# Fibrosis in DMD

Federica Montanaro, Ph.D.

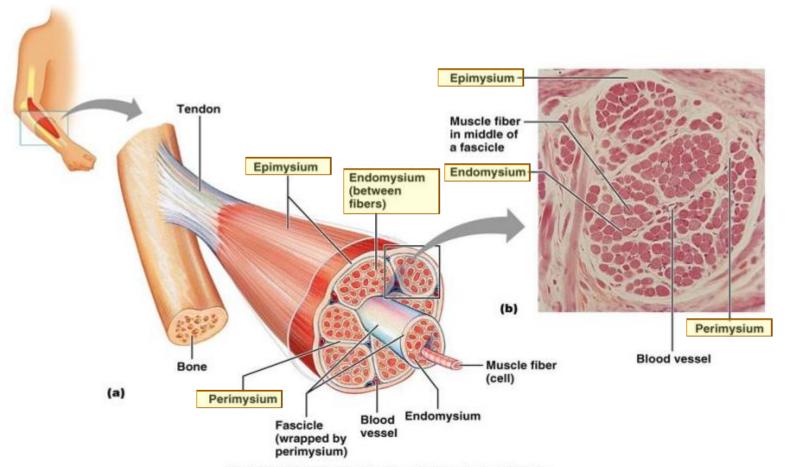
MVIMG# 7470: Fundamentals of Muscle Biology: Duchenne Muscular Dystrophy

April 3, 2014

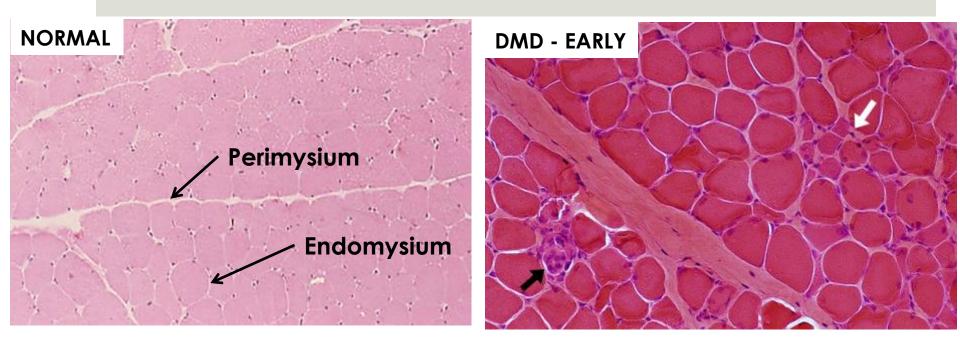
# What is fibrosis?

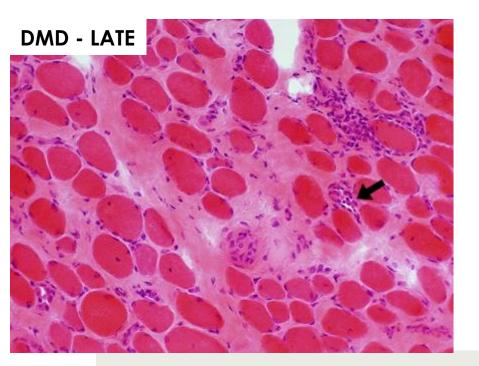
- Basic response of <u>any organ</u> that undergoes <u>repetitive</u> injury and inflammation.
- Characterized by the excessive deposition of extracellular matrix proteins (mainly collagens I and III, fibronectin) thus creating a scar.
- Leads to a disordered tissue structure, disruption of organ function, and ultimately organ failure.
- Major cause of mortality worldwide.
- No available FDA- or EMEA- approved anti-fibrotic therapies.

