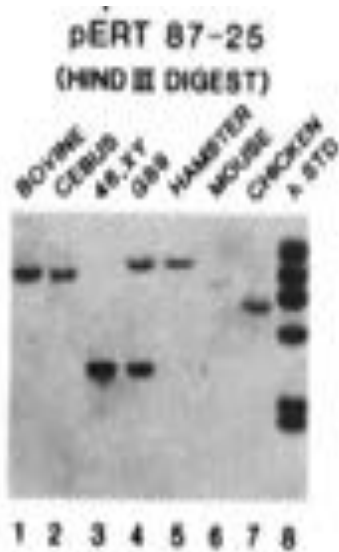
ARTHUR H. M. BURGHEES, CAIRINE LOGAN, XIUYUAN HU, BONNIE BELFALL,
RONALD G. WORTON & PETER N. RAY

Genetics Department and Research Institute, The Hospital for Sick Children and
The Departments of Medical Genetics and Biophysics, University of Toronto, 555
University Avenue, Toronto, Ontario M5G 1X8, Canada

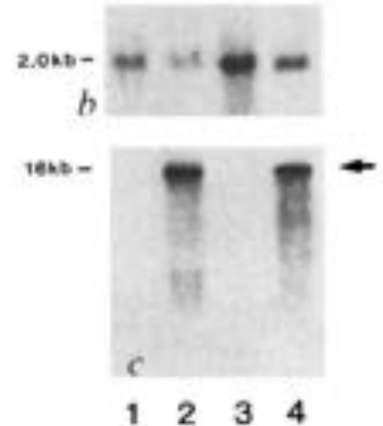
Nature. 1987 Jul 30-Aug 5;328(6129):434-7

Identification of DMD mRNA Monaco et al

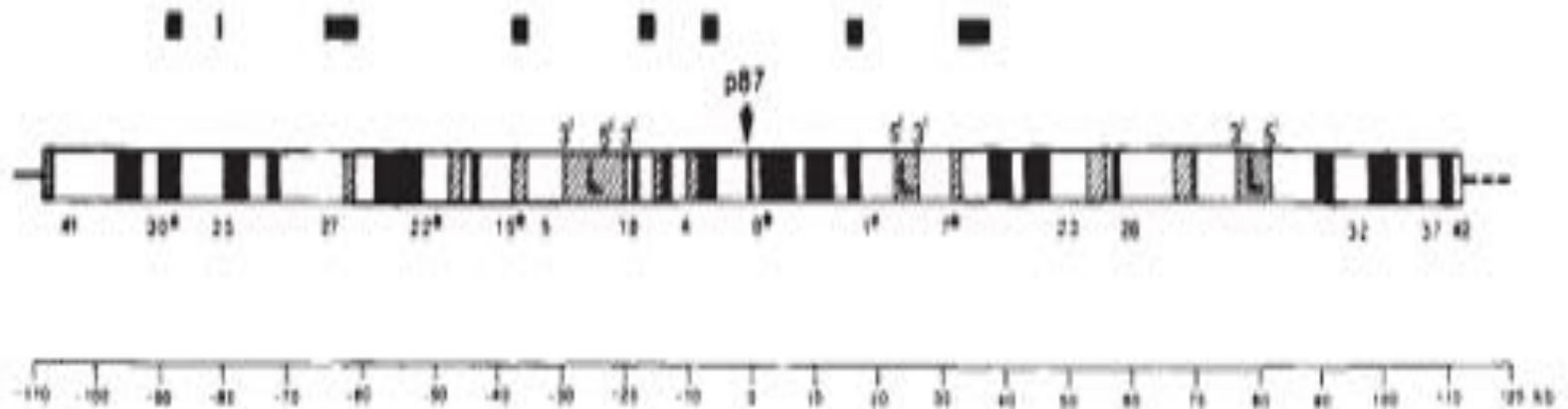
Identification of conserved segments in PERT 87 region
pERT87-25 detected mRNA and cDNA