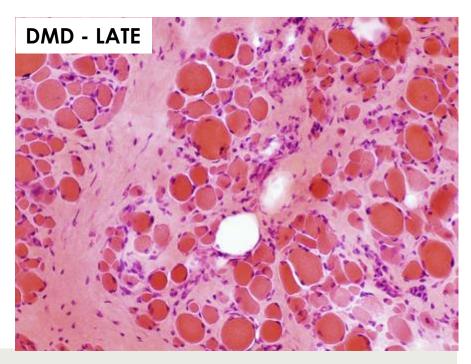
# Impact on disease progression in DMD



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J Neuropathol Exp Neurol Copyright © 2009 by the American Association of Neuropathologists, Inc. Vol. 68, No. 7 July 2009 pp. 762–773

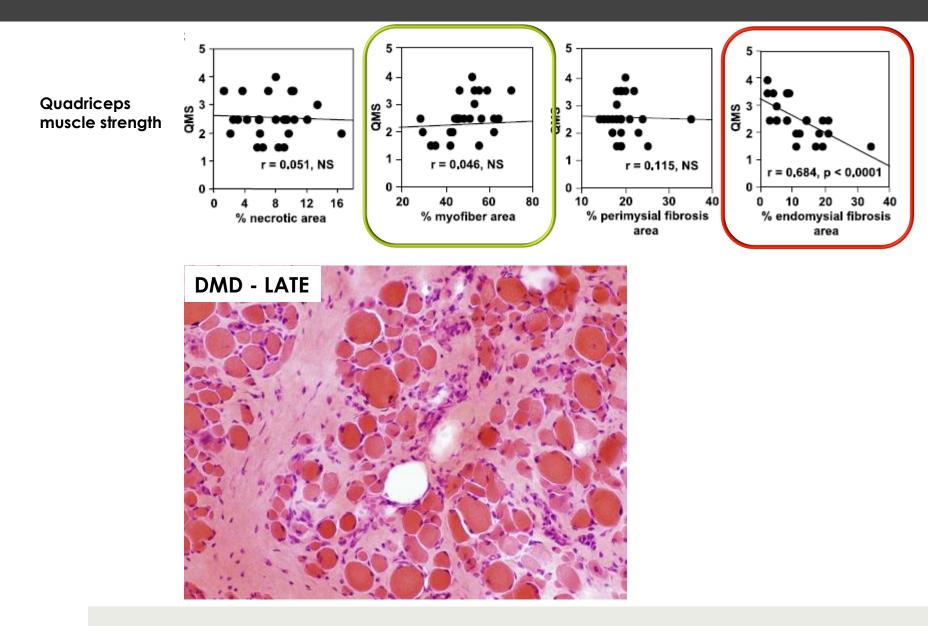
Original Article

### Endomysial Fibrosis in Duchenne Muscular Dystrophy: A Marker of Poor Outcome Associated With Macrophage Alternative Activation

Isabelle Desguerre, MD, Michelle Mayer, MD, France Leturcq, PhD, Jacques-Patrick Barbet, MD, PhD, Romain K. Gherardi, MD, and Christo Christov, MD

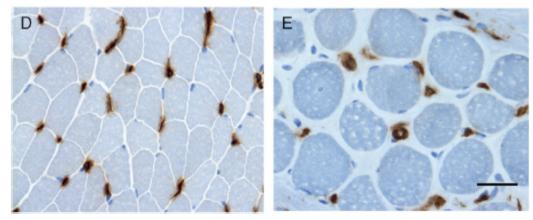
Endomysial fibrosis is the main histopathological parameter that correlates with poor motor outcome in DMD patients

### Consequences of Endomysial Fibrosis



### Consequences of Endomysial Fibrosis

Loss of tight association between muscle fibers and capillaries → decreased oxygenation and nutrients

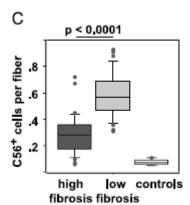


CD31 staining (brown) of capillaries

Control



Decreased number of satellite cells  $\rightarrow impaired regeneration$ 



### Consequences of Endomysial Fibrosis

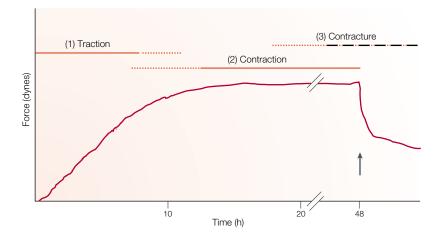
### Tissue contracture

Increased tissue stiffness

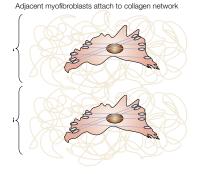
 $\rightarrow$  inhibits the proliferation and differentiation of satellite cells

 $\rightarrow$  Enhances production of matrix proteins by fibrotic cells

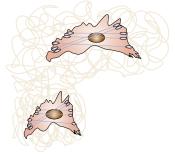
 $\rightarrow$  Interferes with muscle contraction



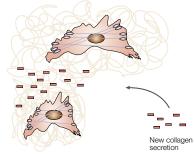
Tomasek et al., 2002, Nature Reviews 3: 349



Myofibroblast B contracts, deforming network B

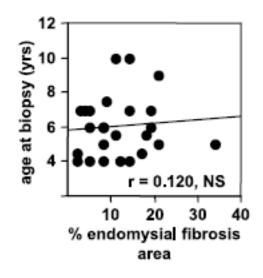


New collagen secretion stabilizes contracted structure of network  ${\bm B},$  relative to network  ${\bm A}$ 



Cell re-spreads and process is repeated

### Genetic modifiers of fibrosis



Variability in the rate of disease progression among patients, even if they have the same genetic mutation in the *DMD* gene or lack of dystrophin protein expression

# Latent TGF-β-binding protein 4 modifies muscular dystrophy in mice

Ahlke Heydemann,<sup>1</sup> Ermelinda Ceco,<sup>2</sup> Jackie E. Lim,<sup>3</sup> Michele Hadhazy,<sup>1</sup> Pearl Ryder,<sup>1</sup> Jennifer L. Moran,<sup>4</sup> David R. Beier,<sup>4</sup> Abraham A. Palmer,<sup>2</sup> and Elizabeth M. McNally<sup>1,2,3</sup>

<sup>1</sup>Department of Medicine, Section of Cardiology, <sup>2</sup>Committee on Cell Physiology, and <sup>3</sup>Department of Human Genetics, University of Chicago, Chicago, Illinois, USA. <sup>4</sup>Genetics Division, Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts, USA.

The Journal of Clinical Investigation http://www.jci.org Volume 119 Number 12 December 2009

### LTBP4 Genotype Predicts Age of Ambulatory Loss in Duchenne Muscular Dystrophy

Kevin M. Flanigan, MD,<sup>1,2,3</sup> Ermelinda Ceco, BS,<sup>4</sup> Kay-Marie Lamar, BS,<sup>4</sup>
Yuuki Kaminoh, BS,<sup>1</sup> Diane M. Dunn, BS,<sup>5</sup> Jerry R. Mendell, MD,<sup>1,2,3</sup>
Wendy M. King, PT,<sup>3</sup> Alan Pestronk, MD,<sup>6</sup> Julaine M. Florence, DPT,<sup>6</sup>
Katherine D. Mathews, MD,<sup>7</sup> Richard S. Finkel, MD,<sup>8</sup> Kathryn J. Swoboda, MD,<sup>9</sup>
Eduard Gappmaier, PhD,<sup>10</sup> Michael T. Howard, PhD,<sup>5</sup> John W. Day, MD, PhD,<sup>11</sup>
Craig McDonald, MD,<sup>12</sup> Elizabeth M. McNally, MD, PhD,<sup>4</sup> and Robert B. Weiss, PhD<sup>5</sup> for the United Dystrophinopathy Project