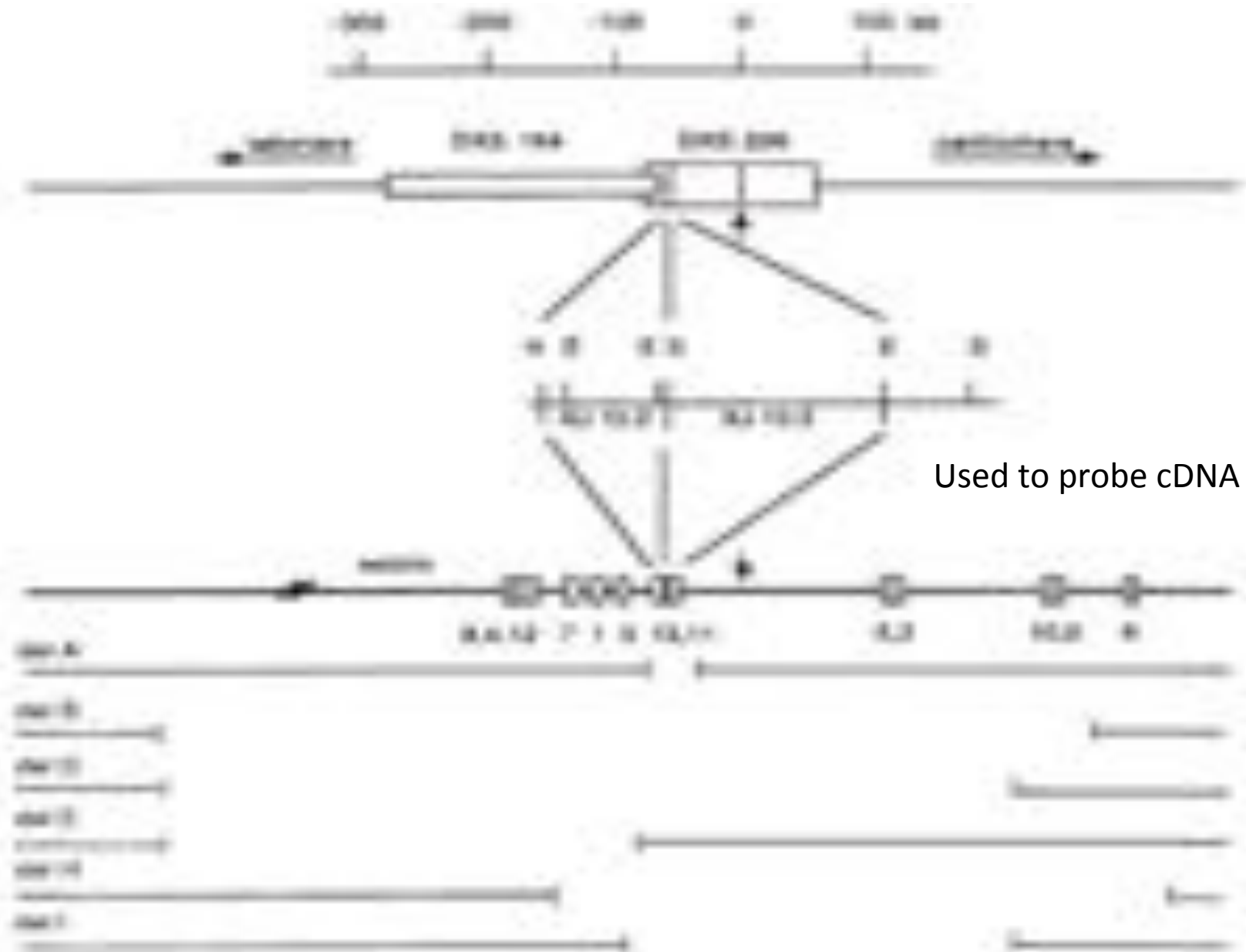
14kb dystrophin
mRNA



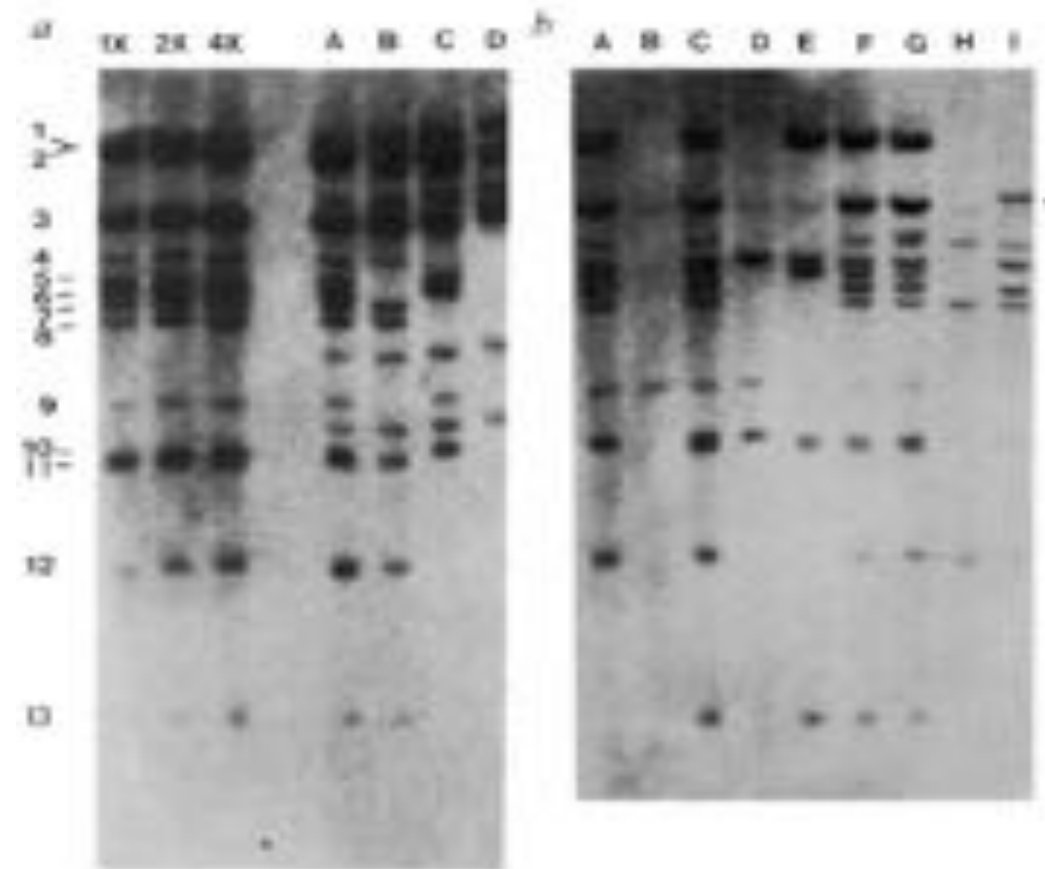
Distribution of dystrophin exons in pERT region



Identification of cDNA/mRNA from DX206 and 110kb intron of dystrophin



Identification of multiple exons and deletions at the five prime end of dystrophin

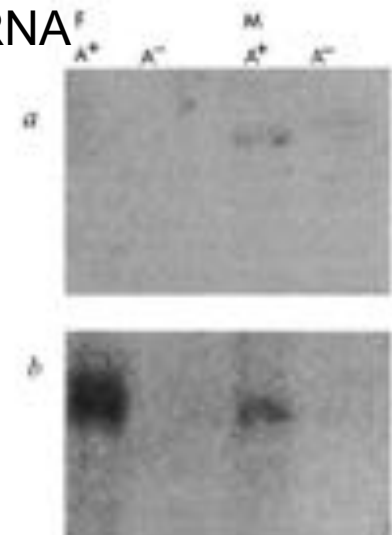


Southern blot of patient DNA probed with 2.0kb Dystrophin cDNA

Table 1 Exon mapping using translocation hybrids and deletion patients

Exon*	Fragment size (kb)	Translocation chromosome		Deletion patients									
		Der(X)	Der(21)	A	B	C	D	E	F	G	H	I	
E8	5.3		+	+	+				+	+	+	+	
E4	3.7		+	+	+				+	+	+	+	
E12	0.8		+	+	+				+	+	+	+	
E7	4.0		+	+	+				+	+		+	
E1	15		+	+	+				+	+		+	
E3*(+)	6.6		+	+	+				+	+			
E13†	0.7		+			+		+	+	+			
E11†	1.8		+			+		+	+	+			
(X;21)													
E6	4.3	+		+		+		+	+	+			
E2	13.5	+		+		+		+	+	+			
E10	2.1	+		+		+	+	+	+	+		+	
E5	4.6	+		+		+	+	+	+	+		+	
E9	2.6	+		+	+	+	+	+	+	+		+	

Northern detecting 14kb Dystrophin mRNA



Subcellular fractionation of dystrophin to the triads of skeletal muscle

ERIC P. HOFFMAN*, C. MICHAEL KNUDSON‡, KEVIN P. CAMPBELL‡ & LOUIS
M. KUNKEL*†

* Department of Pediatrics, Harvard Medical School and Division of Genetics, Children's
Hospital, and

† Howard Hughes Medical Institute, Boston, Massachusetts 02115, USA

‡ Department of Physiology and Biophysics, University of Iowa, Iowa City, Iowa 52242,
USA

**Here we show that dystrophin is associated with the triadic junctions in skeletal muscle,
and is therefore probably involved with Ca^{2+} homoeostasis.**