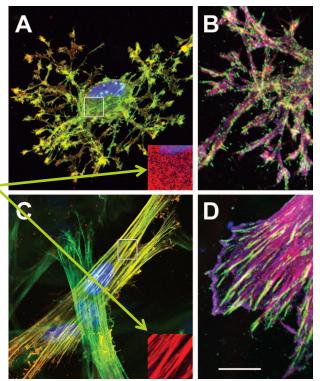
ANN NEUROL 2013;73:481-488

# The Fibroblast

### Versatile in shape

- Versatile in gene expression
- Versatile in function
  - Developingipha-Smooth muscle actin
    - Promote slow muscle myogenesis
    - Fetal to adult switch Myofibroblast /
    - Myoblast fusigetivated fibroblast
  - Adult muscle
    - Regulation of satellite cell selfrenewal and differentiation
    - Tissue integrity

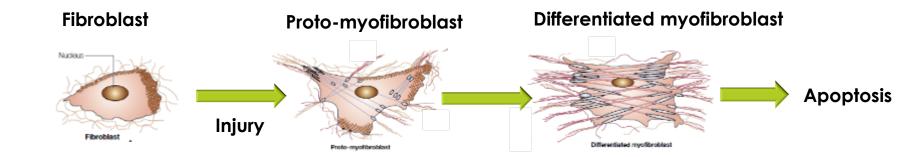
### Lung fibroblasts



Soft

Stiff

# Fibroblast activation



Phenotype Tissue homeostasis

- Migratory cell
- Matrix protein production, including specific forms of FN
- Production of TGF- $\beta$

- Contractile cell
- High matrix protein production
- Production of TGF-β
- Production of ROS

Myofibroblasts are the key pathogenic cells in all fibrotic diseases

Research has focused on identifying:

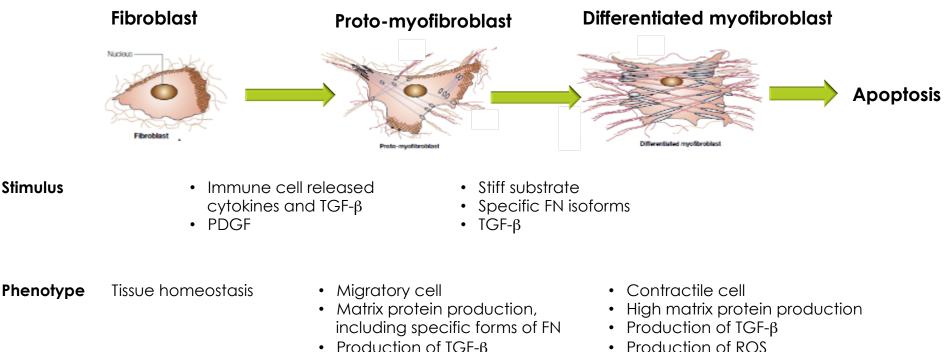
the factors that activate myofibroblasts