Nature. 1987 Dec 24-31;330(6150):754-8

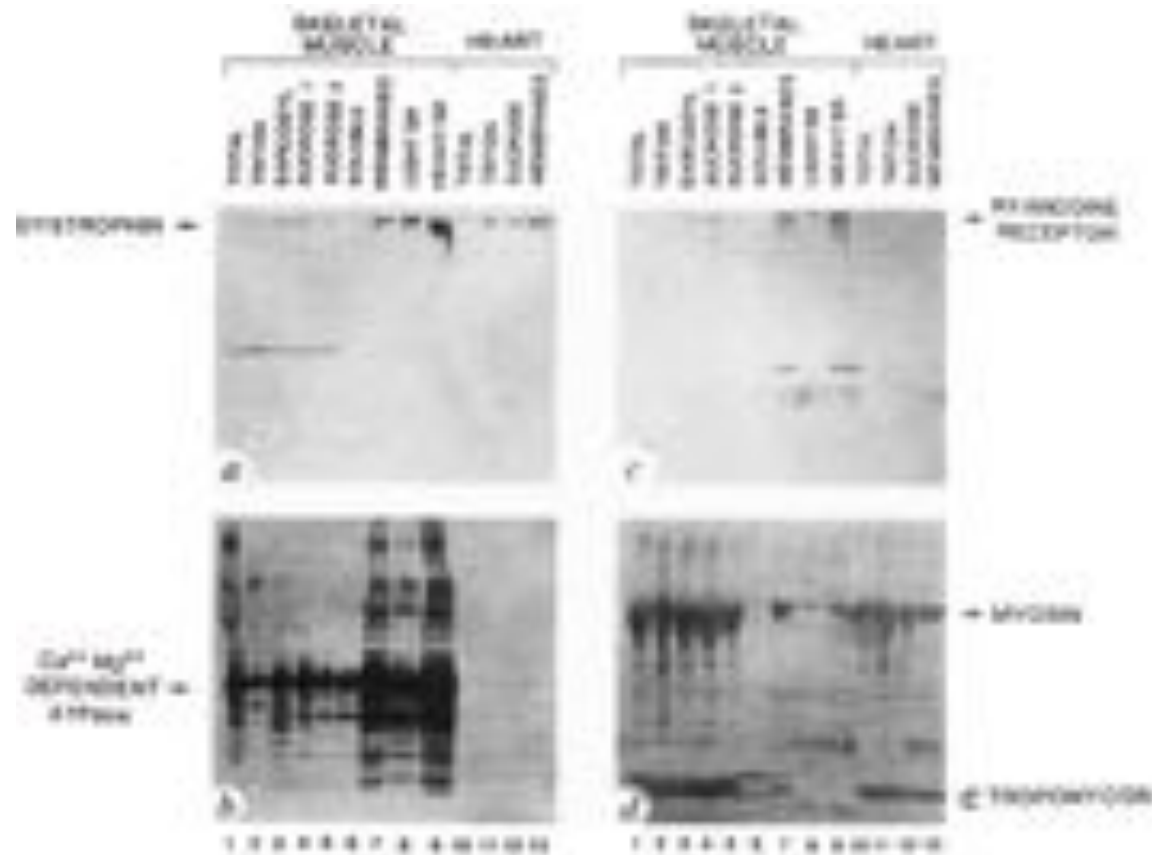
Nature 330 1987

News and views Nature

Muscular dystrophy

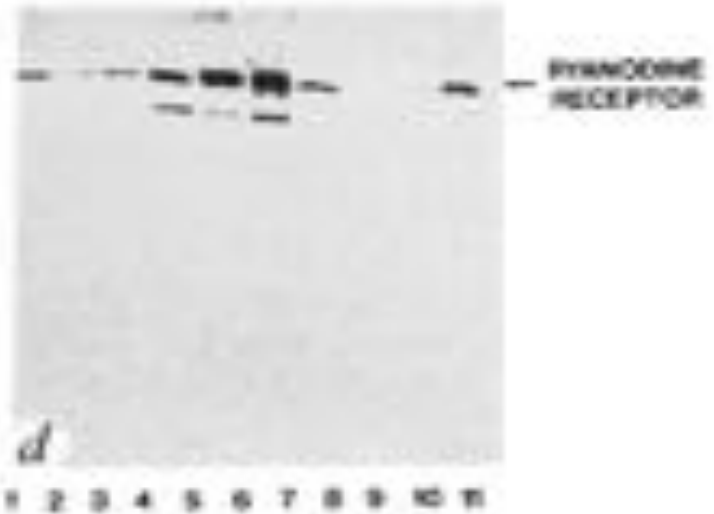