the mechanisms that contribute to myofibroblast apoptosis

the cellular origins of myofibroblasts

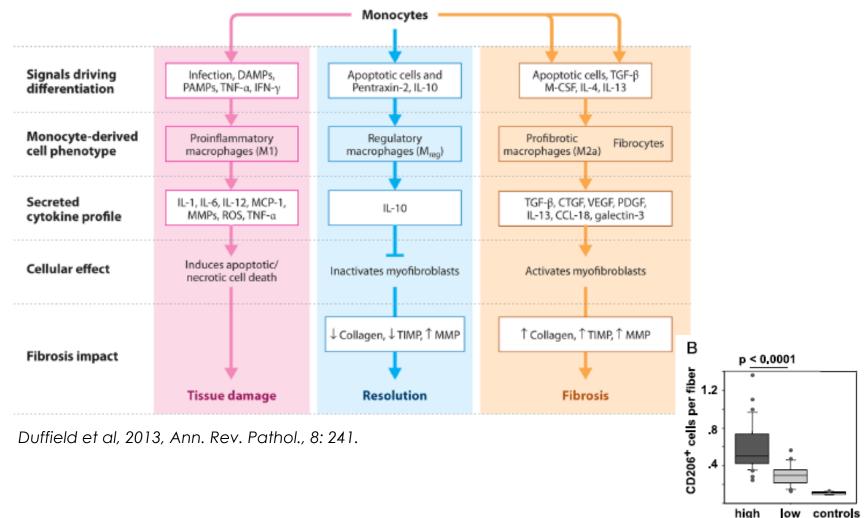
# Fibroblast activation

Stimulus



Production of TGF-β

# Fibroblast activation – impact of immune cells

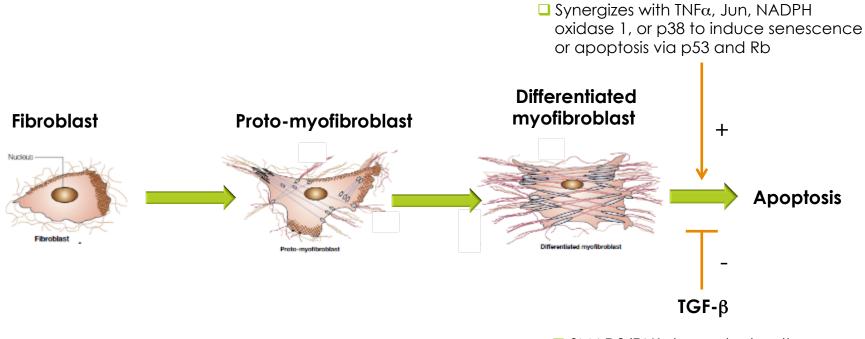


Desguerre et al, 2009, J. Neuropathol. Exp. Neurol., 68(7): 762.

fibrosis fibrosis

# Myofibroblast apoptosis

#### CCN1



SMAD3/FAK-dependent pathway

P38 MAPK/PI3 kinase/Akt dependent signaling

### Cellular origins of fibroblasts/ myofibroblasts

- Circulating Fibrocytes
- Endothelial to mesenchymal transition
- Epithelial to mesenchymal transition
- Mesenchymal progenitors/fibroblast and adipocyte precursors
- Pericytes

- Genetic fate mapping experiments in several organs, including skeletal muscle, brain, kidney, lung skin and liver indicate that mesenchymal progenitors and pericytes are the precursors of myofibroblasts.
- Many parallel genetic fate mapping studies show little or no evidence of direct differentiation of epithelial cells, endothelial cells, or circulating fibrocytes into myofibroblasts

### Mesenchymal progenitors

- Location: interstitium
- Main markers: PDGFR-α, Sca-1, CD34
- Differentiation potential:
  - Fibroblasts
  - Adipocytes
  - Osteogenic
  - Chondrogenic

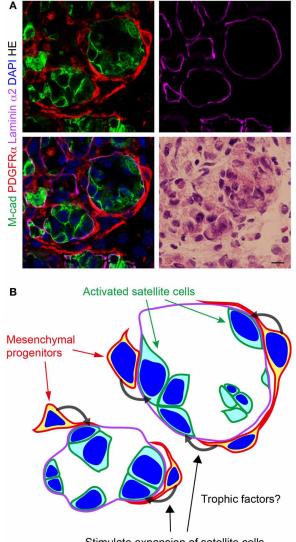
# Acute muscle injury

- Release trophic factors that support satellite cell expansion and myogenic differentiation
- Phagocytose dead cells and cellular debris

### Type 2 Innate Signals Stimulate Fibro/Adipogenic Progenitors to Facilitate Muscle Regeneration

Jose E. Heredia,<sup>1,10</sup> Lata Mukundan,<sup>1,10</sup> Francis M. Chen,<sup>1</sup> Alisa A. Mueller,<sup>6</sup> Rahul C. Deo,<sup>1,3,7,8</sup> Richard M. Locksley,<sup>3,4,5</sup> Thomas A. Rando,<sup>6,9</sup> and Ajay Chawla<sup>1,2,3,\*</sup>

Cell 153, 376-388, April 11, 2013



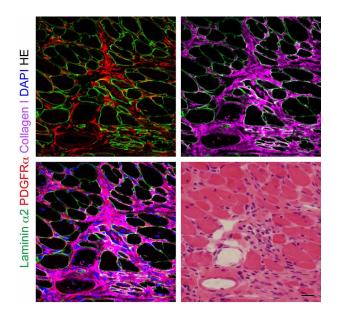
Stimulate expansion of satellite cells Promote satellite cell-dependent myogenesis