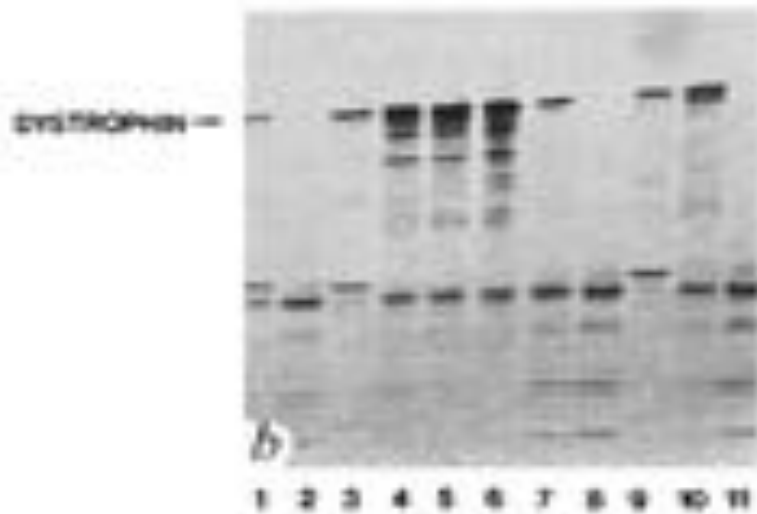
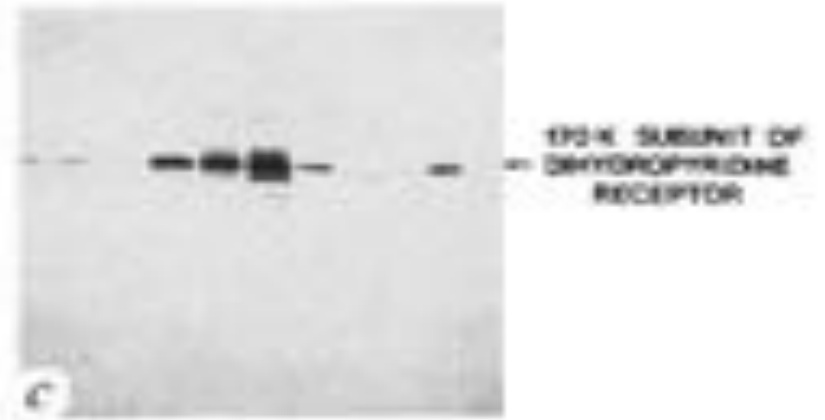
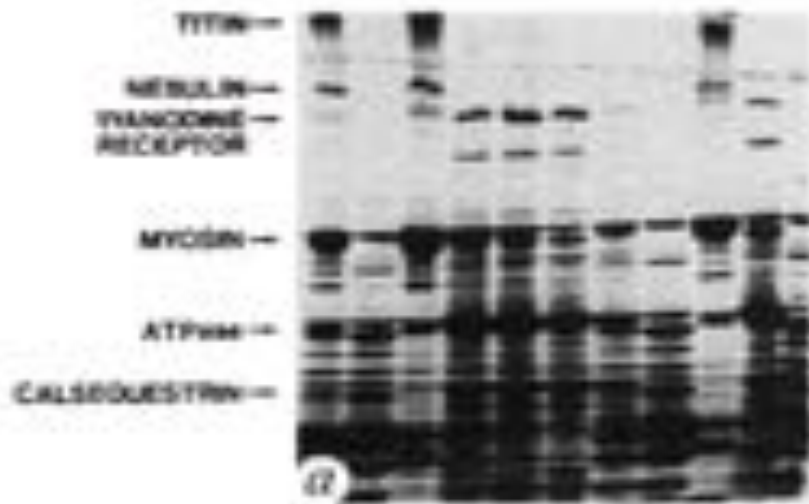
The missing link in DMD?

Clarke R. Slater



This protein species is called dystrophin because of its identification by molecular-genetic analysis of affected individuals. Here we show that dystrophin is associated with the triadic junctions in skeletal muscle, and is therefore probably involved with Ca^{2+} homoeostasis.

400kd protein not Nebulin but dystrophin



But no plasma membrane marker such as muscle Sodium Channel

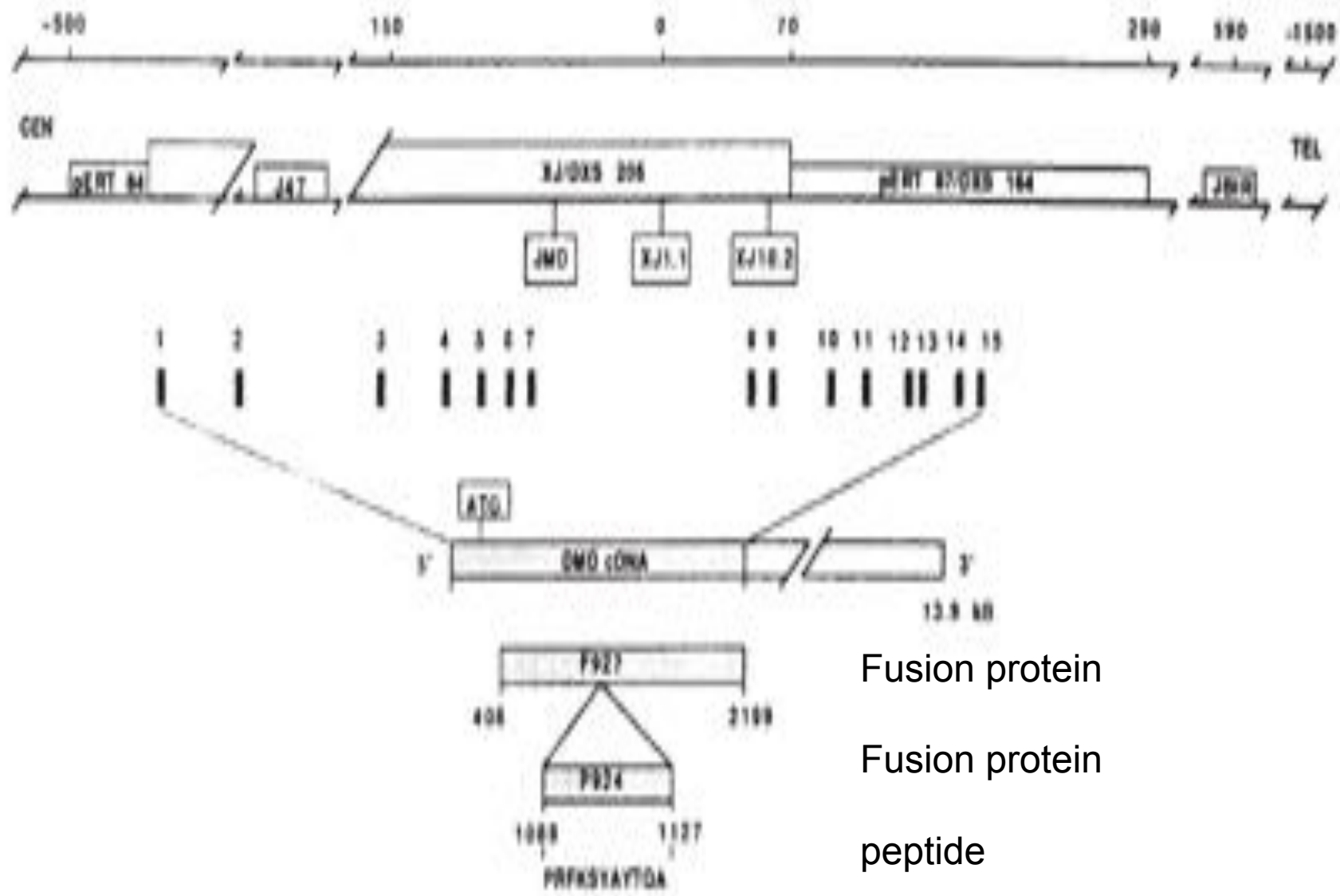
The Duchenne muscular dystrophy gene product is localized in sarcolemma of human skeletal muscle