Uezumi et al., 2014, Frontiers in Physiology, Vol 5, Article 68, p2

# Muscular Dystrophy

### Mesenchymal progenitors:

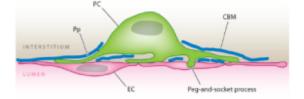
- Produce collagens
- Differentiate into fibroblasts and adipocytes



Uezumi et al., 2014, Frontiers in Physiology, Vol 5, Article 68, p2

### Pericytes

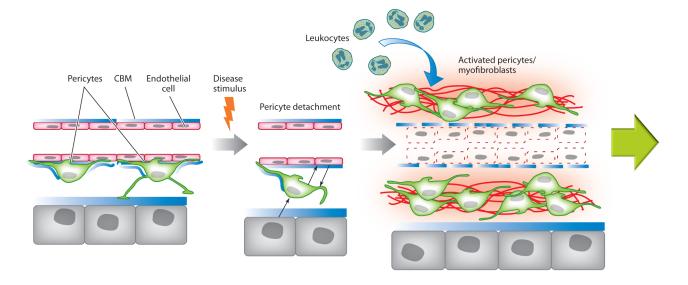
- Location: perivascular, around capillaries
- **D** Main markers: PDGFR- $\beta$ , NG2
- $\square$  Activated by PDGF, VEGF, TGF- $\beta$



- Main function: Microvasculature homeostasis
- Differentiation potential:
  - Myogenic
  - Adipogenic
  - Osteogenic
  - Fibrogenic

(Duloroy et al., 2012, Nat. Med., 18:1262)

### Pericyte activation



Fibrosis

Compromised microvascular homeostasis

Duffield et al., 2013, Annu. Rev. Pathol., 8: 241

# Summary

- Fibrosis is a major determinant of disease progression in DMD
- Replaces muscle tissue and impairs the function of residual muscle fibers
  - Inhibition of satellite cell proliferation
  - Impaired interactions with the microvasculature
  - Stiffens the matrix
- Tight relationship between fibrotic and immune cells
- Treatment targets:
  - Immune modulation
  - Inhibition of differentiation of fibroblast progenitors
  - Inhibition of fibroblast differentiation into myofibroblasts
  - Induction of apoptosis/senescence of myofibroblasts

### Anti-fibrotic treatment targets in DMD

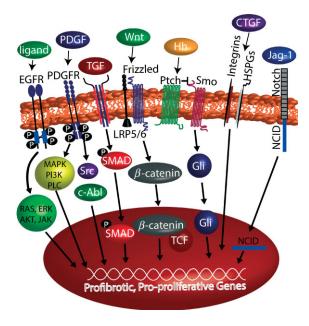
### Inflammation

- Nfk-B inhibition (Flavocoxid [Phase 1], VBP15 [preclinical])
- TNF-α inhibition (BKT-104, cV1q, LMP420, etanercept [preclinical])

### Pro-fibrotic pathways

- TGF-β (ACE inhibitors, Myostatin inhibitors [MYO-029, ACE-031, Follistatin]
- ROS (CoQ10 [Phase 2/3], Sunphenon Epigallocatechin-Gallate [Phase 2/3], Catena)
- Pro-regenerative pathways
  - □ IGF-1 [Phase 2]
  - □ Tissue vascularization (Tadalafil, Sildenafil, PDE inhibitors)

### Anti-fibrotic treatments are a challenge



#### Kramann et al, 2013, Journal of Pathology, 231:273

 Table 1. Therapeutics that are currently being tested or have been tested in fibrotic diseases (this list does not claim to be exhaustive)