**Elizabeth E. Zubrzycka-Gaarn*, Dennis E. Bulman*,
George Karpati†, Arthur H. M. Burghes*,
Bonnie Belfall*, Henry J. Klamut*, Jim Talbot‡,
Robert S. Hodges‡, Peter N. Ray*
& Ronald G. Worton***

* Genetics Department and Research Institute, The Hospital for Sick Children and The Departments of Medical Genetics and Medical Biophysics, University of Toronto, 555 University Avenue, Toronto, Ontario M5G 1X8, Canada

† Neuromuscular Research Group, Montreal Neurological Institute, McGill University, Montreal, Quebec H3A 2B4, Canada

‡ Department of Biochemistry, University of Alberta, Edmonton, Alberta T6G 2H7, Canada

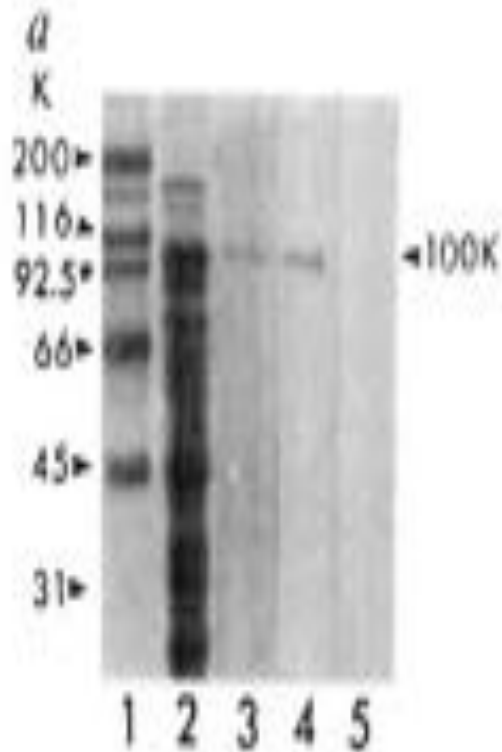


Fusion protein

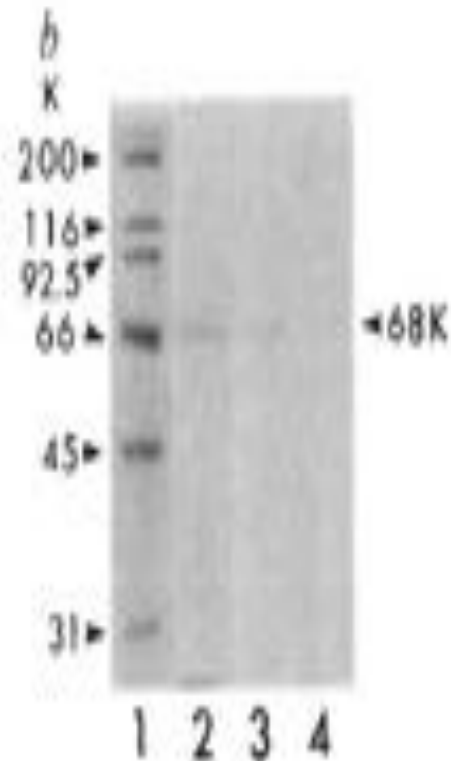
Fusion protein

peptide

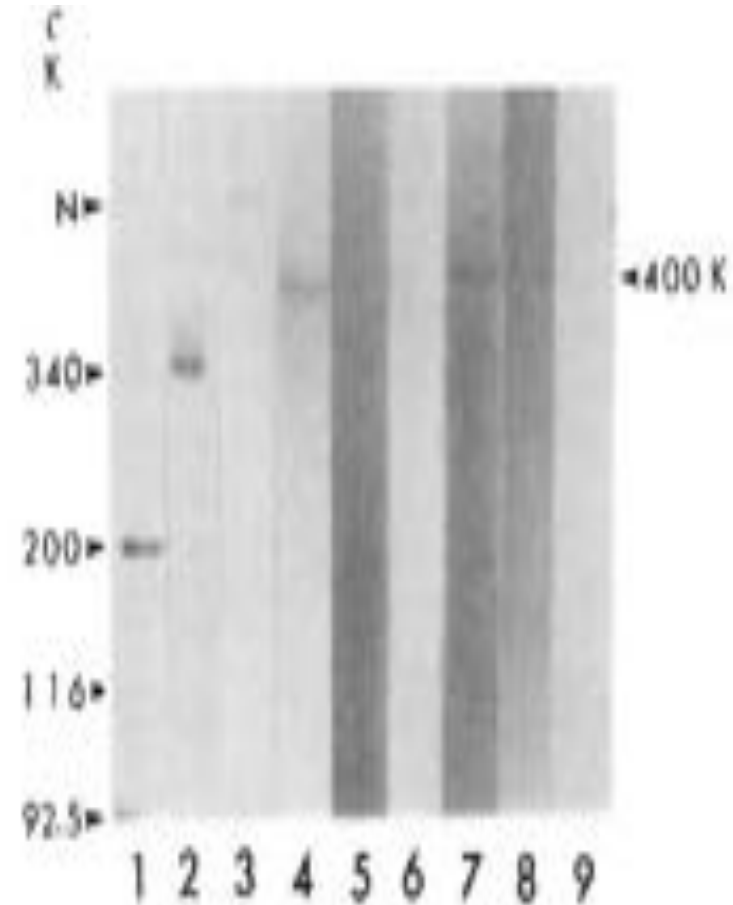
Antibodies do recognize dystrophin



E.Coli Fusion protein
of DMD cDNA
In pATH and PRIT vector
3)Peptide P924
4)Fusion F927
5) pre-immune

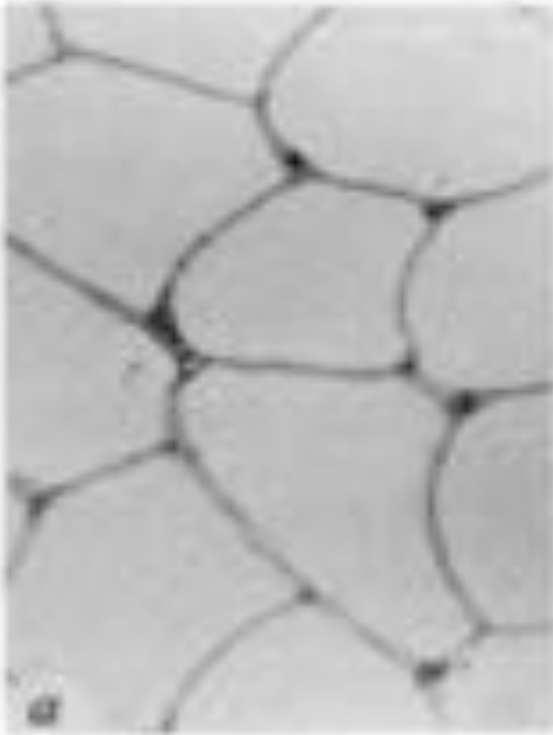


In-vitro translation
of DMD cDNA



Muscle fractionated
(Not a whole leg)
4)P924
7)F927

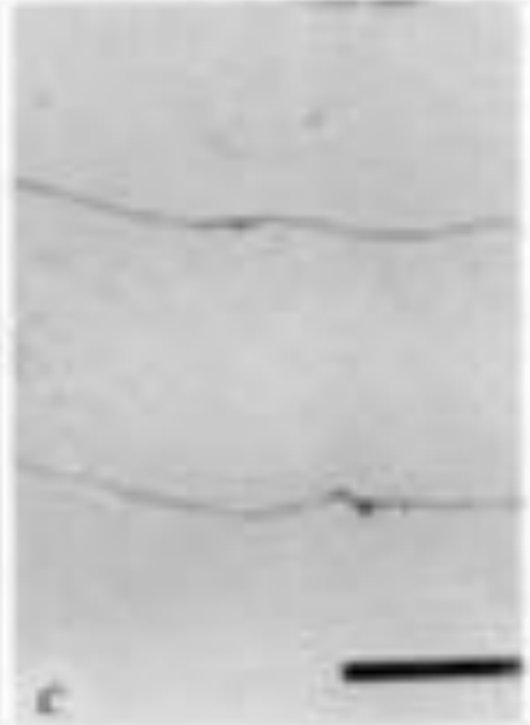
Dystrophin is at the membrane



Normal muscle stained
with F927

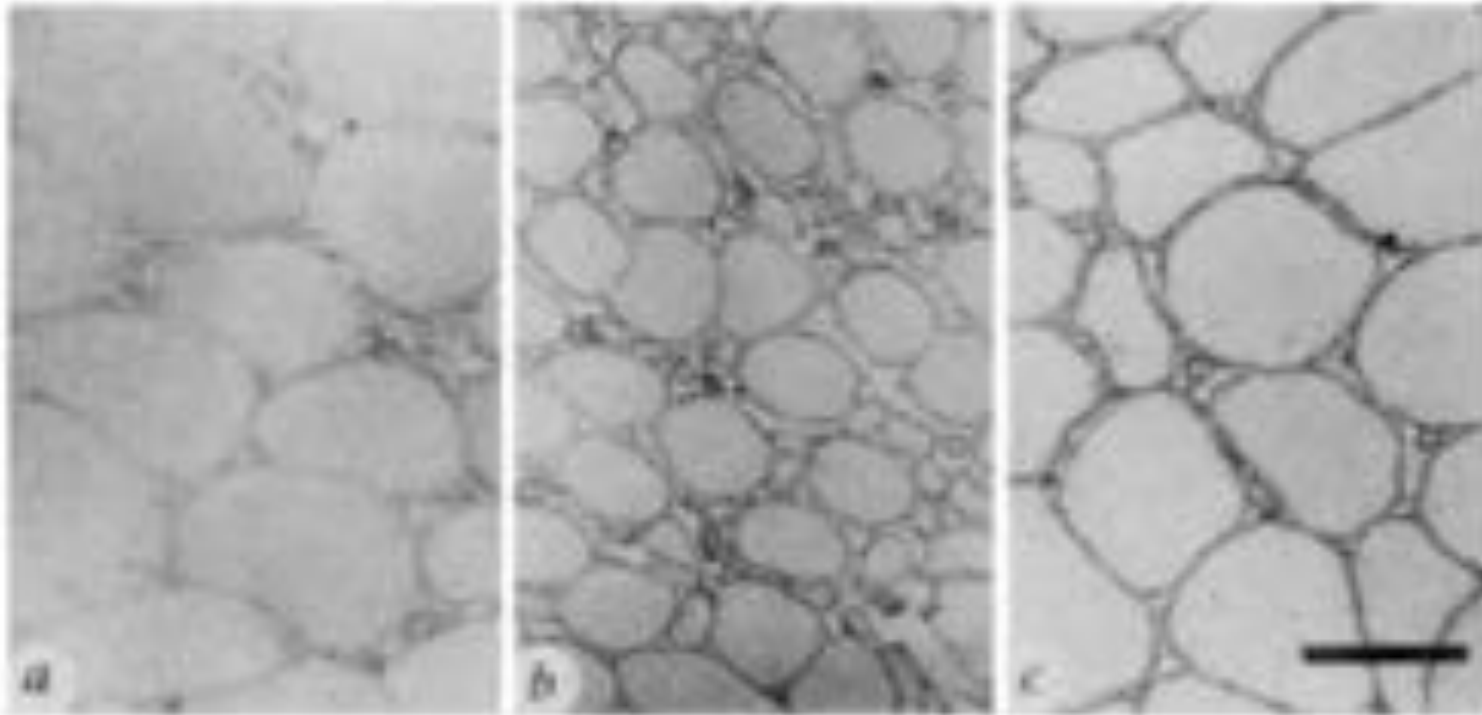


Normal muscle stained
With pre-immune serum



Normal muscle longitudinal
stained with F927

F927 immunostaining shows absence of dystrophin in DMD



5 year old DMD

Nemaline rod
myopathy

Dermatomyositis

Immunostaining of skeletal and cardiac muscle surface membrane with antibody against Duchenne muscular dystrophy peptide

Kiichi Arahata, Shoichi Ishiura, Tsuneo Ishiguro*, Toshifumi Tsukahara, Yoshihiro Sahara, Chikahiko Eguchi*, Tadayuki Ishihara†, Ikuya Nonaka, Eijiro Ozawa & Hideo Sugita

National Institute of Neuroscience, NCNP, 4-1-1 Ogawa-Higashi, Kodaira, Tokyo 187, Japan

* Central Research Laboratories, Ajinomoto Co., Inc., Kawasaki-ku, Kawasaki 210, Japan

† National Higashi-Saitama Hospital, Saitama 349-01, Japan

Immunolectron microscopic localization of dystrophin in myofibres

Simon C. Watkins, Eric P. Hoffman*, Henry S. Slayter & Louis M. Kunkel*†

Structural Molecular Biology, Dana-Farber Cancer Institute and Harvard Medical School, Boston, Massachusetts 02115, USA

* Division of Genetics, Children's Hospital, and Harvard Medical School, Boston, Massachusetts 02115, USA

† Howard Hughes Medical Institute, Children's Hospital, Boston, Massachusetts 02115, USA

Two historical references with some of the information

Duchenne Muscular Dystrophy

[Alan Emery](#) and [Francesco Muntoni](#)

**Genome: The Story of the Most Astonishing Scientific
Adventure of Our Time--The Attempt to Map All the Genes
in the Human Body**

By Jerry Bishop and Michael Waldholz