Drug name	Company	Target/MOA	Indication	Phase/notes	Clinical Trials.gov identifier
Pirfenidone	Intermune	p38/TGFβ inhibitor	IPF	Approved in Europe and Asia, phase III in USA (ongoing)	NCT01366209
Fresolimumab	Sanofi	Anti-TGFβ monoclonal antibody	Diffuse systemic sclerosis	Phase I (recruiting)	NCT01284322
		,	FSGS IPF	Phase II (recruiting) Phase 1 (completed)	NCT01665391 NCT00125385
LY2382770	Lilly	Anti-TGFß monoclonal antibody	Diabetic kidney disease; diabetic nephropathy, diabetic glomerulosclerosis	Phase II (recruiting)	NCT01113801
STX-100	Biogen Idec	Anti-a <sub>v</sub> b <sub>6</sub> monoclonal antibody	IPF	Phase II (recruiting)	NCT01371305
Macitentan	Actelion	Endothelin receptor antagonist ET-A and ET-B	IPF	Phase II (fail)	NCT00903331
Bosentan	Actelion	Endothelin receptor antagonist, ET-A and ET-B	IPF	Phase III (fail)	NCT00631475
			Digital ulcers in SSc patients	Approved in EU	NCT00077584 NCT00319696
			Interstital lung disease with SSc	Phase II/III (did not improve outcomes versus natural course)	NCT00319033
Ambrisentan	Gilead	Endothelin receptor antagonist selective for ET-A	IPF	Phase III (fail)	NCT00879229
RE-021	Retrophin	Selective endothelin type A receptor antagonist	FSGS	Phase II (not yet open)	NCT01613118
FG-3019	Fibrogen	Anti-CTGF	Liver fibrosis due to HBV IPF	Phase II (ongoing) Phase II (ongoing, with promising preliminary results)	NCT01217632 NCT01262001
			Adolescents and adults with FSGS	Phase I (terminated)	NCT00782561
			Diabetic nephropathy Locally advanced or metastatic pancreatic cancer	Phase II (terminated) Phase I (ongoing)	NCT00913393 NCT01181245
PF-06473871 RXI-109	Pfizer RXi Pharmaceuticals	Antisense CTGF CTGF RNAi	Hypertrophic skin scarring Dermal scar prevention	Phase II (recruiting) Phase I (ongoing) Phase I (recruiting)	NCT01730339 NCT01640912 NCT01780077
SAR156597 Tralokinumab	Sanofi MedImmune	Bi-specific IL-4/IL-13 mAB IL-13 inhibition	IPF IPF	Phase I/II (recruiting) Phase II (recruiting)	NCT01529853 NCT01629667
QAX576	Novartis	IL-13 inhibition	Pulmonary fibrosis secondary to SSc	Phase II -Terminated due to SAE	NCT00581997
			IPF	Phase II (terminated)	NCT01266135
Rilonacept	Regeneron	IL-1 trap	SSc	Phase I/II (recruiting)	NCT01538719
CNTO 888	Centocor	MCP-1(CCL2) inhibition	IPF	Phase II (completed)	NCT00786201
Etanercept	Pfizer/Amgen	TNF inhibition	IPF	Phase II (fail)	NCT00063869
Actimmune Interferon-α lozenge	Intermune Amarillo	Human interferon-γ Oral IFNα	IPF IPF	Phase III (fail) Phase II (completed) Phase	NCT00075998 NCT01442779
PRM-151	Biosciences Promedior	Recombinant pentraxin-2	IPF	II (terminated) Phase I (completed, improvements in FVC and 6MWT)	NCT00690885 NCT01254409
Belimumab	GlaxoSmithKline	Anti-BAFF mAB	Scarring in trabeculectomy Membranous	Phase II (completed) Phase II (recruiting)	NCT01064817 NCT01610492
Pomalidomide	Celgene	Multiple; anti angiogenic and immunomodulatory	glomerulnephritis IPF	Phase II (not yet recruiting)	NCT01135199
			SSc	Phase II (recruiting)	NCT01559129
IW001	United Therapeutics	Collagen V solution as immunomodulator	IPF	Phase I (completed)	NCT01199887